

Hydrogen peroxide effects on *Streptococcus thermophilus* CNRZ368 cell viability

Annabelle Thibessard, Annabelle Fernandez, Brigitte Gintz, Nathalie Leblond-Bourget, Bernard Decaris

► **To cite this version:**

Annabelle Thibessard, Annabelle Fernandez, Brigitte Gintz, Nathalie Leblond-Bourget, Bernard Decaris. Hydrogen peroxide effects on *Streptococcus thermophilus* CNRZ368 cell viability. *Research in Microbiology*, Elsevier, 2001, 152, pp.593 - 596. 10.1016/S0923-2508(01)01234-7 . hal-01631261

HAL Id: hal-01631261

<https://hal.univ-lorraine.fr/hal-01631261>

Submitted on 8 Nov 2017

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Hydrogen peroxide effects on *Streptococcus thermophilus* CNRZ368 cell viability

Annabelle Thibessard, Annabelle Fernandez, Brigitte Gintz, Nathalie Leblond-Bourget*,
Bernard Decaris

Laboratoire de Génétique et Microbiologie, UMR INRA 1128, Faculté des sciences de l'Université Henri Poincaré Nancy I, BP239,
54506 Vandoeuvre-lès-Nancy, France

Received 25 July 2000; accepted 2 February 2001

Abstract – *Streptococcus thermophilus* CNRZ368 is an anaerobic aerotolerant bacteria and its ability to survive under aerobic growth conditions raises the question of the existence of a putative defence system against oxidative stress. Thus, survival of CNRZ368 in the presence of increasing concentrations of hydrogen peroxide was studied. Moreover, the influence of the physiologic state of the cells, as well as that of a preexposition with sublethal doses of hydrogen peroxide, upon *S. thermophilus* CNRZ368 survival were determined. The results suggest that *S. thermophilus* displays a defence system against oxidative stress and that this system is inducible. © 2001 Éditions scientifiques et médicales Elsevier SAS

hydrogen peroxide / oxidative stress / *S. thermophilus* CNRZ368

1. Introduction

The oxidative stress response has been well characterized for some microorganisms and especially for the facultative aerobic Gram-negative bacteria *Escherichia coli* (for a review, [17]). In this organism, two distinct regulons, OxyR [4] and SoxR/S [9, 18], have been identified and are known to be induced by two distinct oxidative molecules, H₂O₂ and O₂⁻ respectively. The case of streptococci is less well documented since only the existence of some anti-oxidant enzymes have been reported [3, 8, 10, 13–15, 19]. Moreover, these bacteria lack catalase [16].

Our research focuses on the facultative anaerobic Gram-positive bacterium *Streptococcus thermophilus*, widely used in dairy product fermentation. Its ability to grow under aerobic conditions raises the question of the existence of a defence system against oxidative stress. Indeed, such a system might be necessary to rescue cells from damage caused by the inevitable reactive oxygen species produced under aerobic growth conditions.

The aim of our work was to test the effect of oxidative stress on *S. thermophilus* survival. For this purpose, H₂O₂ was used as the oxidative agent:

hydrogen peroxide is notably known to react with reduced iron or copper ions to generate hydroxyl radicals (OH•) in the Fenton reaction [11, 12].

2. Materials and methods

2.1. Strains and culture conditions

S. thermophilus CNRZ368 was purchased from the Centre National de la Recherche Zootechnique belonging to the INRA strains collection (France). D₁ was isolated in the progeny of strain CNRZ368 after serial subclonings. All experiments were performed on this clone. First, D₁ was grown overnight at 42°C in milk medium. Overnight culture was then diluted 100-fold in fresh TPPY [1] (0.7% tryptose, 0.7% proteose-peptone, 0.2% bacto yeast extract, 2% lactose) and grown at 42°C.

2.2. Sensitivity to oxidizing agents

D₁ was grown overnight in milk medium, then inoculated into fresh TPPY medium. During exponential growth (OD₆₀₀ = 0.6), culture was exposed to oxidative stress by the addition of hydrogen peroxide (0.25 to 30 mM). Cells were left in contact with hydrogen peroxide for 15 min. Aliquots of the culture were diluted in TPPY, and appropriate dilutions plated on TPPY agar plates to determine cell viability.

* Correspondence and reprints.

E-mail address: bourget@nancy.inra.fr (N. Leblond-Bourget).

Plates were stocked in jars provided with the GENbox Anaer system (bioMérieux). Colonies were counted after 20-h incubation under anaerobic conditions. Percentage survival was determined by comparing viability with or without treatment. All experiments were performed at least in triplicate.

3. Results and discussion

3.1. Effect of increasing concentrations of H₂O₂ on survival

The ability of D₁ (derived from CNRZ368) to respond to 15-min exposure to a range of hydrogen peroxide concentrations (0.25–30 mM) was investigated and led to a depletion of cell viability (*figure 1A*). When D₁ was treated with low doses, the H₂O₂ dose response curve was found to be biphasic: between 0 and 0.5 mM H₂O₂ viability decreased, but then rose again above 0.5 to 1.0 mM (*figure 1B*). Further increases in the level of H₂O₂ above 1 mM resulted in a further loss of viability. The kill curves seen in *figure 1* are very similar to those seen in *E. coli*, although *S. thermophilus* appears to be inherently more sensitive to H₂O₂.

Two hypotheses can be proposed to explain this biphasic shape: the curve may reflect two different

modes of killing by H₂O₂. Such an explanation has been proposed to account for two kinetically distinguishable modes of cell killing by H₂O₂ in *E. coli*. In the first mode, cells are killed primarily by DNA damage caused by OH• radicals produced during the Fenton reaction. However, higher levels of H₂O₂ are thought to scavenge OH•, thus suppressing mode one killing. As the concentration of H₂O₂ rises still further, viability is decreased by a second mode of killing, although the cellular targets in this instance are not known [11, 12]. Alternatively, the curve shape could account for the induction of a protective response above a concentration of 0.5 mM H₂O₂. Above the concentration of 1 mM, viability again decreased, because of the saturation of the protective system.

3.2. The influence of preexposure to H₂O₂ on resistance to oxidative stress

The ability of *E. coli* to resist stress has been shown to increase following prior exposure to low doses of the same stress [5]. Therefore, the viability of D₁ was tested with or without pretreatment with a sublethal level of H₂O₂. As shown in *figure 2*, a 0.25-mM pretreatment led to a 15-fold increase in survival when challenged with 5 mM H₂O₂, compared to strains that had not been pretreated. Moreover, when challenged with 10 mM H₂O₂, no more than 80% of the 0.25 mM H₂O₂ pretreated population was killed, compared to 99% loss of viability for the non-pretreated control. A 1-mM H₂O₂ preexposure led to even higher levels of survival to subsequent exposure: no more than 30% of the population was killed when subsequently challenged with 10 mM H₂O₂. Thus, D₁, when exposed to sublethal levels of H₂O₂, was better able to cope with subsequent toxic doses. These results suggest that *S. thermophilus* cells respond to H₂O₂, probably by activating groups of genes encoding antioxidant and repair enzymes.

3.3. The effect of growth phase on H₂O₂ tolerance

In *E. coli* and *S. typhimurium*, stationary-phase cells are more resistant to environmental stress than exponentially growing cells [6]. To test whether this is the case for *S. thermophilus*, the effect of growth phase on H₂O₂ tolerance was investigated. A culture of D₁ was split into five subcultures, and H₂O₂ was added at four different time points during the ensuing growth period. One culture remained untreated

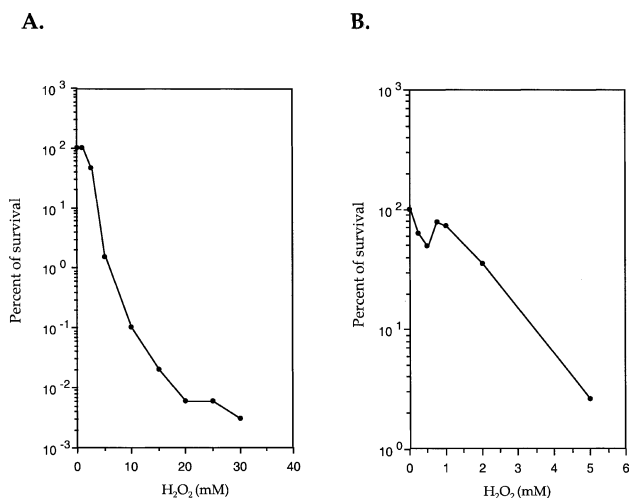


Figure 1. Sensitivity of D₁ isolate to killing by H₂O₂. Survival was determined by plating appropriate dilutions of samples after a 15-min exposure to H₂O₂ of exponentially growing cells (OD_{600 nm} = 0.6). Surviving fractions were calculated relative to colony counts without treatment. These experiments were performed in triplicate and the results of typical experiments are displayed in the figure. A. High concentrations of H₂O₂. B. Low concentrations of H₂O₂.

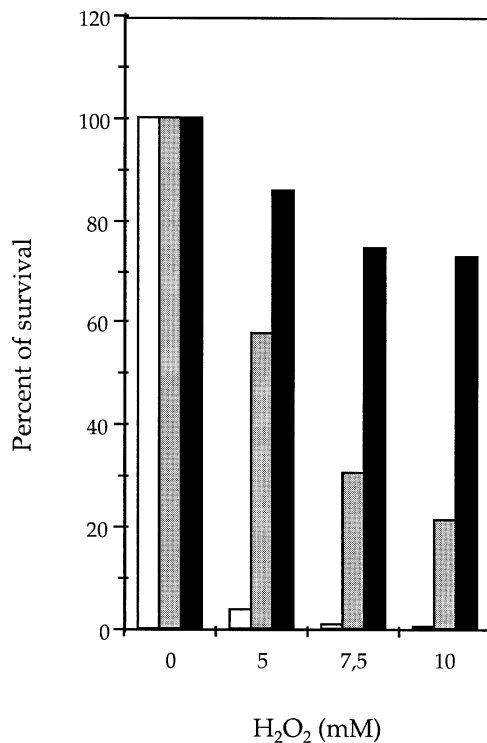


Figure 2. Effect of H₂O₂ pretreatment on cells viability. Cells were preincubated for 15 min at 42°C in TPPY medium in the absence (□) or in the presence of 0.25 mM H₂O₂ (▒) or 1 mM H₂O₂ (■). Cells were then treated again with hydrogen peroxide at the indicated concentration and plated. Percentage of surviving cells is indicated on the graph.

and served to determine the growth rate, which proceeded in TPPY with a doubling time of 30 min⁻¹ (figure 3A). Figure 3B shows H₂O₂ tolerance as a function of the growth phase. H₂O₂ had little effect on the viability of stationary phase cells (OD₆₀₀ = 2.1), even up to 5 mM. In contrast, cells in early exponential phase (OD₆₀₀ = 0.15) were extremely sensitive to H₂O₂, and resistance increased throughout the growth period (OD₆₀₀ = 0.6; OD₆₀₀ = 0.85). Thus, nongrowing *S. thermophilus* cells appeared to be more tolerant to H₂O₂ than their actively growing counterparts as found in *E. coli*. Moreover, since the resistance is settled down progressively through the growth curve, the oxidative stress response in *S. thermophilus* would have at least two modes of induction, one by H₂O₂ and the other by starvation, as seen in *Bacillus subtilis* [7].

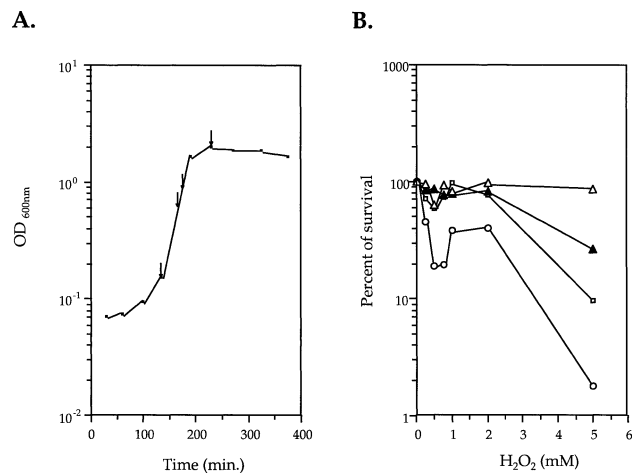


Figure 3. A. Growth curve of *S. thermophilus* D1 strain. B. Change in H₂O₂ tolerance as a function of physiological state. Each point represents the average of a triplicate experiment. Cells were grown in TPPY medium. H₂O₂ was added at different times during the ensuing growth period as a function of the OD₆₀₀. Cells were treated at OD_{600nm} = 0.15 (○), OD_{600nm} = 0.60 (□), OD_{600nm} = 0.85 (▲) and OD_{600nm} = 2.10 (△).

4. Conclusion

Our results suggest the existence of an inducible defence system against H₂O₂ in *S. thermophilus* CNRZ368, possibly of the type OxyR-like regulon. Although the search for the structural homologue of OxyR (the regulator protein of the OxyR regulon) in *Streptococcus mutans* has failed [19], the hypothesis of a functional substitution for the OxyR protein can be drawn. Such functional substitution has already been identified in another Gram-positive bacteria as PerR in *Bacillus subtilis* [2].

Acknowledgements

We are grateful to Mark Paget for his critical reading and his help in the English formulation of the manuscript.

This work was supported by Le Ministère de l'Éducation Nationale, de la Recherche et de la Technologie.

References

- [1] Bracquart P, An agar medium for the differential enumeration of *Streptococcus thermophilus* and *Lactobacillus bulgaricus* in yoghurt, J. Appl. Bacteriol. 51 (1981) 303–305.

- [2] Bsat N., Herbig A., Casillas-Martinez L., Setlow P., Helmann J.D., *Bacillus subtilis* contains multiple Fur homologues: identification of the iron uptake (Fur) and peroxide regulon (PerR) repressors, *Mol. Microbiol.* 29 (1998) 189–198.
- [3] Chang S.K., Hassan H.M., Characterization of superoxide dismutase in *Streptococcus thermophilus*, *Appl. Environ. Microbiol.* 63 (1997) 3732–3735.
- [4] Christman M.F., Storz G., Ames B.N., OxyR, a positive regulator of hydrogen peroxide-inducible genes in *Escherichia coli* and *Salmonella typhimurium*, is homologous to a family of bacterial regulatory proteins, *Proc. Natl. Acad. Sci. USA* 86 (1989) 3484–3488.
- [5] Crawford D.R., Davies J.A., Adaptive response and oxidative stress, *Environ. Health. Perspect.* 102 (1994) 25–28.
- [6] Demple B., Regulation of bacterial oxidative stress genes, *Annu. Rev. Genet.* 25 (1991) 315–337.
- [7] Dows B.C., Murphy P., McConnell D.J., Devine K.M., Relationship among oxidative stress, growth cycle, and sporulation in *Bacillus subtilis*, *J. Bacteriol.* 169 (1987) 5771–5775.
- [8] Gibson C.M., Mallett T.C., Claiborne A., Caparon M.G., Contribution of NADH oxidase to aerobic metabolism of *Streptococcus pyogenes*, *J. Bacteriol.* 182 (2000) 448–455.
- [9] Greenberg J.T., Monach P., Chou J.H., Josephy P.D., Demple B., Positive control of a global antioxidant defense regulon activated by superoxide-generating agents in *Escherichia coli*, *Proc. Natl. Acad. Sci. USA* 87 (1990) 6181–6185.
- [10] Higuchi M., Yamamoto Y., Poole L.B., Shimada M., Sato N., Takahashi N., Kamio Y., Functions of two types of NADH oxidases in energy metabolism and oxidative stress of *Streptococcus mutans*, *J. Bacteriol.* 181 (1999) 5940–5947.
- [11] Imlay J.A., Chin S.M., Linn S., Toxic DNA damage by hydrogen peroxide through the Fenton reaction in vivo and in vitro, *Science* 240 (1988) 640–642.
- [12] Imlay J.A., Linn S., DNA damage and oxygen radical toxicity, *Science* 240 (1988) 1302–1309.
- [13] Nakayama K., Nucleotide sequence of *Streptococcus mutans* superoxide dismutase gene and isolation of insertion mutants, *J. Bacteriol.* 174 (1992) 4928–4934.
- [14] Pébay M., Holl A.C., Simonet J.M., Decaris B., Characterization of the gor gene of the lactic acid bacterium *Streptococcus thermophilus* CNRZ368, *Res. Microbiol.* 146 (1995) 371–383.
- [15] Poole L.B., Higuchi M., Shimada M., Calzi M.L., Kamio Y., *Streptococcus mutans* H₂O₂-forming NADH oxidase is an alkyl hydroperoxide reductase protein, *Free Radic. Biol. Med.* 28 (2000) 108–120.
- [16] Sneath P.H.A. (Ed.), *Bergey's manual of systematic bacteriology*, Williams and Wilkins, Baltimore, 1986.
- [17] Storz G., Zheng M., in: Storz G., Hengge-Aronis R. (Eds.), *Bacterial stress responses*, ASM press, Washington, D.C., 2000, pp. 47–59.
- [18] Tsaneva I.R., Weiss B., soxR, a locus governing a superoxide response regulon in *Escherichia coli* K-12, *J. Bacteriol.* 172 (1990) 4197–4205.
- [19] Yamamoto Y., Kamio Y., Higuchi M., Cloning, nucleotide sequence, and disruption of *Streptococcus mutans* glutathione reductase gene (*gor*), *Biosci. Biotechnol. Biochem.* 63 (1999) 1056–1062.