

# **Variability in Sexual Pheromones Questions their Role in Bumblebee Pre-Mating Recognition System**

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 **Abstract -** Sex-specific chemical secretions has been widely used as diagnostic characters in chemotaxonomy. The taxonomically confused group of bumblebees has reaped the benefit of this approach through the analyses of cephalic labial gland secretions (CLGS). Most of currently available CLGS descriptions concern species from the West-Palearctic region and few from the New World. Here, the CLGS descriptions of four East-Palearctic species *B. deuteronymus*, *B. filchnerae*, *B. humilis* and *B. exil* (subgenus *Thoracobombus*) are performed. Our results underline a strong variability in main compounds in *B. exil*. In contrast, we describe a low differentiation in CLGS compounds between *B. filchnerae* and its closely phylogenetically related taxon *B. muscorum*. Moreover, the chemical profile of *B. filchnerae* and *B. muscorum* is characterized by a highly concentrated component of the length C18 while it is a C16 in the other *Thoracobombus* species. It raises a possibility that the courtship behavior as well as environmental constraints could affect the role of the bumblebee males cephalic labial glands secretions.

**Key Words** - Courtship behavior, pheromones, cephalic glands

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#### INTRODUCTION

 In many animal species, sex-specific chemical secretions are involved in the pre-mating communication (Patricelli et al. 2003; Hay 2011; Lecocq et al. 2015a). This key role in reproductive behavior has led to the development of the chemotaxonomy in which such chemical traits are used as diagnostic characters to assess taxonomic statuses (Quicke 1993; Bagnères et al. 2003; Pokorny et al.2014). The systematics of insects has especially reaped the benefits of this approach (Lockey 1991). Bumblebees give a stunning example of the improvements raised by chemotaxonomy (Bertsch and Schweer 2011; Lecocq et al. 2015b). In this taxonomically confused group, morphological traits are often poorly efficient diagnostic characters because different species can be morphologically highly similar while conspecific populations can be extraordinarily divergent in their coat color pattern (Michener 1990; Williams 1998; Lecocq et al. 2015b). In contrast, the male cephalic labial gland secretions (CLGS) are known to be species-specific semiochemicals (Calam 1969; Ayasse and Jarau 2014). These secretions are produced *de novo* (Luxová et al. 2003; Žáček et al. 2013) and they attract conspecific virgin females (Ayasse and Jarau 2014). They appear to provide efficient diagnostic characters for species delimitation (Svensson 1979; Bergström 1981; Terzo et al. 2003; Rasmont et al. 2005; Lecocq et al. 2015c).

 During the last decades, an increasing number of studies has based their taxonomic conclusions on inter-taxa comparisons of CLGS composition (Lecocq et al. 2015b,d). However, global taxonomic assessment of bumblebee species through chemotaxonomy requires an extensive overview of the CLGS composition. Currently the CLGS composition have been described for approximately 65 species (Terzo et al. 2003; Terzo et al. 2007; Rasmont et al. 2005; Ayasse and Jarau 2014; Lecocq et al. 2015c,d; Brasero et al. 2015) out of a total of 250 bumblebee species around the world (Williams 1998).

 Most of currently available CLGS descriptions concern species from the West-Palearctic region (Calam 1969; Rasmont et al. 2005; Lecocq et al. 2015b) and some from the New World (Bertsch et al. 2004, 2008; Brasero et al. 2015). In contrast, bumblebee fauna from East-Palearctic region has received far less attention to date (Bergström et al. 1981). Yet, this area hosts the main bumblebee species diversity hotspot (Williams 1998).

 Beyond taxonomic matter, more and more ecological questions about the role of compounds emitted by bumblebees emerge within the scientific community (Lhomme et al. 2015; Brasero et al. 2017). However, compared to several neural coding studies focused on sex pheromones in honey bees (Galizia and Menzel 2001; Sandoz 2012; Carcaud et al. 2015), only few studies have begun to describe neurophysiological structures in bumblebees (Fonta and Masson 1984; Strube-Bloss et al. 2015).

 In this study, the CLGS of four East-Palearctic taxa were investigated: *Bombus deuteronymus superequester* (Skorikov 1914) (Fig. 1 A); *B. filchnerae* Vogt 1908 (Fig. 1 B), *B. humilis subbaicalensis* Vogt 1911 (Fig. 1 C) and *B. exil* (Skorikov 1923) (Fig. 1 D) all belonging to the *Thoracobombus* Dalla Torre, 1880 subgenus*.* Moreover, we compared the CLGS composition of *B. filchnerae* with that of its nearest Palearctic taxon; *B. muscorum* (L. 1758) (Fig. 1 E, F).

#### METHODS AND MATERIALS

 *Sampling.* 65 queens of *B. deuteronymus superequester, B. exil, B. filchnerae* and *B. humilis subbaicalensis* have been collected in Mongolia in 2014. Twenty were transported alive and reared in Belgium. In order to produce males, we maintained queens in a wooden box in a 91 controlled climate, dark room at  $26/30^{\circ}$ C and 50-60% relative humidity, and fed ad libitum with salix pollen and sugar syrup (Lhomme et al. 2013). We produced 51 males (Tab. 1): *B. deuteronymus superequester* (n=10), *B. exil* (n=13), *B. filchnerae* (n=15) and *B. humilis*   *subbaicalensis* (n=13). We assessed the taxonomic identification of males using the taxonomical revision of North China bumblebees published by An et al. (2014).

 Prior to chemical analyses, all males were killed by freezing at -20°C and their CLGS were 97 extracted in 400 µl hexane (De Meulemeester et al. 2011). Samples were stored at -40<sup>o</sup>C prior to analyses. In order to compare the CLGS compositions of *B. filchnerae*, we used its phylogenetic sister taxon *B. muscorum* (n=24) partially published (Lecocq et al. 2015d) (Tab. 1).

 *Chemical Analyses.* We identified the CLGS composition by gas chromatography-mass spectrometry (GC/MS) using a Focus GC (Thermo Scientific) with a non-polar DB-5 ms capillary column [5% phenyl (methyl) polysiloxane stationary phase; column length 30 m; inner diameter 0.25 mm; film thickness 0.25 μm] coupled to DSQ II quadrupol mass analyzer (Thermo Scientific) with 70 eV electron ionization. We used a splitless injection mode 106 (220 $^{\circ}$ C) and helium as a carrier gas (1 ml/min). The temperature program of the oven was set 107 to 70 $^{\circ}$ C for two minutes and then heated up at a rate of 10 $^{\circ}$ C/min to 320 $^{\circ}$ C. The temperature 108 was then held at 320 $\degree$ C for five minutes. Compounds were identified in Xcalibur<sup>TM</sup> using the 109 retention times  $(t_r)$  and mass spectra of each peak, in comparison to those at National Institute of Standards and Technology library (NIST, U.S.A). Double bond positions (C=C) were determined by dimethyl disulfide (DMDS) derivatization (Cvacka et al. 2008).

 We quantified all samples using a gas chromatograph-flame ionization detector Shimadzu GC-2010 with a SLB-5ms non-polar capillary column (5% phenyl (methyl) polysiloxane stationary phase; 30-m column length; 0.25-mm inner diameter; 0.25-µm film thickness) with the same chromatographic conditions as in GC/MS. Peak areas of compounds were detected in GCsolution Postrun (Shimadzu Corporation) with automatic peak detection and noise measurement. We calculated relative amounts (RA in %) of compounds in each sample by dividing the peak areas of compounds by the total area of compounds in each sample. We

 discarded all compounds for which RA were lower than 0.1% for all specimens (De Meulemeester et al. 2011). Moreover, All compounds having a RA equal to or greater than 1% are considered to be abundant compounds. We elaborated the data matrix as the alignment of each compound between all samples performed with GCAligner 1.0 (Dellicour and Lecocq, 2013a,b)(see supplementary file 1).

 *Statistical Analyses.* Statistical analyses were performed using R (R Development Core Team 2013) to detect CLGS differentiations between taxa. Data consisting of the relative contents of all compounds were standardized (mean=0, standard deviation=1), to reduce the sample-concentration effect (Brasero et al. 2015).

 Clustering method was used to detect the differentiation between taxa. A Pearson correlation matrix based on CLGS data matrix was computed. An unweighted pair group method with arithmetic mean (UPGMA) was used as clustering method (R-package ape,(Suzuki and Shimodaira 2011)). We assessed the uncertainty in hierarchical cluster analysis using *p*-values calculated via multiscale bootstrap resampling with 50000 bootstrap replications (R-package pvclust, (Suzuki and Shimodaira 2011)). We calculated CLGS differentiations of taxa by performing a *multiple response permutation procedure (MRPP)* (R package vegan, (Oksanen et al. 2011)).

 To determine compounds specific to each taxa (indicator compounds, IC), we used the *indicator value (IndVal)* method (Dufrêne and Legendre 1997; Claudet et al. 2006). The value given is the product of relative abundance and relative frequency of occurrence of a compound within a group. The statistical significance of an indicator compound (threshold of 0.01) was evaluated with a randomization procedure (Dufrêne and Legendre, 1997).

 In order to visualize the indicator compounds, we compared *B. filchnerae* and its sister species *B. muscorum* by *principal component analyses* (PCA; R-package MASS, (Venables and Ripley, 2002)).

#### RESULTS

 129 compounds were detected in the CLGS of the species belonging to *Thoracobombus* subgenus: 36 in *B. deuteronymus superequester*, 70 in *B. exil*, 33 in *B. filchnerae,* 35 in *B.* 

- *muscorum* and 42 in *B. humilis* (Supplementary file 1).
- The main compounds detected were (i) for *B. deuteronymus superequester*: octadec-11-en-1-

ol (44.40% - 57.05%); (ii) for *B. exil*: hexadec-9-enal (0.04% - 44.35%), octadec-9-enoic acid

(1.36% - 15.20%), icos-11-en-1-ol (0.20% - 8.88%) and hexadecenyl hexadecenoate (0.69% -

10.53%); (iii) for *B. filchnerae and B. muscorum*: octadec-9-enyl acetate (40.49% - 75.96%)

 and (iv) for *B. humilis subbaicalensis:* octadec-9-en-1-ol (40.12% - 56.64%) (Tab. 2; supplementary file 1).

 Our chemical analyses showed qualitative differentiations between all taxa including specific main compounds (i.e. compounds that have the highest relative amount among all compounds of CLGS at least in one individual of the taxon) except between *B. filchnerae* and *B. muscorum* (Tab. 2) where only minor compounds diverged (Tab. 3).

 Our statistical analyses confirmed the differentiations of our sampling in five groups supported by high multiscale bootstrap resampling values: (i) *B. exil*, (ii) *B. humilis subbaicalensis,* (iii) *B. deuteronymus superequester,* (iv) *B. filchnerae* and (v) *B. muscorum* (Fig. 2A). Moreover, a multiple response permutation procedure (MRPP) confirmed these differentiations (all p-values < 0.001): *B. deuteronymus vs B. exil* (A= 0.2993); *B. deuteronymus vs B. filchnerae* (A=0.3326); *B. deuteronymus vs B. humilis* (A=0.3868); *B. deuteronymus vs B. muscorum* (A=0.2551); *B. exil vs B. filchnerae* (A=0.2876); *B. exil vs B. humilis* (A=0.3059); *B. exil vs B. muscorum* (A=0.2501); *B. filchnerae vs B. humilis*  (A=0.3547); *B. filchnerae vs B. muscorum* (A=0.186) and *B. humilis vs B. muscorum* (A=0.2648). Several significant indicator compounds were revealed by the IndVal method 168 (IndVal  $> 0.70$ ) (Supplementary file 1).

#### DISCUSSION

 *B. humilis subbaicalensis and B. deuteronymus superequester.* Based on main and abundant compounds (Tab. 1), our CLGS analyses underline a differentiation between *B. humilis subbaicalensis* and *B. deuteronymus superequester* (Fig. 2A). This differentiation is also observed in genetic markers between these two species (Cameron et al. 2007). However, both genetic and chemical analyses are based on few samples only. This could lead to a misestimation of the inter-taxon differentiation (Lecocq et al. 2015c). Further studies based on larger sampling are needed to assess this differentiation.

 *The intraspecific variability of main compounds in Bombus exil. Bombus exil* is characterized by a high intraspecific variability of its CLGS composition (supplementary file 1). Indeed, *B. exil* displays four main components (Tab. 2) and 45 indicator compounds (supplementary file 1). Moreover, males of *B. exil* have the last three articles of the antennae broadened and ventrally concave (Tkalců 1974) (Fig. 1D). They press this clubbed part of the antennae against the lower part of the face of female bees in a characteristic way (P. Williams: com. pers. based on unpublished personal observations from Moerdaga, Neimenggu, 18/08/2009). These modified antennae deserve more functional morphology studies and histological researches dealing particularly on the sensillae in the concave area on the last article.

 *The case of Bombus filchnerae and B. muscorum questions the role of minor compounds.* As mentioned in Terzo et al. (2005) for *B. ruderarius*, it is possible that abundant compounds are absent within a same taxon (i.e. for *B. filchnerae*; Tab. 1). Despite this intraspecific variability, low interspecific CLGS differentiation is observed between *B. muscorum* and *B. filchnerae* (Supplementary file 1; Tab. 2,3; Fig. 2 A,B). Such a low CLGS differentiation  pattern could be explained by (i) the conspecificity of the two taxa or (ii) the lack of role of CLGS in pre-mating communication. While further taxonomic assessment is needed, we speculate that the latter hypothesis could explain our results. First, CLGS is a complex mixture of (mainly aliphatic) compounds (Calam 1969; Coppée et al. 2008) that can be classified into four groups: (i) C1 to C4 exclusively considered as gas; (ii) C5 to C8 considered as volatile liquids; (iii) C9 to C16 as non-volatile liquids and (iv) C16 and more considered as solid at a temperature of 20°C (Cicolella 2008). By comparing composition between these two species and other *Thoracobombus* (*B. exil, B. deuteronymus superequester, B. humilis subbaicalensis, B. pascuorum*, *B. inexspectatus*, *B. ruderarius* and *B. sylvarum*) (Terzo et al. 2005; Lecocq et al. 2015b; Brasero et al. 2017), we highlight that the chemical profile of *B. filchnerae* and *B. muscorum* is characterized by a high concentrated component of the length C18 in the beginning of the spectra while they are mostly C16 in the other species of *Thoracobombus*. This could suggest that the CLGS of *B. muscorum* and *B. filchnerae* are not used for long distance intersex attraction but rather could play a recognition role at a short distance. This is further supported by observed and described aggregations of *B. muscorum* males in close proximity to mature nests entrance in order to mate with virgin queens (Krüger 1951; Darvill et al.2007). Considering these observations, a sexual selection favoring reproductive isolation through CLGS divergence is not expected if reproductive isolation is mediated by alternative cues such as nest site choice (i.e. the habitat) or nest scent. Further studies on the pre-mating recognition of bumblebee species are needed to assess this hypothesis.

 While most authors only take into account the major compounds in their discussion, our results show that between *B. filchnerae* and *B. muscorum*, the CLGS differentiation is marked on minor compounds. This observation questions the role of these minor compounds on  bumblebee olfaction. In this perspective, an approach on the neuronal process induced by odorant information in the bumblebee brain will provide first elements of thought.

 Indeed, several neural coding studies in Hymenoptera have shown the capacity of their olfactory system to grant special properties to mixtures of pheromonal components as well as to recognize individual components (Galizia and Menzel 2001; Sandoz 2012; Carcaud et al. 2015). In social insects, species use a wide range of different components which induce a simultaneous activation of several classes of olfactory receptor neurons (Sandoz et al. 2007). In honey bees, most of the results were obtained using volatile hydrocarbons that differ in carbon chain length (C5-C10) and functional groups (Sachse et al. 1999; Carcaud et al. 2012). However, some studies addressed the effect of temperature on neural responses to odorants of low volatility (Carcaud et al. 2015) and realized that brood and queen compounds (C10 to C20) were detected only at hive temperature, i.e. around 35°C (Brill et al. 2013; Carcaud et al. 2015).

 Since environmental conditions appear to influence the neural response to odorants (Carcaud et al. 2015), it is difficult to identify which compounds emitted by the CLGS of males induce an neuronal answer under natural conditions in bumblebees (in or out of the nest). In vivo calcium imaging or neural representation studies, as developed in Carcaud et al (2012) and Strube-Bloss et al. (2015), tested in wild conditions (temperature and humidity) would mark a great first step to go deeper into the biological relevance understanding of the emitted males message by observing what is or is not detected by females.

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## **Figure Legends**



 **Fig. 1** A: *Bombus deuteronymus superequester* from Mongolia (photo: P. Rasmont). B: *B. filchnerae* from Mongolia (photo: P. Rasmont). C: *B. humilis subbaicalensis* from Mongolia (photo: P. Rasmont). D: *B. exil* from Mongolia (photo: P. Rasmont). E: *B. muscorum muscorum* from Holland. F: *B. muscorum liepetterseni* from Norway



 **Fig. 2** A: *Unweighted pair group method with arithmetic mean cluster based on a correlation matrix calculated from matrix of cephalic labial gland secretions of Bombus deuteronymus superequester, B. filchnerae, B. humilis subbaicalensis, B. exil and B. muscorum*. Values above branch represent multiscale bootstrap resampling. The letters indicate the geographic origins (C: Corsica; R: Russia; I: Ireland; N: Norway, P: Poland, D: Denmark; S: Sweden, F: France, A: Aran island and H: Holland). B: CLGS Principle components analyses (PCA) of *B. muscorum* (blue squares) and *B. filchnerae* (yellow squares). PC1 and PC2 are first and second axes of PCA. In the left corner, the variable factor map. Each letter (code) correspond to characteristic compounds revealed by IndVal method (Tab. 3)

#### 432 **Supplementary file**

433 **Supplementary file 1** - Data matrix of cephalic labial gland secretions (CLGS) (relative 434 concentration of each compound), list of the identified compounds, kovats indice and IndVal 435 analysis with specific compounds in *B. deuteronymus superequester*, *B. exil*, *B. filchnerae*, *B.*  436 *muscorum* and *B. humilis subbaicalensis*. U indicates undetermined compounds.

437 *Table* 

### 438 Table 1. SAMPLING



439 Collecting sites and number (N) of bred specimens for *Bombus deuteronymus superequester,*  440 *B. filchnerae, B. humilis subbaicalensis and B. exil, B. muscorum* were integrated in the study

- in order to check their CLGS composition with their East-Palearctic subspecies or closely
- related species. Coordinate are given with the reference to the WGS84.

### Table 2. LIST OF MAIN COMPOUNDS



Main compounds (grey background) identified in cephalic labial gland secretions of *Bombus deuteronymus superequester*, *B. exil*, *B. filchnerae*,

*B. muscorum* and *B. humilis subbaicalensis*. Carbons: the number of carbon atoms; MW: the molecular weight; M: median of relative

concentration of compound (%). Abundant compounds are on black background. All identified compounds in Supplementary file1.

## Table 3. LIST OF COMPOUNDS CALCULATED WITH THE INDICATOR VALUE (INDVAL) METHOD FOR *B. MUSCORUM* AND *B.*

#### *FILCHNERAE*



List of compounds identified in cephalic labial gland secretions of *B. muscorum* and *B. filchnerae* after the identification of characteristic compounds (grey background) calculated with the Indicator Value (IndVal) method (>0.70). MW: the molecular weight; M: median of relative concentration of compound (%). The compounds we were not able to determine are indicated as *Ux*. The full table with minimum and maximum data (min and max [%]) are in the supplementary file 1.