



**HAL**  
open science

## Variation in variance means more than mean variations: What does variability tell us about population health status?

Simon Devin, Laure Giambérini, Sandrine Pain-Devin

### ► To cite this version:

Simon Devin, Laure Giambérini, Sandrine Pain-Devin. Variation in variance means more than mean variations: What does variability tell us about population health status?. *Environment International*, 2014, 73, pp.282-287. 10.1016/j.envint.2014.08.002 . hal-01726624

**HAL Id: hal-01726624**

**<https://hal.univ-lorraine.fr/hal-01726624>**

Submitted on 8 Mar 2018

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **Variation in Variance means more than Mean Variations: What does variability tell us about**  
2 **population health status?**

3

4 **Simon Devin<sup>1</sup>, Laure Giamberini and Sandrine Pain-Devin**

5 **Université de Lorraine, CNRS UMR 7360, Laboratoire Interdisciplinaire des Environnements**

6 **Continentaux (LIEC) - Metz, France**

7

8

9 Corresponding author: Simon Devin (simon.devin@univ-lorraine.fr), LIEC UMR 7360, Campus  
10 Bridoux, Rue Claude Bernard, Metz, F-57070, France

11

12

1 **Abstract**

2 In environmental science, the variability of biological responses in natural or laboratory populations  
3 is a well known and documented phenomenon. However, while an extensive literature aims to  
4 explain and understand the origin of variability, few try to use it as a demonstration of the  
5 population's response facing a stress. We propose here a theoretical framework that explores  
6 various patterns of variability both within and among populations, and seeks methods useful in  
7 bioevaluation methodologies. We also introduce the concept of "ecotoxicological niche" to  
8 characterize the ability of a population to endure contamination.

9

10 **Keywords**

11 Biomarkers, Variability, Niche, Ecotoxicology, Risk assessment, Population acclimatation

12

13 **Highlights**

- 14 • Variance in biomarkers response along environmental gradients shouldn't be neglected  
15 • Variability reflects acclimatation abilities and could be a precocious response to stress  
16 • Ecotoxicological niche allow to interpret ecological significance of toxicity biomarkers

17

18

1 **Preamble**

2 What is one of the most annoying things that an ecotoxicologist is confronted with? A high variability  
3 of responses between replicates, and perhaps most annoying, a variable variability across conditions.  
4 Indeed, the within-site/condition variability of the measured biological responses is a strong  
5 limitation on detecting the central tendency between site/condition differences, whatever the  
6 statistical test used. As a consequence, the variance is generally only considered as a parameter  
7 limiting the access to parametric statistical analysis, *i.e.* the (h)allowed student and ANOVAs tests.

8

9 **How is variability considered in ecotoxicology literature?**

10 Our scientific literature is strongly dominated by the tyranny of the Golden Mean introduced by  
11 Bennett (1987), that restricts our vision of the dataset to a unique endpoint, and prevents us from  
12 fully exploiting the richness of this dataset. Variance is therefore considered at best as illustrative  
13 information, at worst as a noise, that prevents us from discerning/depicting the true biological  
14 effects, *i.e.* variations of mean along environmental gradients, still considered to be the most useful  
15 way to understand the studied phenomenon.

16 In a recent synthesis, Artigas et al. (2012) proposed a framework for ecotoxicology that pointed out  
17 some new and some recurrent questions, including relevance of ecotoxicological tools. A key part of  
18 this relevance is to go beyond the question of biological endpoints by defining some quantitative  
19 parameters that we should focus on: the mean is of course a pertinent measure, but the variance is  
20 another, as well as differences of basal levels across populations and their range of variation.

21 Indeed, variability is a key parameter in population genetics (Browne et al. 2002), that relies on the  
22 partitioning of phenotypic variance between its genetic and environmental component , as well as in  
23 physiology (Bennett 1987). A rebirth of interest for the variance is also observed in risk analysis (Maul

1 2014) and in ecology, after having been neglected for decades (Violle et al. 2012). In ecotoxicology  
2 also, a quite abundant literature deals with the variability of biological responses. However, it mainly  
3 tries to understand the mechanisms underlying the observed variability, proposes solutions to limit  
4 this variability and deals with the statistical issues and methods to cope with the variability, but few  
5 have explored the information contained in those variability patterns as a new method to better  
6 understand the effect of a contaminant on the studied system (Calow 1996). By focusing on the  
7 mean only, we neglect an important part of the information in our interpretation and probably  
8 misunderstand the true effect of anthropogenic stressors on populations. Indeed, the mean is often  
9 interpreted as a parameter that reflects the highest density of individuals in a distribution, while it's  
10 only its gravity center. In some case (bimodal distribution, asymmetric distribution with extreme  
11 values), it could be a real problem. However, data are often summarized by the mean and the  
12 standard deviation, that gave some indications on the data distribution.

13 If we consider now a classic case of normal distribution of the biological response, the mean +/-  
14 standard deviation interval encompasses roughly only 68% of the population. As pointed out by  
15 Depledge (1990), we thus neglect 32% of the population. These 32%, far from being atypical  
16 individuals, constitute a significant proportion of the population and need to be included in our  
17 reflection about effect of the pollution. Indeed, those individuals could be particularly resistant to  
18 some stresses (Depledge 1990) and survive high pollution, being the basis for a future population  
19 which reconstitutes after an episodic stress or that has adapted in the case of a chronic stress.

20 Thus, since the eighties, regularly, some people have been advocating for a better consideration of  
21 variance in data analysis, and desired that it become a topic in ecotoxicology (Depledge 1990;  
22 Depledge and Fossi 1994; Handy and Depledge 1999; Keppler and Ringwood 2001; Williams 2008;  
23 Corton et al. 2012; Smith et al. 2012). However, we have not yet observed a shift of paradigm from  
24 mean to mean/variance studies, and therefore propose here an additional step in this direction.

1 So, how can we explain inter and intra-population variations of the biological responses, and what  
2 are the general patterns we could define across contamination levels? Once these issues are  
3 understood, the usefulness of this information can be assessed more accurately. The way variability  
4 will be considered will affect environmental risk assessment strategies

5

## 6 **Observed patterns of variability**

7 First, we should define at which level we consider variability. The objects usually compared to each  
8 other are populations. However, the term population can define different objects depending on the  
9 context in which it is used. Calow (1996) deals with the responses of populations of several species  
10 facing a stress, and debates their varying tolerance to pollutants. However, this is not the type of  
11 variance that is most often considered in ecotoxicology, and we focus on variations within and  
12 among populations of one species only. Within-population studies are generally laboratory  
13 experiments where a field population is exposed to a gradient of pollutant, while between-  
14 population studies are generally field experiments where several sites across an environmental  
15 gradient are considered.

16 While such studies mostly focus on the mean, the importance of variance among individuals has  
17 been described in several studies. The major observed patterns are detailed below. However,  
18 explanations for these patterns are rarely proposed by the authors.

19 Depledge and Lundebye (1996) observed high inter-individual variations in the heart rate of crabs in  
20 contaminated sites, and suggested that it was linked to a high response of sensitive individuals while  
21 resistant ones exhibited low responses. Bard (2000) also noticed a higher inter-individual variability  
22 of P-glycoprotein expression in a contaminated site, but only link this to variable abilities of  
23 individuals to respond to P-gp inducers. Odum et al. (1979) presents the increase of variability with  
24 increasing perturbation as a proof of a decrease of system stability, leading to a decrease of its global

1 performance. In the meantime, Luoma (1996) observed a narrowing of the variance of mussel  
2 growth rates on polluted sites, which he explained by a selection of some typical response by the  
3 contaminants. He stated that "high variance in a response may be typical at low pollutant  
4 concentration". Apart from these two opposite patterns, some studies showed either less clear  
5 patterns (Kepler and Ringwood 2001), or both patterns. For example, Baird and Barata (1999)  
6 showed that variance within population can either increase or decrease between sublethal and lethal  
7 contamination levels, depending on the contaminant tested. It thus became excessively low or high  
8 compared to the basal variability observed without contamination (Cairns Jr. 1992).

9

#### 10 **How can we explain differences of variability levels? What are the mechanisms are involved?**

11 While variability is not used *per se* to understand the effect of contamination on natural systems, a  
12 quite abundant literature aims to explain why responses are variable. Mechanisms that structure  
13 variability within populations are acting at four different levels.

14 The first two mechanisms have an intrinsic origin. Differences in the genetic heritage of individuals  
15 enable populations sufficiently large to avoid genetic homogenization, through drift or inbreeding for  
16 example. A conceptual framework for these interactions between genes and environment in the  
17 specific context of contaminated ecosystems has been proposed by Morgan et al. (2007) or Steinberg  
18 et al. (2008). It has been illustrated by Baird & Barata (1999) for *Daphnia* clonal populations, and by  
19 Feckler et al. (2012) for natural populations of *Gammarus fossarum*. However, even in clonal  
20 populations without genetic variability, physiological status (varying according to age, reproduction  
21 or feeding status for example) may influence all the biological responses. Many works have focused  
22 on this physiological status, seeking for sensitive stage (Depledge 1994; Hyne and Maher 2003;  
23 Karimi and Folt 2006; Corton et al. 2012).

1 The latter two mechanisms involve the interactions of the population with its biocenosis and its  
2 biotope, and are of extrinsic origin. They are generally considered through the angle of confounders  
3 that could mask the effect of contaminant and lead to false evaluation of population and ecosystem  
4 health. They could be abiotic factors of natural (temperature, salinity...) or anthropogenic origin  
5 (contamination, but also pH, organic matter...) or biotic factors such species interaction, food  
6 availability, parasitism (Beketov and Liess 2008; Nikinmaa and Tjeerdema 2010; Beketov et al. 2011;  
7 Minguéz et al. 2012; Knillmann et al. 2013).

8 All those mechanisms generally act simultaneously, and the identification of the origin of the  
9 variation is not easy. However, whatever the origin of variation, it contains information which is of no  
10 profit.

11 **What are the expected patterns of variability along environmental gradients, and what is their**  
12 **significance?**

13 We focus only on adverse effects of contaminants on biological systems, thus hormesis (Costantini et  
14 al. 2010) is not implicitly considered here. However, whether beneficial effects of low doses are  
15 homogenous within a population, or if resistant individuals are less affected by low doses than  
16 sensitive individuals should also be considered. The different patterns explored below exclude  
17 extrinsic "non mechanistic" sources of variation (measurement errors, experimental unit differences  
18 - block effects, system stochasticity - see box 1 for the experimental design associated with variance  
19 study). However, we can leave out that an initial source of variation lies on the field sampling  
20 procedure of the population. This factor cannot be controlled easily, while it remains possible to  
21 evaluate the sample size needed to stabilize the sample variance, for example by increasing sample  
22 size until no change in variance was observed. To precisely estimate inter-individual variability and to  
23 compare variability across samples (samples being either from the same population or from different



1 populations), the blame pseudoreplication frequently observed in ecotoxicology became the most  
2 powerful experimental design, limiting the source of variation to the sole individual effect.

3 Finally, the patterns presented thereafter only considered sublethal biological response that  
4 increased following exposure to contaminants. However, the theoretical framework we develop here  
5 is also applicable to biological responses that decrease following exposure to contaminants. These  
6 biological responses could be either a detoxication system, a defense or exposure biomarker, express  
7 the modification of physiological performance, an increase or a decrease in growth rate or in  
8 fecundity.....

### 9 *Intra-population variability along environmental/pollution gradients*

10 During laboratory experiments or active biomonitoring, one population is used in several  
11 conditions. Two dimensions should be considered. For a given exposure condition, the intra-  
12 population variation mainly reflects genetic variability. When all the exposure conditions are  
13 considered, variance patterns show how this genetic variability allows populations to cope with  
14 contamination. We first focus on sublethal levels of exposure, *i.e.* the stage where the contaminant  
15 begin to have an impact on the organism without leading to the death of any individual. Obviously,  
16 this stage is strongly dependant of the exposure duration, with differences between short and long  
17 term lethality thresholds. However, a general pattern can be drawn, and the main difference  
18 between these two exposure conditions is in the scale of the horizontal axis rather than in the  
19 general aspect of the response. Thus, three patterns are possible: a stable variance, a decreasing or  
20 an increasing variance. They are presented in box 2. Only the principal trends are detailed there, and  
21 obviously intermediate and mix situations could be observed, with for example a canalization of the  
22 response at sublethal levels, followed by an increase in variance at lethal levels, when  
23 regulation/defense systems are overcome.

1 Once the lethality threshold is exceeded, that is, when the first deaths occur (indicated by  $CL_{10}$  in box  
2 2), we should observe either no modification of the variance or a decrease, following the death of  
3 individuals exhibiting the most extreme values. It could be accompanied by a shift of the mean,  
4 according to the relationships between individual response and survival. It could be the driver of  
5 stabilizing selection (box 2c), cases (d) and (e) are directional selection. They all led, over some  
6 generations, to a type (a) response, but with probable differences in basal and maximum levels.

7 These variation patterns could help us to understand the results of active biomonitoring, with a  
8 population sampled in a reference station and transplanted into a contaminated one. Thus, with this  
9 theoretical framework, we will develop three situations of mean variation (Fig. 1).

10 Case 1 is the simplest one: when facing contamination, all individuals die. Thus the diagnosis is easy,  
11 the environment is toxic! Case 2 is the favorite of scientists and editors: a clear and indubitable  
12 increase in mean is observed. Beyond the mean, the variance could also change, with the patterns  
13 detailed in box 2.

14 Finally, case 3 is classically considered as a "no effect" response. However, while it's true for the  
15 population 1a, the two others presented variation in variance, if not in mean. We must not neglect  
16 this effect that could be the early warning signal that everybody is looking for. Indeed, the shift in  
17 variance can occur before the shift in mean, though numerous case studies are needed to correctly  
18 interpret such results.

19

## 20 *Inter-population variability along environmental/pollution gradients*

21 Passive biomonitoring cases are the most difficult to interpret, because they are the outcome  
22 of a gene x environment interaction. By comparing several populations to a reference system, several  
23 patterns could be observed. The values observed are not the responses to a change of environmental

1 condition, but the chronic, basal level of the studied parameter in the local environmental condition.  
2 Thus, the objective is to understand whether the differences observed could be relevant indicators of  
3 population health (Fig. 2).

4 The simplest result is the stability of both mean and variance that could be easily interpreted as the  
5 absence of environmental stress. If no difference in mean is observed, while a difference in variance  
6 exists (Fig 2., A vs B vs C), we can postulate that the chronic levels of contamination are similar  
7 among the two studied sites. However, the phenotypic plasticity revealed by the response variability  
8 tells about the genetic variability and the adaptative abilities of the population. It has been shown  
9 that genetic variability is a key factor for population health, thus populations at risk are those with  
10 the lowest variability.

11 The most complicated cases are those where several populations exhibit different mean levels of  
12 their responses (A-B-C vs D-E-F). Indeed, without information on the temporal variation, we cannot  
13 distinguish short term induction in response to a temporary stress or long term adaptation or  
14 acclimatation to a chronic stress. In such situations, even the study of the variance is of no help.  
15 Either temporal trends of the biological response or results of laboratory stress on stress  
16 experiments would be helpful to understand these complex interactions. This leads us to question, in  
17 an interpopulation perspective, the meaning of the basal level (in mean and variance), and how  
18 useful it is to predict and interpret the response pattern. Such approaches are poorly developed.  
19 They reveal within and between population phenotypic variability, that could be the outcome of  
20 stochastic phenomenon (genetic drift or founder effect), but also the results of local adaptation  
21 through selection by the contaminant pressure, or a simple acclimatation to local conditions. More  
22 studies are necessary, either in the field or laboratory, to explore and understand the mechanisms  
23 and the consequences of modification of variance patterns.

24

## 1 **Towards the ecotoxicological niche: an ecological perspective to ecotoxicology**

2 Information necessary to understand and exploit the different patterns of response within and  
3 between populations relies on the natural variability, *i.e.* the range of values of the studied biological  
4 response in a range of healthy ecosystems and for healthy populations, to correctly define the  
5 baseline level (Xuereb et al. 2009; Coulaud et al. 2011; Lacaze et al. 2011; Jubeaux et al. 2012).

6 We can then define the ecotoxicological niche as the ability of species to cope with contamination  
7 and to maintain basal physiological function when facing anthropogenic stress. This ecotoxicological  
8 niche is thus a particular case of the traditional ecological niche, where the environmental gradient is  
9 replaced by a chemical compound. Using this well known conceptual framework, we can adapt the  
10 Shelford law (1931) and define a well-being zone (Figure 3), corresponding to the realized niche of a  
11 population, a stress zone, corresponding to the potential niche, and a lethal zone, where acclimation  
12 abilities are overwhelmed.

13 The main dimensions of this niche should be the classical ecotoxicological parameters related to  
14 genetic and cellular damages, physiological and behavioral modifications, defense or exposure  
15 system activation, but also more integrative parameters directly related to fitness (reproduction,  
16 survival and growth). For each of these dimensions, the realized niche is defined as the inter-  
17 individual variability (95% confidence interval, for instance) observed *in situ*. To get a complete  
18 overview of this realized niche, those parameters should at least be measured at several time scales,  
19 depending on the life cycle of the species. The theoretical niche is not conceptually difficult to define,  
20 as it should be drawn from the observation of several populations of the considered species, in  
21 various environmental conditions (of pollution, geochemistry, hydromorphology...). Thus, it is just a  
22 compilation of many realized niches. The most difficult parameter to evaluate is the potential niche,  
23 which is the extreme deviation from the baseline that allows the population to maintain itself in the  
24 ecosystem. This raises two problems: how to make sure that we measured the extreme deviation,

1 and how to ensure the sustainability of the population? It needs to combine an experimental and a  
2 modeling approach. Experiments performed in realistic conditions (mesocosms), on long term to  
3 allow acclimation phenomena, will be used to measure biological response levels. In the meantime,  
4 population dynamics models based on the Dynamic Energy Budget in Toxicology (DEBtox) need to be  
5 built to assess population sustainability (Billoir et al. 2007). The combination of those two  
6 information will define the potential ecotoxicological niche defined above.

7 Once the three niche scales described, the population-level information (realized and potential) can  
8 be compared to the species level information (theoretical niche), this comparison solving the  
9 problem of interpopulation comparison, giving a general framework to interpret every response  
10 observed. Indeed, if populations get different realized niches, but similar potential niches, we can  
11 suppose that they are living in contrasted environments according to contamination. In the  
12 meantime, if those differences in realized niche are small compared to the theoretical niche of the  
13 species, we can suppose that the level of stress is far from the maximal level that they are able to  
14 undergo. Finally, the potential niche breadth informs us on the sensitivity of each population to  
15 stress. A next step should be to link ecotoxicological niche breadth to biological and ecological traits  
16 such as feeding habits and food, maximum size, substrate preferences, trophic status (Poisot et al.  
17 2011; Colas et al. 2014).

18

### 19 **Consequences for Environmental Risk Assessment (ERA) approaches**

20 The main objective of ERA approaches is to define the health status of a population. A frequent  
21 confusion/debate is about the scale at which we should interpret biological responses measured on  
22 an individual. We think that a misuse/misunderstanding is to consider that those parameters should  
23 be interpreted as proxy of the individual health status. Well, they obviously reflect the individual  
24 health status, but those individuals are sampled from a population, and thus also reflect the

1 population health status. Thus, individual-based measurements are population relevant indicators,  
2 and as a consequence, variance between individuals should be interpreted as a populational  
3 response.

#### 4 *Sacrifice complexity (and realism) on the altar of reproducibility*

5 Biomarker approaches are often criticized for their lack of ecological relevance (McCarty et al 2002).  
6 Indeed, to get fine information about the ecosystem health, a common approach is to focus on the  
7 most sensitive species and on the most sensitive stages. However, finding such species looks like an  
8 unattainable goal (Cairns Jr. 1992) and takes us away from a pertinent ecological risk assessment  
9 (Baird and Barata 1999). In the same way, considering the spatio-temporal variability of physiological  
10 status, the most sensitive stage at one time (which has been selected for ecotoxicology tests) could  
11 be more resistant at another time (*i.e.* females should be more sensitive to lipophilic contaminants  
12 before gestation than during).

13 Moreover, with the aim of controlling variability, ecotoxicology tests are based on a very specific  
14 category of natural population. However, doing that, these tests are not suitable to build metric  
15 representatives of the whole population. The response variability evidences phenotypic variants  
16 within a population. For example, decreasing genetic diversity by selecting some clonal population of  
17 *Daphnia* leads to an increase in uncertainty when performing field extrapolation (Barata et al. 2002).  
18 Thus, with a limited, biased variability, we do not get a good idea of the adaptative possibilities  
19 within the population, nor of the resilience/resistance abilities of the population. Therefore, rather  
20 than minimize variability, it would be better to model the relative influence of intrinsic and extrinsic  
21 factors on the studied biological response to better understand the system (Handy et al. 2003).

22 Finally, to demonstrate the effects of pollution and to predict population health, variance must  
23 become a component of the evaluation scheme, as it can reflect system resistance and resilience

1 against perturbation (the assimilative capacity hypothesis (Cairns Jr. 1977, 1999; Death 2010; Steudel  
2 et al. 2012)).

### 3 **Acknowledgments**

4 We want to thank Sharon Kruger for her linguistic corrections. This approach was developed within  
5 the EC2CO Sydepop and ANR IPOC programs. We thank the reviewers whose comments helped us to  
6 improve the manuscript.

### 7 **References**

- 8 Artigas J, Arts G, Babut M, Caracciolo AB, Charles S, Chaumot A, et al. Towards a renewed research  
9 agenda in ecotoxicology. *Environmental Pollution*. 2012; 160: 201–6.
- 10 Baird DJ, Barata C. Genetic variation in the response of *Daphnia* to toxic substances: implications for  
11 risk assessment. *Genetics and ecotoxicology*. Taylor and Francis. London: Forbes V.E.; 1999.  
12 p. 207–21.
- 13 Barata C, Markich SJ, Baird DJ, Taylor G, Soares AMVM. Genetic variability in sublethal tolerance to  
14 mixtures of cadmium and zinc in clones of *Daphnia magna* Straus. *Aquatic Toxicology* 2002;  
15 60: 85–99.
- 16 Bard SM. Multixenobiotic resistance as a cellular defense mechanism in aquatic organisms. *Aquatic*  
17 *Toxicology*. 2000 ;48: 357–89.
- 18 Bastos AC, Monaghan KA, Pestana JLT, Lillebø. AI, Loureiro S. A comment on the Editorial “Replication  
19 in aquatic biology: The result is often pseudoreplication.” *Aquatic Toxicology* 2013; 126: 467-  
20 470
- 21 Beketov MA, Liess M. An indicator for effects of organic toxicants on lotic invertebrate communities:  
22 independence of confounding environmental factors over an extensive river continuum.  
23 *Environmental Pollution*. 2008; 156: 980–7.
- 24 Beketov MA, Speranza A, Liess M. Ultraviolet radiation increases sensitivity to pesticides: synergistic  
25 effects on population growth rate of *Daphnia magna* at low concentrations. *Bulletin of*  
26 *environmental contamination and toxicology*. 2011; 87: 231–7.
- 27 Bennett AF. Interindividual variability: an underutilized resource. *New Directions in Ecological*  
28 *Physiology*. Cambridge University Press. Cambridge: Feder M.E., Bennett A.F., Burggren W.W.  
29 and Huey R.B.; 1987.
- 30 Billoir E, Péry ARR, Charles S. Integrating the lethal and sublethal effects of toxic compounds into the  
31 population dynamics of *Daphnia magna*: A combination of the DEBtox and matrix population  
32 models. *Ecological Modelling*. 2007; 203: 204–14.

- 1 Browne R., Moller V, Forbes V., Depledge M. Estimating genetic and environmental components of  
2 variance using sexual and clonal *Artemia*. Journal of Experimental Marine Biology and  
3 Ecology. 2002; 267: 107–19.
- 4 Cairns Jr. Aquatic ecosystem assimilative capacity. Fisheries. 1977; 2: 5.
- 5 Cairns Jr. J. The threshold problem in ecotoxicology. Ecotoxicology. 1992; 1: 3–16.
- 6 Cairns Jr. J. Assimilative capacity — the key to sustainable use of the planet. Journal of Aquatic  
7 Ecosystem Stress and Recovery. 1999; 6: 259–63.
- 8 Calow P. Variability: noise or information in ecotoxicology? Environmental Toxicology and  
9 Pharmacology. 1996; 2: 121–3.
- 10 Colas F, Vigneron A, Felten V, Devin S. The contribution of a niche-based approach to ecological risk  
11 assessment: Using macroinvertebrate species under multiple stressors. Environmental  
12 Pollution. 2014; 185: 24–34.
- 13 Corton JC, Bushel PR, Fostel J, O’Lone RB. Sources of variance in baseline gene expression in the  
14 rodent liver. Mutation Research - Genetic Toxicology and Environmental Mutagenesis. 2012;  
15 746: 104–12.
- 16 Costantini D, Metcalfe NB, Monaghan P. Ecological processes in a hormetic framework. Ecology  
17 Letters. 2010; 13:1435–47.
- 18 Coulaud R, Geffard O, Xuereb B, Lacaze E, Quéau H, Garric J, et al. *In situ* feeding assay with  
19 *Gammarus fossarum* (Crustacea): Modelling the influence of confounding factors to improve  
20 water quality biomonitoring. Water Research. 2011; 45:6417–29.
- 21 Death RG. Disturbance and riverine benthic communities: What has it contributed to general  
22 ecological theory? River Research and Applications. 2010; 26: 15–25.
- 23 Depledge M. The rational basis for the use of biomarkers as ecotoxicological tools. In: Fossi M,  
24 Leonzio C, editors. Nondestructive biomarkers in vertebrates. Lewis Publishers; 1994. p. 261–  
25 85.
- 26 Depledge M.H., Lundebye A.K. Physiological Monitoring of Contaminant Effects in Individual Rock  
27 Crabs, *Hemigrapsus Edwardsi*: The Ecotoxicological Significance of Variability in Response.  
28 Comparative Biochemistry and Physiology -- Part C: Pharmacology, Toxicology and  
29 Endocrinology. 1996; 113: 277–82.
- 30 Depledge MH. New approaches in ectotoxicology: can inter-individual physiological variability be used  
31 as a tool to investigate pollution effects? Ambio. 1990; 19: 251–2.
- 32 Depledge MH, Fossi MC. The role of biomarkers in environmental assessment (2). Invertebrates.  
33 Ecotoxicology. 1994; 3: 161–72.
- 34 Drummond GB, Vowler SL. Variation: use it or misuse it – replication and its variants. Journal of  
35 Physiology. 2012; 590: 2539–42.



- 1 Feckler A, Thielsch A, Schwenk K, Schulz R, Bundschuh M. Differences in the sensitivity among cryptic  
2 lineages of the *Gammarus fossarum* complex. *Science of The Total Environment*. 2012; 439:  
3 158–64.
- 4 Handy RD, Depledge MH. Physiological Responses: Their Measurement and Use as Environmental  
5 Biomarkers in Ecotoxicology. *Ecotoxicology*. 1999; 8: 329–49.
- 6 Handy RD, Galloway TS, Depledge MH. A Proposal for the Use of Biomarkers for the Assessment of  
7 Chronic Pollution and in Regulatory Toxicology. *Ecotoxicology*. 2003; 12: 331–43.
- 8 Hyne RV, Maher WA. Invertebrate biomarkers: links to toxicosis that predict population decline.  
9 *Ecotoxicology and Environmental Safety*. 2003; 54: 366–74.
- 10 Jubeaux G, Simon R, Salvador A, Lopes C, Lacaze E, Queau H, et al. Vitellogenin-like protein  
11 measurement in caged *Gammarus fossarum* males as a biomarker of endocrine disruptor  
12 exposure: Inconclusive experience. *Aquatic Toxicology* 2012; 122: 9–18.
- 13 Karimi R, Folt CL. Beyond macronutrients: element variability and multielement stoichiometry in  
14 freshwater invertebrates. *Ecology Letters*. 2006; 9: 1273–83.
- 15 Keppler C, Ringwood AH. Expression of P-glycoprotein in the gills of oysters, *Crassostrea virginica*:  
16 seasonal and pollutant related effects. *Aquatic Toxicology*. 2001; 54: 195–204.
- 17 Knillmann S, Stampfli NC, Noskov YA, Beketov MA, Liess M. Elevated temperature prolongs long-term  
18 effects of a pesticide on *Daphnia* spp. due to altered competition in zooplankton  
19 communities. *Global Change Biology*. 2103; 19: 1598-1609.
- 20 Lacaze E, Devaux A, Mons R, Bony S, Garric J, Geffard A, et al. DNA damage in caged *Gammarus*  
21 *fossarum* amphipods: A tool for freshwater genotoxicity assessment. *Environmental*  
22 *Pollution*. 2011; 159: 1682–91.
- 23 Lazic S. The problem of pseudoreplication in neuroscientific studies: is it affecting your analysis? *BMC*  
24 *Neuroscience*. 2010; 11: 5.
- 25 Luoma SN. The developing framework of marine ecotoxicology: Pollutants as a variable in marine  
26 ecosystems? *Journal of Experimental Marine Biology and Ecology*. 1996; 200: 29–55.
- 27 Maul A. Heterogeneity: A Major Factor Influencing Microbial Exposure and Risk Assessment. *Risk*  
28 *Analysis*. 2014; In press : DOI: 10.1111/risa.12184.
- 29 Minguez L, Buronfosse T, Beisel J-N, Giambérini L. Parasitism can be a confounding factor in assessing  
30 the response of zebra mussels to water contamination. *Environmental Pollution*. 2012; 162:  
31 234–40.
- 32 Morgan AJ, Kille P, Stürzenbaum SR. Microevolution and Ecotoxicology of Metals in Invertebrates.  
33 *Environmental Science and Technology* 2007; 41: 1085–96.
- 34 Nikinmaa M, Tjeerdema R. Environmental variations and toxicological responses. *Aquatic Toxicology*.  
35 2013; 127: 1.
- 36 Nikinmaa M., Celander M., Tjeerdema R. Replication in aquatic biology: The result is often  
37 pseudoreplication. *Aquatic Toxicology*. 2012; 116-117: iii–iv.

- 1 Odum EP, Finn JT, Franz EH. Perturbation Theory and the Subsidy-Stress Gradient. *BioScience*. 1979;  
2 29: 349–52.
- 3 Poisot T, Bever JD, Nemri A, Thrall PH, Hochberg ME. A conceptual framework for the evolution of  
4 ecological specialisation. *Ecology Letters*. 2011; 14: 841–51.
- 5 Smith R, Mann R, Fox D. Variability in ecotoxicology: deliberate ignorance or just not getting it. 6th  
6 SETAC World Congress / SETAC Europe 22nd Annual Meeting , Berlin, Germany; 2012.
- 7 Steinberg CEW, Stürzenbaum SR, Menzel R. Genes and environment — Striking the fine balance  
8 between sophisticated biomonitoring and true functional environmental genomics. *Science*  
9 of The Total Environment. 2008; 400: 142–61.
- 10 Steudel B, Hector A, Friedl T, Löffke C, Lorenz M, Wesche M, et al. Biodiversity effects on ecosystem  
11 functioning change along environmental stress gradients. *Ecology Letters*. 2012; 15: 1397–  
12 405.
- 13 Violle C, Enquist BJ, McGill BJ, Jiang L, Albert CH, Hulshof C, et al. The return of the variance:  
14 intraspecific variability in community ecology. *Trends in Ecology & Evolution*. 2012; 27: 244–  
15 52.
- 16 Williams TD. Individual variation in endocrine systems: moving beyond the “tyranny of the Golden  
17 Mean.” *Philosophical Transactions of the Royal Society – Biological Sciences*. 2008; 363:  
18 1687–98.
- 19 Xuereb B, Chaumot A, Mons R, Garric J, Geffard O. Acetylcholinesterase activity in *Gammarus*  
20 *fossarum* (Crustacea Amphipoda): Intrinsic variability, reference levels, and a reliable tool for  
21 field surveys. *Aquatic Toxicology*. 2009; 93: 225–33.

22

23

24

25

26

27

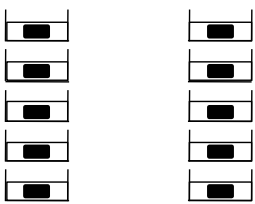
28

29

30

- 1 Box 1: experimental design (for recent review and analysis of pseudoreplication, see(Bastos et al.;
- 2 Lazic 2010; Drummond and Vowler 2012; Nikinmaa M. et al. 2012).

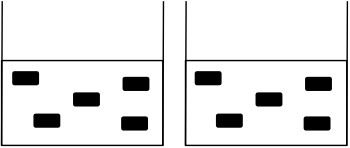
If you focus on mean, the good practice is :



5 « independant » experimental units per condition  
 1 measure per unit  
 = 5 replicates

Condition A Condition B

BUT if you focus on variance, the good practice is :



1 experimental units per condition  
 5 measure per unit  
 = 5 pseudoreplicates

Condition A Condition B

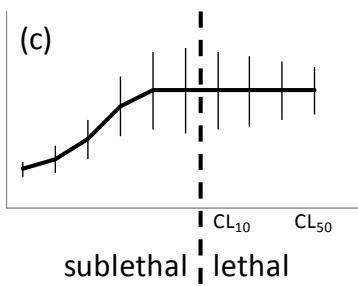
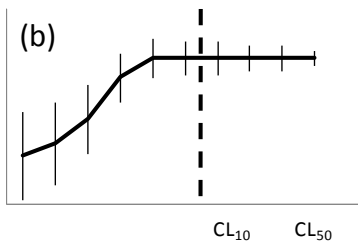
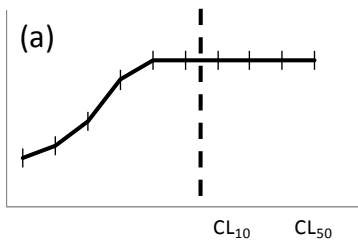
Replicated samples take into account two sources of variability : experimental units differences and inter-individual differences. Pseudo-replicated samples only consider the variability within population. The best way is to mix the two methods, that allows to work on the mean and on the variance.

3

4

1

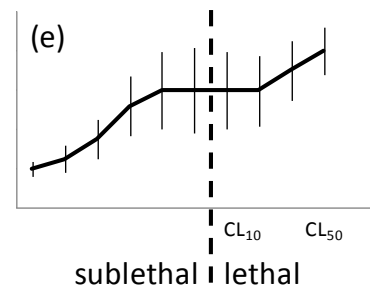
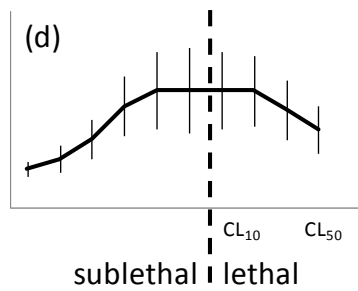
## 2 Box 2 :Intra-population variance scenarios



The case considered is an increase of the measured parameter in response to contamination. Beside the mean, several trends in the variance could be observed.

**Sublethal exposure.** The variance could be constant across conditions (a), revealing a low phenotypic variability, probably associated to a low genetic variability. A decrease in variance (b), often associated to a high variability of basal levels, could be interpreted as a canalization of the response to the most efficient one. Finally, the variance could increase (c). This last case could be linked to a continuous variation of the response in the interval observed, or to the constitution of several distinct groups of individuals (low, intermediate and high values for instance). Thus this high variability at concentrations near the lethality threshold is linked to individuals that exhibit extreme values, reflecting either low resistance abilities or a risk for the biological responses to be overcome (i.e. once the highest possible value is attained, health impairments are expected). Those two extreme situations can lead to death if the stress is maintained.

**Lethal exposure.** Over the lethality threshold, a decrease in variance indicates that all individuals displaying extreme values are dead. If the mean decreases (d), we can suppose that individuals exhibiting the highest responses are dead and that high induction is a signal of a soon overcoming of regulation/defense system. The last case (e) is an increase of the mean, indicating that low values of the response are linked to insufficient induction and an inability of individuals to protect themselves.

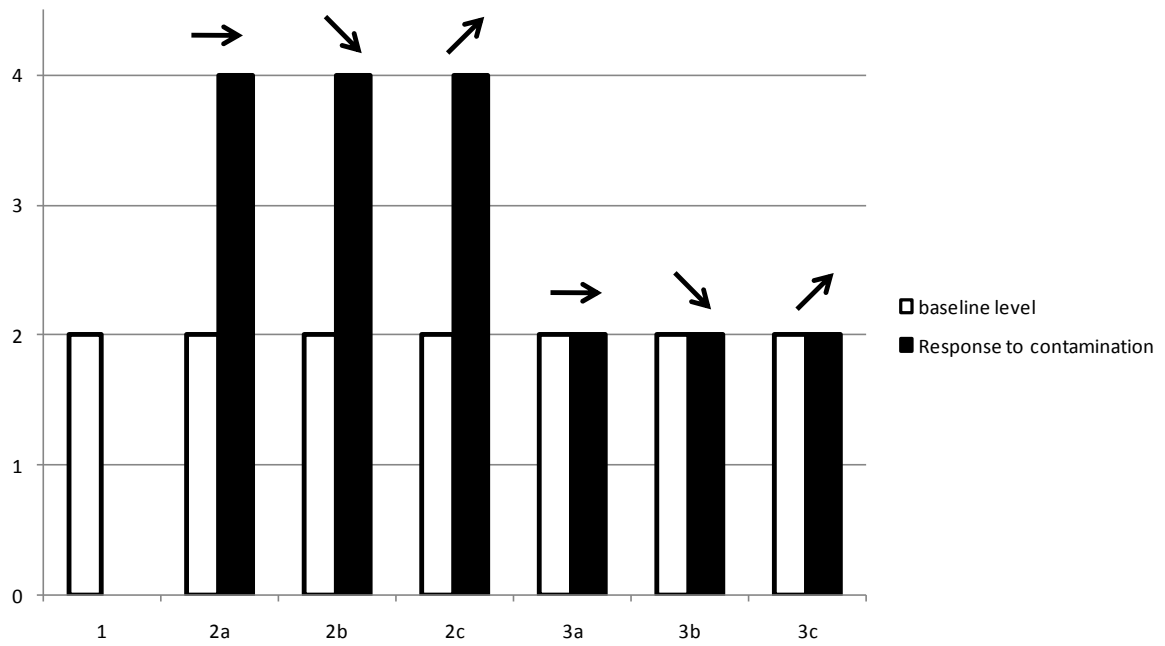


3

4

1

2 Figure 1



3

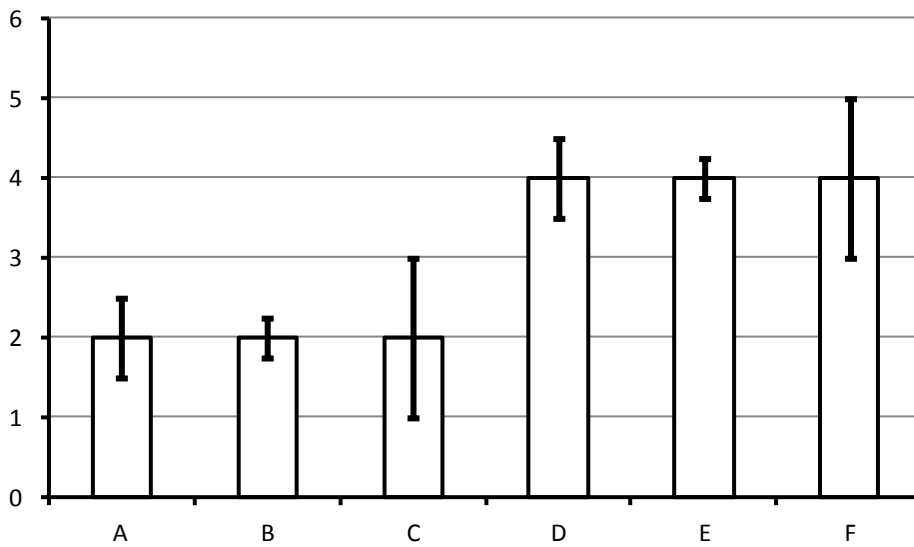
4 Figure 1: Main mean/variance response to contamination in an active biomonitoring study. Arrows

5 indicates the direction of variance changes.

6

1

2 Figure 2.

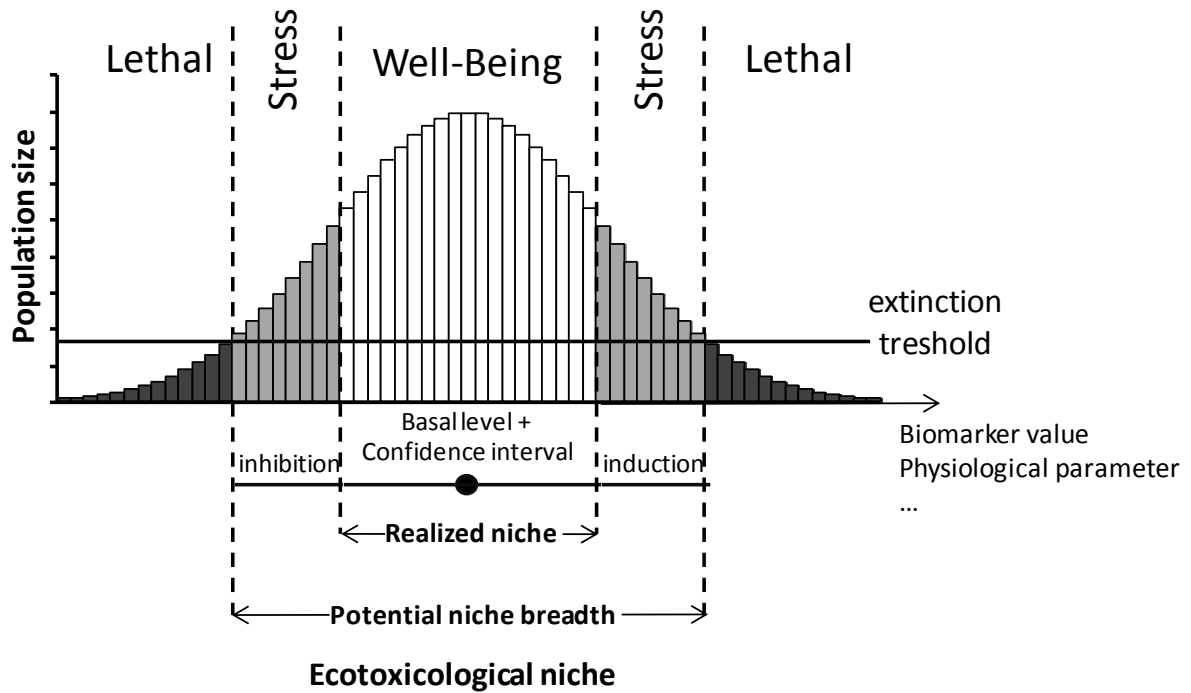


3

4 Figure 2: Main mean/variance response to contamination in a passive biomonitoring study. The  
5 observed biological response is induced when organisms are exposed to a toxicant. A is the reference  
6 population. By comparing other population to the reference one, we can observe either a stability of  
7 mean with differences in variance (B and C) or differences in means without (D) or with (E, F)  
8 differences in variance.

9

1



2

3 Figure 3: The ecotoxicological niche. The optimal health status for a given population is attained for  
4 its realized niche, *i.e.* when the natural, chronic conditions are encountered, contamination included.  
5 This niche thus encompasses acclimation and adaptation phenomenon. The presence of a  
6 supplementary contamination represents an important deviation from these standard conditions,  
7 leading to an increase or a decrease of the biological response, depending on the parameter  
8 considered. A first level still allows the population to maintain itself, with lower abundances: it's the  
9 potential niche. Then this potential niche is overcome, the environment became too far from the  
10 adaptative abilities of the population, which lead to local extinction.

11