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► **To cite this version:**

Francisco Cejudo, Andreas Meyer, Jean-Philippe Reichheld, Nicolas Rouhier, Jose Traverso. Thiol-based redox homeostasis and signaling. *Frontiers in Plant Science*, Frontiers, 2014, 5, pp.450 - 450. <10.3389/fpls.2014.00266>. <hal-01579721>

HAL Id: hal-01579721

<https://hal.univ-lorraine.fr/hal-01579721>

Submitted on 31 Aug 2017

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Thiol-based redox homeostasis and signaling

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Reviewed by:

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Keywords: thioredoxin, glutaredoxin, glutathione, redox regulation, redox signaling, plants

Plants imperatively have to cope with adverse conditions owing to their lack of mobility and to the high amounts of reactive oxygen species (ROS) generated from both respiration and photosynthetic metabolism. Although thiol redox homeostasis in plants is mainly preserved by the cellular glutathione pool, specific strategies have been adopted by the plant kingdom during evolution to manage these “extra” pro-oxidative conditions. Unlike human or yeast, plants generally possess a higher number of genes coding for antioxidant proteins, including protein families responsible of dithiol/disulfide exchange reactions. During the last decades, redox-dependent post-translational modifications of proteins proved to be pivotal to many cellular functions. In particular, this is critically important under some situations of environmental constraints taking into account the alterations and fine adjustment of the cellular redox status occurring during and after any biotic or abiotic stresses.

Indeed, thiol groups of cysteinyl residues are highly sensitive to oxidation which might critically perturb cellular homeostasis. Members of the thioredoxin superfamily are key proteins involved in the regulation of cysteine/protein redox state. They share two common and well-known features: (i) the presence of an active center containing at least one catalytic cysteine residue, and (ii) a highly conserved 3D-structure, the so-called thioredoxin fold, which consists of a four-stranded anti-parallel β -sheet surrounded by three α -helices. Key members of this super family are thioredoxins (TRX) and glutaredoxins (GRX). Representatives of both subgroups are distributed in most cellular compartments and contain at least one TRX motif in their structures. While TRXs are generally reduced by thioredoxin reductases (TR), the reduction of GRXs depends on reduced glutathione (GSH).

The 19 reports of this Research Topic provide timely overviews and new insights into redox regulation, focusing on both TR/TRX and GSH/GRX reduction systems in plants. The biochemical characteristics of these systems as well as their target proteins and functions in metabolic and signaling pathways are discussed. Several contributions to this Research Topic deal with the role

of TRX systems in plastid metabolism. Michelet et al. (2013) summarize the seminal contributions reported in the 70s and 80s indicating that several Calvin-Benson cycle enzymes are regulated by the ferredoxin/ferredoxin-thioredoxin-reductase/thioredoxin system. Besides this, based on the observation that multiple redox post-translational modifications including glutathionylation and nitrosylation may affect a single protein, Michelet and collaborators propose that these multiple layers of redox regulation could serve for the fine-tuning of the Calvin-Benson cycle enzyme activities in response to changing conditions. Serrato et al. (2013) reviewed the current knowledge on the functions of plastidial TRXs and discuss their emerging role in non-photosynthetic organs. By analyzing transplastomic tobacco plants overexpressing plastidial TRXs f or m, Rey et al. (2013) show that these types of TRXs fulfill distinct physiological functions and propose a role of TRXm in linking photosynthetic activity, redox homeostasis and antioxidant mechanisms in the chloroplast. In another report, López-Calcastro et al. (2014) discuss the possibility that CP12 proteins are part of a redox-mediated metabolic switch in response to rapid environmental changes, in addition to their classical regulatory role of the Calvin-Benson cycle.

Within the photosynthetic context, Karamoko et al. (2013) propose that the enzymatically assisted sulfhydryl oxidation in plant thylakoid lumen is required for the biogenesis of the energy-transducing membrane systems associated to photosynthesis. Their suggestions are based on both the existence of several luminal disulfide bond containing proteins in *Arabidopsis*, and on the discovery of trans-thylakoid redox pathways controlling disulfide bond formation and reduction.

In addition, the importance of NADPH thioredoxin reductase C (NTRC) in plastid redox regulation is also reported in four articles. The paper by Puerto-Galán et al. (2013) discusses the role of NTRC in the control of the overoxidation status of chloroplast 2-Cys peroxiredoxins (2-Cys-PRX), thus having a crucial function balancing the toxic and signaling activities of hydrogen peroxide. Moreover, Richter and Grimm (2013)

emphasize the relevant function of NTRC in conjunction with the FTR/TRX pathway on the redox regulation of tetrapyrrole biosynthesis. This pathway needs to be tightly regulated to adjust chlorophyll content of photosynthetic cells and to avoid the accumulation of chlorophyll biosynthesis intermediates, which may cause oxidative damage. In support of the overarching significance of redox regulation for chloroplast function and adaptation to environmental changes, Chi et al. (2013) discuss the molecular mechanisms and physiological significance of redox-dependent structural changes observed for some proteins including transcriptional factors and co-activators, which switch the chaperone activity of these proteins. Finally, Toivola et al. (2013) report an *in vivo* approach showing the positive effect of NTRC overexpression in plant performance.

The importance of redox regulation in plant mitochondria is also highlighted in this Research Topic by Lázaro et al. (2013). These authors update and discuss the complex antioxidant systems of this organelle, including the complex thiol-based TRX/PRX/Sulfiredoxin (SRX) system. Moreover, these authors focus on the function of this system in the response of plants to abiotic stresses and the regulation by redox post-translational modifications.

Some aspects of the cytosolic redox regulation pathways have also been developed in this Research Topic. Hara and Hisabori (2013) analyzed the kinetics of interactions of a cytosolic TRX *h* with selected target proteins using surface plasmon resonance. The presented data reveal a stronger preference of TRX *h* for an oxidized target, thus explaining the selective association of TRX with oxidized proteins. Zaffagnini et al. (2013) review how cysteine-based modifications of the plant cytosolic glyceraldehyde phosphate dehydrogenase (GAPDH) provoke the inactivation of classical enzymatic activity of this enzyme, but provide additional non-metabolic functions in stress signaling pathways, similar to those that have been identified in animal cells.

GRXs are oxidoreductases of the TRX family which display a particularly rich diversity in higher plants. Beyond classification in three main subgroups based on sequence and structural features, Couturier et al. (2013) indicated that class I GRXs can be subdivided into different groups which can be distinguished by different biochemical and catalytic properties. The importance of a GRX-dependent redox regulation is also evidenced here by Sánchez-Riego et al. (2013) who demonstrate, by performing a physiological and biochemical characterization of single, double and triple *grx* mutants, that GRXs are essential for stress adaptation in cyanobacteria. Finally, Knaff and Sutton (2013) describe an interesting work about the use of GRX to initiate a long-term educational project which will allow examining the structural and biochemical changes in GRXs according to single-point mutational replacements.

Glutathione is a key component in regulation and maintenance of cellular thiol redox homeostasis. It also plays key roles in different aspects of plant development. Here, Rahantaniaina et al. (2013) provide an overview of the main pathways influencing the glutathione redox status and their impact on signaling pathways through regulation of protein thiol status. Schnaubelt et al. (2013) examine the influence of glutathione in plant development and stress tolerance in *Arabidopsis*. The data demonstrate that cellular

glutathione homeostasis influences the root architecture and the leaf area under optimal and stress conditions. Another aspect where glutathione and homoglutathione are crucial molecules is nodule development and in the context of legume-rhizobium mutualistic interactions. Indeed, these organs are peculiar due to the formation of a bacteroid in which the oxygen-sensitive nitrogenase reduces di-nitrogen to ammonia. However, the importance of other redox systems in this unique organ has been poorly documented. Frendo et al. (2013) review the current knowledge on the roles played by redox components in nitrogen-fixing symbioses. Finally, Traverso et al. (2013) summarize the insights of the precise involvement of thiol/disulfide-containing proteins at different stages of the sexual plant reproduction, suggesting specific and critical involvement of thiol-based redox modifications in different reproductive processes.

REFERENCES

- Chi, Y. H., Paeng, S. K., Kim, M. J., Hwang, G. Y., Melencio, S. M. B., Oh, H. T., et al. (2013). Redox-dependent functional switching of plant proteins accompanying with their structural changes. *Front. Plant Sci.* 4:277. doi: 10.3389/fpls.2013.00277
- Couturier, J., Jacquot, J. P., and Rouhier, N. (2013). Toward a refined classification of class I dithiol glutaredoxins from poplar: biochemical basis for the definition of two subclasses. *Front. Plant Sci.* 4:518. doi: 10.3389/fpls.2013.00518
- Frendo, P., Matamoros, M. A., Alloing, G., and Becana, M. (2013). Thiol-based redox signaling in the nitrogen-fixing symbiosis. *Front. Plant Sci.* 4:376. doi: 10.3389/fpls.2013.00376
- Hara, S. and Hisabori, T. (2013). Kinetic analysis of the interactions between plant thioredoxin and target proteins. *Front. Plant Sci.* 4:508. doi: 10.3389/fpls.2013.00508
- Karamoko, M., Gabilly, S. T., and Hamel, P. P. (2013). Operation of trans-thylakoid thiol-metabolizing pathways in photosynthesis. *Front. Plant Sci.* 4:476. doi: 10.3389/fpls.2013.00476
- Knaff, D. B., and Sutton, R. B. (2013). Utility of *Synechocystis* sp. PCC 6803 glutaredoxin A as a platform to study high-resolution mutagenesis of proteins. *Front. Plant Sci.* 4:461. doi: 10.3389/fpls.2013.00461
- Lázaro, J. J., Jiménez, A., Camejo, D., Iglesias-Baena, I., Martí, M. C., Lázaro-Payo, A., et al. (2013). Dissecting the integrative antioxidant and redox systems in plant mitochondria. Effect of stress and S-nitrosylation. *Front. Plant Sci.* 4:460. doi: 10.3389/fpls.2013.00460
- López-Calcano, P. E., Howard, T. P., and Raines, C. A. (2014). The CP12 protein family: a thioredoxin-mediated metabolic switch?. *Front. Plant Sci.* 5:9. doi: 10.3389/fpls.2014.00009
- Michelet, L., Zaffagnini, M., Morisse, S., Sparla, F., Pérez-Pérez, M. E., Francia, E., et al. (2013). Redox regulation of the Calvin-Benson cycle: something old, something new. *Front. Plant Sci.* 4:470. doi: 10.3389/fpls.2013.00470
- Puerto-Galán, L., Pérez-Ruiz, J. M., Ferrández, J., Cano, B., Naranjo, B., Nájera, V. A., et al. (2013). Overoxidation of chloroplast 2-Cys peroxiredoxins: balancing toxic and signaling activities of hydrogen peroxide. *Front. Plant Sci.* 4:310. doi: 10.3389/fpls.2013.00310
- Rahantaniaina, M. S., Tuzet, A., Mhamdi, A., and Noctor, G. (2013). Missing links in understanding redox signaling via thiol/disulfide modulation: how is glutathione oxidized in plants? *Front. Plant Sci.* 4:477. doi: 10.3389/fpls.2013.00477
- Rey, P., Sanz-Barrio, R., Innocenti, G., Ksas, B., Courteille, A., Rumeau, D., et al. (2013). Overexpression of plastidial thioredoxins f and m differentially alters photosynthetic activity and response to oxidative stress in tobacco plants. *Front. Plant Sci.* 4:390. doi: 10.3389/fpls.2013.00390
- Richter, A. S., and Grimm, B. (2013). Thiol-based redox control of enzymes involved in the tetrapyrrole biosynthesis pathway in plants. *Front. Plant Sci.* 4:371. doi: 10.3389/fpls.2013.00371
- Sánchez-Riego, A. M., López-Maury, L., and Florencio, F. J. (2013). Glutaredoxins are essential for stress adaptation in the cyanobacterium *Synechocystis* sp. PCC 6803. *Front. Plant Sci.* 4:428. doi: 10.3389/fpls.2013.00428
- Schnaubelt, D., Schulz, P., Hannah, M. A., Yocgo, R. E., and Foyer, C. H. (2013). A phenomics approach to the analysis of the influence of glutathione on leaf area

- and abiotic stress tolerance in *Arabidopsis thaliana*. *Front. Plant Sci.* 4:416. doi: 10.3389/fpls.2013.00416
- Serrato, A. J., Fernández-Trijueque, J., Barajas-López, J. D., Chueca, A., and Sahrawy, M. (2013). Plastid thioredoxins: a “one-for-all” redox-signaling system in plants. *Front. Plant Sci.* 4:463. doi: 10.3389/fpls.2013.00463
- Toivola, J., Nikkanen, L., Dahlström, K. M., Salminen, T. A., Lepistö, A., Vignols, E., et al. (2013). Overexpression of chloroplast NADPH-dependent thioredoxin reductase in *Arabidopsis* enhances leaf growth and elucidates *in vivo* function of reductase and thioredoxin domains. *Front. Plant Sci.* 4:389. doi: 10.3389/fpls.2013.00389
- Traverso, J. A., Pulido, A., Rodríguez-García, M. I., and Alché, J. D. (2013). Thiol-based redox regulation in sexual plant reproduction: new insights and perspectives. *Front. Plant Sci.* 4:465. doi: 10.3389/fpls.2013.00465
- Zaffagnini, M., Fermani, S., Costa, A., Lemaire, S. D. and Trost, P. (2013). Plant cytoplasmic GAPDH: redox post-translational modifications and moonlighting properties. *Front. Plant Sci.* 4:450. doi: 10.3389/fpls.2013.00450
- Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
- Received: 28 March 2014; accepted: 22 May 2014; published online: 10 June 2014.
Citation: Cejudo FJ, Meyer AJ, Reichheld J-P, Rouhier N and Traverso JA (2014) Thiol-based redox homeostasis and signaling. *Front. Plant Sci.* 5:266. doi: 10.3389/fpls.2014.00266
- This article was submitted to *Plant Physiology*, a section of the journal *Frontiers in Plant Science*.
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