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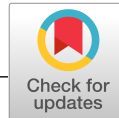
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## RESEARCH ARTICLE

# Engineering *Pseudomonas putida* KT2440 for the production of isobutanol

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We engineered *P. putida* for the production of isobutanol from glucose by preventing product and precursor degradation, inactivation of the soluble transhydrogenase SthA, overexpression of the native *ilvC* and *ilvD* genes, and implementation of the feedback-resistant acetolactate synthase AlsS from *Bacillus subtilis*, ketoacid decarboxylase KivD from *Lactococcus lactis*, and aldehyde dehydrogenase YqhD from *Escherichia coli*. The resulting strain *P. putida* Iso2 produced isobutanol with a substrate specific product yield ( $Y_{\text{Iso/S}}$ ) of  $22 \pm 2$  mg per gram of glucose under aerobic conditions. Furthermore, we identified the ketoacid decarboxylase from *Carnobacterium maltaromaticum* to be a suitable alternative for isobutanol production, since replacement of *kivD* from *L. lactis* in *P. putida* Iso2 by the variant from *C. maltaromaticum* yielded an identical  $Y_{\text{Iso/S}}$ . Although *P. putida* is regarded as obligate aerobic, we show that under oxygen deprivation conditions this bacterium does not grow, remains metabolically active, and that engineered producer strains secreted isobutanol also under the non-growing conditions.

## KEYWORDS

isobutanol, ketoacid decarboxylase, metabolic engineering, microaerobic, *Pseudomonas putida*

## 1 | INTRODUCTION

Biofuel production from renewable feed stocks is of special importance because of the finite nature of the currently used crude oil derivatives and growing concerns about climate change [1]. Isobutanol is an attractive alternative to the employed fossil fuels. It has several advantages such as a higher energy density, compatibility with existing engines, lower vapor pressure and volatility, as well as a lower

corrosivity compared to bio-ethanol [2,3]. Furthermore, isobutanol is used in the chemical industry and can be used to produce the gaseous alkene precursor isobutene [4].

Isobutanol can be synthesized via the branched-chain amino acid biosynthesis and the so-called Ehrlich pathway to convert pyruvate to isobutanol (Figure 1). The first step in this route is the conversion of two pyruvate molecules to 2-acetolactate catalyzed by the acetolactate synthase (AlsS), which is usually feedback inhibited by the branched-chain

Abbreviations: 2-KIV, 2-ketoisovalerate; AlsS, acetolactate synthase; BHI, brain–heart infusion; KDC, ketoacid decarboxylase; LB, Lysogeny broth.

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amino acids L-valine, L-leucine, and L-isoleucine. However, AlsS from *Bacillus subtilis* has been shown to be feedback-resistant and therefore has been applied for isobutanol production in several studies [5,6]. Then, 2-acetolactate is reduced to 2,3-dihydroxyisovalerate and subsequently converted to 2-ketoisovalerate (2-KIV) by the ketoacid reductoisomerase IlvC and dihydroxyacid dehydratase IlvD, respectively. Finally, isobutanol is synthesized from 2-KIV in two more reaction steps of the Ehrlich pathway. The decarboxylation of 2-KIV to isobutyraldehyde is catalyzed by ketoacid decarboxylases (KDCs) that are not widespread in nature. Especially KivD from *Lactococcus lactis* has been proved as an efficient variant in, e.g. *E. coli* and *C. glutamicum* [5,7]. The last step from isobutyraldehyde to isobutanol requires an aldehyde reductase or alcohol dehydrogenase. A number of NADH and NADPH dependent enzymes are available that catalyze this reaction [8].

Several microorganisms have been engineered for isobutanol production such as *E. coli*, *C. glutamicum*, *B. subtilis*, and yeast such as *Saccharomyces cerevisiae* [5,7,9,10]. Although highly efficient *E. coli* and *C. glutamicum* strains have been constructed [6,7], the relatively low tolerance of most microbial systems against isobutanol hampers commercialization of isobutanol production processes. In contrast, pseudomonads have an intrinsic tolerance against organic compounds and solvents [11,12] making them promising candidates for isobutanol production.

Among them, *Pseudomonas putida* is a Gram-negative, saprophytic soil bacterium with a genome size of 6.18 Mbp [13]. It has been reported to promote plant growth, prevent plant diseases, and can efficiently remove organic soil pollutants and environmental contaminants [14]. *P. putida* features a versatile metabolism using the Entner–Doudoroff pathway for glucose catabolism, shows resistance against oxidative stress conditions, and genetic engineering tools are readily available [15–17]. The carbohydrate substrate spectrum is limited and confined to hexoses [18], however, *P. putida* has been recently engineered to concomitantly consume xylose, cellobiose, and glucose, which are the basic building blocks of the abundant polysaccharides cellulose and hemicellulose [19]. As a result of these achievements, *P. putida* has emerged as a promising candidate for industrial biotechnology [20,21]. Recent works have engineered this bacterium for the production of polyhydroxyalkanoates, the nylon precursor *cis,cis*-muconic acid [22] and aromatic compounds like *p*-coumaric acid or *trans*-cinnamate [23,24]. *P. taiwanensis* VLB120 has been applied for the production of phenol [25,26].

In this study, we engineered *P. putida* for the production of isobutanol from glucose by preventing product and precursor degradation and increasing the flux from pyruvate towards isobutanol. We identified KivD from *Carnobacterium maltaromaticum* as a suitable alternative to KivD from *L. lactis*

## PRACTICAL APPLICATION

The relatively low tolerance of most microbial systems against isobutanol hampers commercialization of isobutanol production processes. In contrast, pseudomonads have an intrinsic tolerance against organic compounds and solvents making them promising candidates for isobutanol production. Therefore, we engineered *Pseudomonas putida* KT2440 for the production of this alcohol by preventing product and precursor degradation and increasing the flux from pyruvate toward isobutanol. The achieved overall isobutanol yield is significantly higher compared to other engineered *P. putida* strains; however, rather low compared to tailored *E. coli* and *C. glutamicum* strains. Therefore, this study paves the way to construct more efficient *P. putida* strains for isobutanol production in future studies.

to drive the decarboxylation of 2-ketoisovalerate and finally we showed that isobutanol production can also be achieved under oxygen deprivation conditions with this obligate aerobic bacterium.

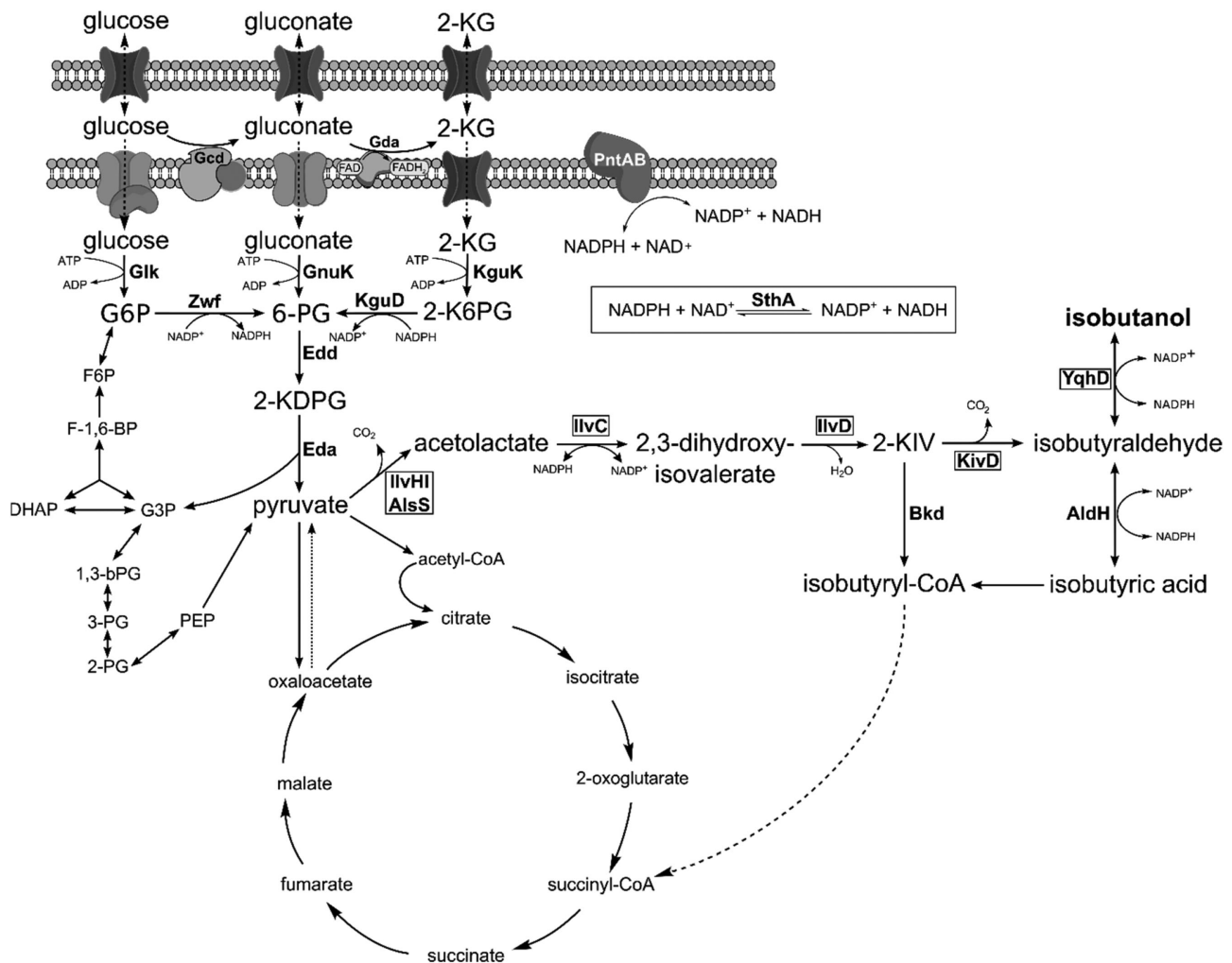
## 2 | MATERIALS AND METHODS

### 2.1 | Bacterial strains and plasmids

Bacterial strains, their respective genotype, plasmids, and oligonucleotides used in this study are listed in Table 1.

### 2.2 | Media and culture conditions

*E. coli* DH5 $\alpha$  was grown aerobically in Lysogeny broth (LB) complex medium containing 10 g/L tryptone, 5 g/L yeast extract, and 10 g/L NaCl [34] at 37°C as 5 mL cultures in glass test tubes on a rotary shaker at 120 rpm (Infors AG, Bottmingen, Switzerland). *C. maltaromaticum* and *L. lactis* were grown in brain–heart infusion (BHI) broth (Carl Roth GmbH & Co. KG, Karlsruhe, Germany) at 30°C on a rotary shaker at 120 rpm. For longtime storage, *P. putida* was kept as 30% (w/v) glycerol stock at –70°C and was streaked out for cultivation on LB solid medium with 15 g/L agar. The first preculture of *P. putida* was prepared by inoculation of 5 mL LB medium in a test tube with a single colony. The culture was cultivated at 30°C on a rotary shaker (Edmund Bühler GmbH, Bodelshausen, Germany) at 175 rpm overnight and used to inoculate, a second overnight preculture to an optical density at 600 nm (OD<sub>600</sub>) of 0.01–0.02 in 50 mL DeBont minimal medium (pH 7) [35], which was supplemented with



**FIGURE 1** The central metabolism of *P. putida* KT2440 with the Ehrlich pathway. Abbreviations (coding genes are given in brackets): G6P: glucose-6-phosphate 2-KG: 2-ketogluconate, 2-K6PG: 2-keto-6-phosphogluconate, 6-PG: 6-phosphogluconate, 2-KDPG: 2-keto-3-deoxy-6-phosphogluconate, G3P: glyceraldehyde-3-phosphate, 1,3-bPG: 1,3-bisphosphoglycerate, 3-PG: 3-phosphoglycerate, 2-PG: 2-phosphoglycerate, PEP: phosphoenolpyruvate, DHAP: dihydroxyacetone-phosphate, F-1,6-bP: fructose-1,6-bisphosphate, F6P: fructose-6-phosphate, CoA: co-enzyme A, Gcd: glucose dehydrogenase (*gcd*), *gad*: gluconate 2-dehydrogenase (*gad*), PQQ: pyrroloquinoline quinone, Glk: glucokinase (*glk*), Zwf: glucose-6-phosphate 1-dehydrogenase (*zwf-1*, *zwf-2*, *zwf-3*), GnuK: gluconate kinase (*gnuK*), KguD: 2-6-phosphoketogluconate reductase (*kguD*), KguK: 2-ketogluconate kinase (*kguK*), Edd: 6-phosphogluconate dehydratase (*edd*), Eda: 2-keto-3-deoxy-6-phosphogluconate aldolase (*eda*), IlvHI/AlsS: acetolactate synthase (*ilvHI/alsS*), IlvC: ketolactate reductoisomerase (*ilvC*), IlvD: dihydroxyacid dehydratase (*ilvD*), KivD: ketoacid decarboxylase (*kivD*), Bkd: branched-chain ketoacid dehydrogenase complex (*bkd*), YqhD: aldehyde reductase (*yqhD*), AldH: aldehyde dehydrogenases, PntAB: pyridine nucleotide transhydrogenase (membrane bound) (*pntAB*), SthA: pyridine nucleotide transhydrogenase (soluble) (*sthA*)

5.4 g/L glucose and 0.5 g/L yeast extract. Cells from the second preculture were harvested by centrifugation ( $4500 \times g$ , 15 min,  $4^\circ\text{C}$ ), resuspended in DeBont medium, and used to inoculate 50 mL DeBont medium, to an  $\text{OD}_{600}$  of about 0.1–0.2. The main culture was supplemented with 5.4 g/L glucose, 0.5 g/L isobutanol, or 2.9 g/L 2-ketoisovalerate, respectively. The second pre- and main cultures were performed in 500 mL baffled Erlenmeyer flasks filled with 50 mL medium on a rotary shaker at 175 rpm at  $30^\circ\text{C}$ . Micro-aerobic shaking flask cultivations were carried out in sealed 100 mL Müller-Krempel bottles as 25 mL cultures that

were inoculated to an  $\text{OD}_{600}$  of 15–20. To obtain sufficient biomass, the second preculture was performed in 100 mL LB medium in a 500 mL baffled Erlenmeyer flask that was cultivated on a rotary shaker (175 rpm) overnight at  $30^\circ\text{C}$ . Cells from the second preculture were harvested by centrifugation ( $4500 \times g$ , 15 min,  $4^\circ\text{C}$ ) and resuspended in 25 mL DeBont minimal medium (pH 7) supplemented with 5.4 g/L glucose and 15 g/L 3-morpholino-propanesulfonic acid. To induce plasmid-based gene expression, 0.2% (*w/v*) L-arabinose was supplemented. For plasmid-bearing strains, 50  $\mu\text{g}/\text{mL}$  kanamycin or 50  $\mu\text{g}/\text{mL}$  apramycin were added to the medium.

**TABLE 1** Overview of strains, plasmids and oligonucleotides used in this study

Strain, plasmid or oligonucleotide	Relevant characteristic(s) or sequence (5' → 3')	Source, reference or purpose
Strains		
<i>Pseudomonas putida</i> KT2440	Wild type strain, DSM-6125, ATCC47054	[27]
<i>Carnobacterium maltaromaticum</i> LMA28		[28]
<i>Lactococcus lactis</i> subsp. <i>cremoris</i> MG1363		[29]
<i>Corynebacterium glutamicum</i>	Wild type strain ATCC13032	American type culture collection
<i>P. putida</i> GN346	<i>P. putida</i> KT2440 $\Delta upp$ , $\Delta pedE$ , $\Delta pedI$ , $\Delta pedH$ , $\Delta aldB-I$	[30]
<i>P. putida</i> EP1	<i>P. putida</i> GN346 $\Delta bkdAA$	This work
<i>P. putida</i> EP2	<i>P. putida</i> EP1 $\Delta sthA$	This work
<i>P. putida</i> EP3	<i>P. putida</i> EP2 $\Delta gcd$	This work
<i>P. putida</i> Iso1	<i>P. putida</i> EP1 + pIP02	This work
<i>P. putida</i> Iso2	<i>P. putida</i> EP2 + pIP02	This work
<i>P. putida</i> Iso3	<i>P. putida</i> EP2 + pIP03	This work
<i>P. putida</i> Iso4	<i>P. putida</i> EP2 + pIP04	This work
<i>P. putida</i> Iso5	<i>P. putida</i> EP3 + pIP02	This work
<i>P. putida</i> Iso6	<i>P. putida</i> EP2 + pIP05	This work
Plasmids		
pBB1	pACYC184/pBL1 derivative, chloramphenicol resistance, P <sub>tac</sub> promoter and <i>trpA</i> terminator	[31]
pSA55	Expression plasmid for <i>adh2</i> of <i>S. cerevisiae</i> and <i>kivD</i> of <i>L. lactis</i>	[5]
pBB1 <i>yqhD</i>	pBB1 P <sub>tac</sub> <i>yqhD</i>	This work
pBB1 <i>kivD yqhD</i>	pBB1 P <sub>tac</sub> <i>kivD yqhD</i>	This work
pNG413.1	pBBR1MCS2 derivative, apramycin resistance, <i>araC</i> , P <sub>BAD</sub> , <i>lacZ</i>	[32]
pSEVA231	pBBR1 derivative, kanamycin resistance, mobilizable ( <i>oriT</i> )	[33]
pIP01	pSEVA231 P <sub>tac</sub> <i>kivD yqhD</i>	This work
pIP02	pNG413 <i>araC</i> P <sub>BAD</sub> <i>kivD yqhD alsS ilvC ilvD</i>	This work
pIP03	pIP02, <i>yqhD</i> was changed for <i>adhA</i> from <i>L. lactis</i>	This work
pIP04	pIP02, <i>yqhD</i> was changed for <i>adhA</i> from <i>C. glutamicum</i>	This work
pIP05	pIP02, <i>kivD</i> was changed for <i>kdcA</i> from <i>C. maltaromaticum</i>	This work
pEMP04	pSEVA231 P <sub>tac</sub> <i>kivD yqhD alsS ilvC ilvD</i>	Ingenza Ltd.
pEMP012	pEMP04, <i>yqhD</i> was changed for <i>adhA</i> from <i>L. lactis</i>	This work
pEMP013	pEMP04, <i>yqhD</i> was changed for <i>adhA</i> from <i>C. glutamicum</i>	This work
pEMP014	pEMP04, <i>kivD</i> was changed for <i>kdcA</i> from <i>C. maltaromaticum</i>	This work
Oligonucleotide		
yqh1	AACTGCAGAACCAATGCATTGGAGGAGACACAACA TGAACAACCTTTAATCTGCACACCCCAACC	Construction of pBB1yqh1, PstI site underlined
yqh2	CCGCTCGAGAAAGCTTAGCGGGCGGCTT CGTATATACG	Construction of pBB1yqh2, XhoI site underlined
kiv1	TCCCCCGGGAGGAGACACAACATGTATACAGTAGGAG ATTACCTAT	Construction of pBB1 kiv1 yqh1, XmaI site underlined
kiv2	CCAATGCATTGGTTCTGCAGTTTTATGATTTATTTGTTT AGCAAAT	Construction of pBB1 kiv2 yqh1, PstI site underlined
bkdaa1	CTGGATCCCATTCAGACCTCCATGACC	Deletion of <i>bkdAA</i>
bkdaa2	CGGCCGCTTCAGAGCTCACATGAGATGAACGA CCACAAC	Deletion of <i>bkdAA</i>
bkdaa3	TGTTGTGGTCGTTTCATCTCATGTGAGCTCTG AAGCGGC	Deletion of <i>bkdAA</i>

(Continues)

TABLE 1 (Continued)

Strain, plasmid or oligonucleotide	Relevant characteristic(s) or sequence (5' → 3')	Source, reference or purpose
bkdaa4	GCTTGTCGACCCGTCGCTACTGCCGTAG	Deletion of <i>bkdAA</i>
bkdaagc1	GTACCGACGATGCCGCT	Verification of <i>bkdAA</i> deletion
bkdaagc2	GCCGTGCCACTAAGATGTAG	Verification of <i>bkdAA</i> deletion
stha1	GCCGCTTTGGTCCCGGATCCACAGCATCCAGTACGT CCGC	Deletion of <i>stha</i>
stha2	GTTGAAATCGGTCTCTCCGACCTGAACGCCGCGCACA TTAAC	Deletion of <i>stha</i>
stha3	GTAAATGTGCGCGGCGTTTCAGGTCGGAGAGACCGATTT CAAC	Deletion of <i>stha</i>
stha4	TTGCATGCCTGCAGGTCGACTGGTTGGGCAAACCCTGC TTGG	Deletion of <i>stha</i>
sthagc1	ATGGCTATTCGACGCTGCTG	Verification of <i>stha</i> deletion
sthagc2	ACTATGGCTGCGAACTGCTG	Verification of <i>stha</i> deletion
gcd1	GCCGCTTTGGTCCCGGATCCTGACCTTGAGTTGTTCC TTG	Deletion of <i>gcd</i>
gcd2	GACCTGACGGAGAACCTACATTAGCCGAGT AAGCGACAC	Deletion of <i>gcd</i>
gcd3	GTGTCGCTTACTCGGCTAATGTAGGTTCTCCGTCA GGTC	Deletion of <i>gcd</i>
gcd4	TTGCATGCCTGCAGGTCGACGACAACATCAGCAACG ACC	Deletion of <i>gcd</i>
gcdgc1	GGGATGGGTTTCAATGGTTC	Verification of <i>gcd</i> deletion
gcdgc2	GGCACAAGATGTTCTCAAGG	Verification of <i>gcd</i> deletion
png1	AGCTCTAAGGAGTTATAAAAACATATGTATACAGTAGG AGATTACC	Construction of pIP02, pIP03, pIP04 and pIP05
png2	GAGAATAGGAACTTCGAACTGCAGGTCGACTCAGAGG CCTCCAGC	Construction of pIP02, pIP03 and pIP04
png3	AGCTCTAAGGAGTTATAAAAACATATGTACTACTGTT GGAAATTATTTGTTA	Construction of pIP05
pbb1	TCGGAGCTCCGCGAATTGCAAGCTGATCCG	Construction of pIP01, SacI site underlined
pbb2	ATCGGATCCCTTAGCGGGCGGCTTCGTAT	Construction of pIP01, BamHI site underlined
kdca1	TTGCTAAACAAAATTCATAAACTGCAGAACCAATGC	Amplification of <i>kdca</i> gene
kdca2	AATGCATTGGTTCTGCAGTTTATGAATTTGTTTACG AAAGACTTTC	Amplification of <i>kdca</i> gene
p41	TTGCTAAACAAAATTCATAAACTGCAGAACCAATG CATTG	Construction of pEMP014
p42	TAATTTCCAACAGTGTACATGTTGTGTCTCCTCCCGG	Construction of pEMP014
p43	TCATTGATTTTACTAAATAAGCCAGGAGGACAGCTAT	Construction of pEMP012
p44	CGTACTACTGCTGCTTTCATGTTGTGTCTCCTCCAATGC	Construction of pEMP012
p45	GTGTGGCGATTTCGTTTCTAAGCCAGGAGGACAGCTA TGAC	Construction of pEMP013
p46	TGGGGTGCAGCAGTGGTCATGTTGTGTCTCCTCCAA TGCATTG	Construction of pEMP013
adha1	GCATTGGAGGAGACACAACATGAAAGCAGCAGTA GTACG	Amplification of <i>adha</i> gene from <i>L. lactis</i>
adha2	GTCATAGCTGTCTCCTGGCTTATTTAGTAAAATC AATGACCATCC	Amplification of <i>adha</i> gene from <i>L. lactis glutamicum</i>
adha3	TGCATTGGAGGAGACACAACATGACCACTGCTGCACC	Amplification of <i>adha</i> gene from <i>C. glutamicum</i>
adha4	GTCATAGCTGTCTCCTGGCTTAGAAACGAATCG CCACACG	Amplification of <i>adha</i> gene from <i>C. glutamicum</i>

(Continues)

TABLE 1 (Continued)

Strain, plasmid or oligonucleotide	Relevant characteristic(s) or sequence (5' → 3')	Source, reference or purpose
alss1	GAGGAAAGCGGCCGCTCTTCGGGGCGGAGCTTGTTG	Construction of pEMP04, NotI site underlined
alss2	TTAGATCTCGAGGCTCTTCGGGCCTAGAGAGCTTTTCG TTTTTCATG	Construction of pEMP04, XhoI site underlined
ilvc1	GAGGAAGCGGCCGCTCTTCGAAGAAAGTCGCCATCATC	Construction of pEMP04, NotI site underlined
ilvc2	TTAGATCTCGAGGCTCTTCGGGCTTAGTTCTTGGTC TTGTCGAC	Construction of pEMP04, XhoI site underlined,
ilvd1	GAGGAAGCGGCCGCTCTTCGCGGCGCCCGTG	Construction of pEMP04, NotI site underlined
ilvd2	TTAGATCTCGAGGCTCTTCGGGCTCAGAGGCCTTCCAG	Construction of pEMP04, XhoI site underlined
pip011	GAGGAAGCGGCCGCTCTTCGCGTGACTGGGAAAACCC TGGCGACTAGTCTTGGACTC	Construction of pEMP04, NotI site underlined
pip012	TTAGATCTCGAGGCTCTTCGGGCTTAGCGGGCGGCTTCG TATATACGGCGGCTGA	Construction of pEMP04, XhoI site underlined

### 2.3 | Recombinant DNA work

Standardized cloning procedures such as PCR and DNA restrictions were carried out according to Sambrook and Russell, 2001. Plasmids were isolated from 5 mL liquid cultures using the E.Z.N.A.<sup>®</sup> Plasmid Mini Kit (Omega Bio-tek, Inc., Norcross, USA) following manufacturer's instructions. PCR fragments were purified with the NucleoSpin<sup>®</sup> Gel and PCR Clean-up Kit (Macherey-Nagel GmbH & Co. KG, Düren, Germany) according to the manufacturer's instructions. Chromosomal DNA of *E. coli* MG1655, *P. putida*, *C. maltaromaticum*, and *L. lactis* was isolated using the Nucleospin<sup>®</sup> Microbial DNA Kit (Macherey-Nagel GmbH & Co. KG, Düren, Germany) following the protocol of the manufacturer. Electrocompetent cells were prepared for *E. coli* and *P. putida* as described previously [53,54]. *E. coli* DH5 $\alpha$  and *P. putida* strains were electroporated with an Eporator (Eppendorf AG, Hamburg, Germany) at 2.5 kV with 600  $\Omega$  resistance. All enzymes for recombinant DNA work were obtained from Thermo Fisher Scientific Inc. (Darmstadt, Germany) and oligonucleotides were synthesized by biomers.net GmbH (Ulm, Germany, listed in Table 3).

### 2.4 | Plasmid construction

*yqhD* was amplified from genomic DNA of *E. coli* MG1655 using the primers *yqhD1/yhqD2*, digested with PstI/XhoI and ligated into PstI/XhoI-digested pBB1 yielding pBB1 *yqhD*. *kivD* was subsequently added before *yqhD*, amplified from pSA55 with the primer pair *kivD1/kivD2*, digested with PstI/XmaI, and ligated into PstI/XmaI-digested pBB1

creating plasmid pBB1 *kivD yqhD*.  $P_{tac}$ , *kivD*, and *yqhD* were amplified from plasmid pBB1 *kivD yqhD* using the primers *pbb1/pbb2*. The resulting PCR fragment was digested with BamHI/SacI and subsequently ligated into BamHI/SacI-digested pSEVA231 to create plasmid pIP01. Plasmid pEMP04 was constructed using the inABLE DNA assembly method from Ingenza Ltd. The *B. subtilis alsS* and *P. putida ilvC* and *ilvD* genes were amplified using primer pairs *alss1/alss2*, *ilvc1/ilvc2*, and *ilvd1/ilvd2*, respectively. Additionally, a 5' truncated version of pIP01 was amplified using primer pair *pip011/pip012*. The PCR products were digested using SapI and annealed oligonucleotides were ligated at each terminus. Ligation of the oligonucleotides results in the generation of 5' and 3' 16 nt single stranded overhangs that are complementary between fragments resulting in the DNA assembling in the predefined order. The genes of pEMP04 were amplified using the primers *png1/png2* and cloned by Gibson Assembly [36] into NdeI/SalI-digested pNG413.1 yielding plasmid pIP02. *kdcA* from *C. maltaromaticum* LMA28, *adhA* from *L. lactis* MG1363, and *adhA* from *C. glutamicum* was amplified using the respective genomic DNA with the primers *kdca1/kdca2*, *adha1/adha2*, and *adha3/adha4* and cloned together via Gibson Assembly with a PCR fragment from pEMP04 that was amplified with the primers *p41/p42*, *p43/p44*, or *p45/p46* to construct plasmid pEMP014, pEMP012, and pEMP013. To exchange  $P_{tac}$  with  $P_{BAD}$  the genes of pEMP012, pEMP013 and pEMP014 were amplified using the primers *png1/png2* for pEMP012/013 and *png3/png2* for pEMP014 and cloned by Gibson Assembly into NdeI/SalI-digested pNG413.1, constructing the plasmids pIP03, pIP04, and pIP05.

## 2.5 | Determination of $\mu$ and $Y_{X/S}$

Growth rates were determined by linear regression of  $\ln(\text{OD}_{600})$  plotted against time (in hours) during the exponential growth phase. Biomass yields  $Y_{X/S}$  (g/g) were calculated by linear regression of the biomass concentration  $c_x$  (g/L) plotted against the respective glucose concentration (g/L) during the exponential growth phase.

## 2.6 | Construction of *P. putida* deletion mutants

Chromosomal deletions in *P. putida* were carried out using the 5-fluorouracil (5-FU)/*upp* counterselection system [37]. Deletions of the *bkdAA* gene (encoding the  $\alpha$ -subunit of the ketoacid dehydrogenase complex), the *sthA* gene (encoding soluble transhydrogenase) and the *gcd* gene (encoding glucose dehydrogenase) were performed using the integration vector pJOE6261.2. The flanking regions (about 500 bp) of each gene were amplified by PCR from chromosomal DNA of *P. putida* using the primer pairs *bkdaa1/bkdaa2* and *bkdaa3/bkdaa4*, *stha1/stha2*, and *stha3/stha4*, *gcd1/gcd2* and *gcd3/gcd4*. The two respective PCR fragments were purified and cloned into *Sall*/*Bam*HI-restricted pJOE6261.2 by Gibson Assembly. Finally, the assembly mix was used to transform *P. putida* by electroporation. The first selection was carried out on LB agar with 50  $\mu\text{g/L}$  kanamycin and a kanamycin-resistant clone was afterward grown in liquid LB medium for 24 h. The second recombination event was induced by plating cells on LB agar with 50  $\mu\text{g/L}$  5-FU. Deletion mutants were identified by colony PCR using the primer pairs *bkdaagc1/bkdaagc2*, *sthagc1/sthagc2*, and *gcdgc1/gcdgc2*, respectively.

## 2.7 | Analytics

Biomass formation was measured by determination of the  $\text{OD}_{600}$  (Ultraspex 10, GE Healthcare, USA) at specific time points. The cell dry weight ( $\text{g}_{\text{CDW}}/\text{L}$ ) was correlated to the  $\text{OD}_{600}$  in several independent cultivations with a correlation factor of 0.346  $\text{g}_{\text{CDW}}/\text{L}$  per OD (data not shown). Shaking flasks were sampled directly in the incubator using an injection syringe (100 Sterican<sup>®</sup>, 0.80  $\times$  120 mm, B. Braun, Melsungen, Germany). For the determination of isobutanol, 2-KIV, 2-ketogluconate (2-KG), and glucose concentrations, 2 mL of the main culture was harvested by centrifugation (12 100  $\times g$ , 5 min, room temperature (RT)) and the supernatant was analyzed via HPLC. Glucose concentrations were measured enzymatically with a test kit from r-biopharm (r-biopharm AG, Darmstadt, Germany).

## 2.8 | HPLC metabolite quantification

Isobutanol, 2-KIV and 2-KG were measured with a Agilent 1200 series HPLC system equipped with a Rezex ROA

organic acid H (8%) column (300 by 7.8 mm, 8  $\mu\text{m}$ ; Phenomenex) protected by a Phenomenex guard column carbo-H (4 by 3.0 mm inside diameter) [38]. Samples and standards were treated with a phosphate precipitation protocol before HPLC measurements. More precisely, 500  $\mu\text{L}$  of sample volume was mixed with 45  $\mu\text{L}$  4 M  $\text{NH}_3$  and 50  $\mu\text{L}$  1.2 M  $\text{MgSO}_4$  followed by 5 min incubation at RT and centrifugation for 5 min at 7000  $\times g$ . Pellets were discarded and the supernatant was mixed with 500  $\mu\text{L}$  0.1 M  $\text{H}_2\text{SO}_4$ , incubated for 15 min at RT, and centrifuged for 15 min at 7000  $\times g$ . The resulting supernatant was used for HPLC injection with an injection volume of 10  $\mu\text{L}$ . Separation was carried out under isocratic conditions at 50°C column temperature for 60 min with 5 mM  $\text{H}_2\text{SO}_4$  as the mobile phase at a constant flow rate of 0.4 mL/min. Detection of isobutanol, 2-KIV, and 2-KG was achieved with a refractive index detector at 32°C. Quantification of all analytes was done with a 7-point calibration curve for each component as an external reference standard.

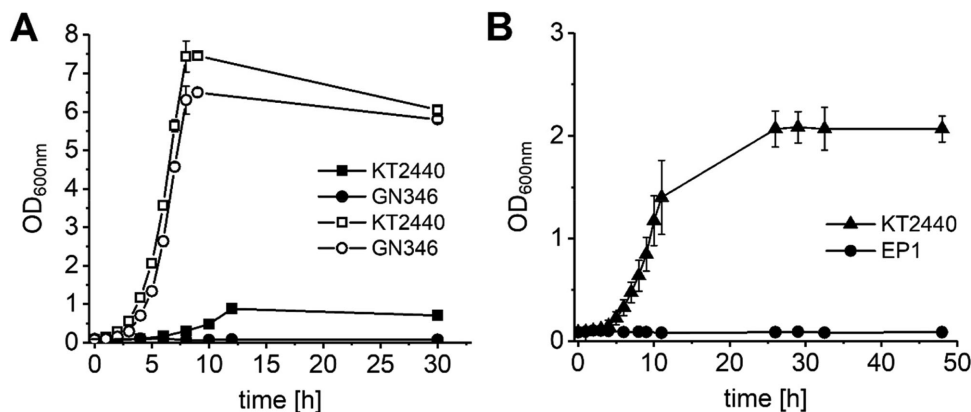
## 3 | RESULTS

### 3.1 | Preventing product and precursor degradation

*Pseudomonads* are well-known for their ability to degrade a variety of organic substances to utilize them as carbon and energy sources [21]. Since the genomic repertoire provides annotated routes for the degradation of isobutanol and 2-ketoisovalerate (Figure 1), we initially characterized growth on both compounds (Figure 2). *P. putida* showed exponential growth on isobutanol with a  $\mu$  of  $0.27 \pm 0.01 \text{ h}^{-1}$  as well as on 2-KIV with a  $\mu$  of  $0.33 \pm 0.01 \text{ h}^{-1}$  that is 52% of the growth rate on glucose (Figure 2A,B). Recently, several enzymes involved in *n*-butanol degradation were identified [39] and Simon et al. [30] constructed *P. putida*  $\Delta upp \Delta pedE \Delta pedI \Delta pedH \Delta aldB-I$  (*P. putida* GN346) to inactivate two alcohol dehydrogenases (PedE, PedH) and two aldehyde dehydrogenases (PedI, AldB-I) and showed that the introduced deletions prevented *n*-butanol consumption. Accordingly, *P. putida* GN346 was unable to utilize isobutanol as sole carbon and energy source (Figure 2A).

*P. putida* possesses a branched chain ketoacid dehydrogenase (BCKDH) complex that converts 2-ketoacids to the respective decarboxylated CoA-derivatives [40,41] which are, after further conversion steps, funneled into the TCA cycle. To prevent the consumption of the precursor 2-KIV, we inactivated the  $\alpha$ -subunit of the BCKDH by deletion of the *bkdAA* gene in *P. putida* GN346. In contrast to the wild-type, the resulting strain *P. putida* EP1 was unable to grow on 2-KIV as carbon source (Fig. 2B), and therefore was used as basis for further strain engineering.





**FIGURE 2** (A) Growth of *P. putida* KT2440 and *P. putida* GN346 in DeBont minimal medium containing 0.5 g/L isobutanol (filled symbols) or 5.4 g/L glucose (open symbols). (B) Growth of *P. putida* and *P. putida* EP1 in DeBont minimal medium containing 2.9 g/L 2-ketoisovalerate. Experiments were performed in triplicates and error bars represent the corresponding standard deviation

**TABLE 2** Overview of growth, 2-ketogluconate (2-KG) and isobutanol production of *P. putida* and its engineered derivatives

Strain	$\mu$ [ $\text{h}^{-1}$ ]	$Y_{X/S}$ [g/g]	$Y_{2\text{-KG/S}}$ [mg/g <sub>GLC</sub> ]	$Y_{\text{Iso/S}}$ [mg/g <sub>GLC</sub> ]
KT2440	$0.62 \pm 0.01$	$0.40 \pm 0.01$	0	0
GN346	$0.59 \pm 0.01$	$0.39 \pm 0.01$	0	0
Iso1	$0.56 \pm 0.02$	$0.30 \pm 0.01$	0	0
Iso2	$0.25 \pm 0.01$	$0.13 \pm 0.01$	$438 \pm 20$	$22 \pm 2$
Iso3	$0.14 \pm 0.01$	$0.06 \pm 0.01$	$833 \pm 100$	$13 \pm 1$
Iso4	$0.19 \pm 0.01$	$0.09 \pm 0.02$	$771 \pm 15$	$14 \pm 0.0$
Iso5	$0.18 \pm 0.02$	$0.28 \pm 0.01$	0	0
Iso6	$0.28 \pm 0.01$	$0.10 \pm 0.01$	$633 \pm 39$	$21 \pm 1$

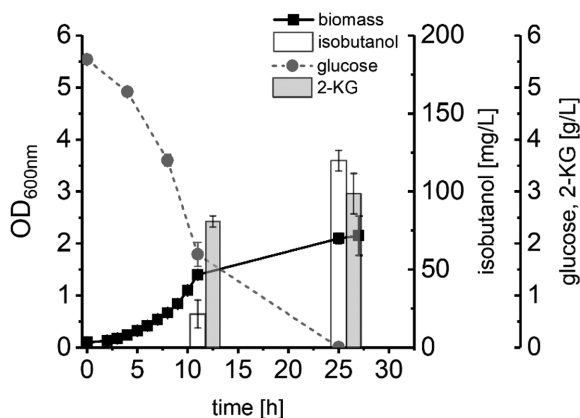
### 3.2 | Engineering *P. putida* for isobutanol production

To drain the carbon from pyruvate to 2-KIV, we constructed a plasmid harboring the *alsS* gene encoding the acetolactate synthase from *Bacillus subtilis*, which is not feedback inhibited by branched chain amino acids, and the native *ilvCD* genes encoding the ketolacid reductoisomerase and dihydroxyacid dehydratase (Figure 1). For the conversion of 2-KIV to isobutanol, we additionally cloned *kivD* encoding the KDC from *Lactococcus lactis* and *yqhD* encoding an aldehyde reductase from *E. coli* (Fig. 1). *AlsS*, *KivD*, and *YqhD* were previously applied for isobutanol production in other hosts such as *C. glutamicum* and *E. coli* [5,7]. The resulting plasmid pIP02 expresses all cloned genes under control of the L-arabinose inducible  $P_{BAD}$  promoter and was used to transform *P. putida* EP1 yielding *P. putida* Iso1. In minimal medium with glucose, *P. putida* Iso1 showed a  $\mu = 0.56 \pm 0.02$ . Although the  $Y_{X/S}$  was reduced by 25% compared to the wild-type, no isobutanol was produced during the cultivation (Table 2).

The synthesis of isobutanol from glucose requires 2 mol NAD(P)H per mol isobutanol. The reduction of acetolactate is catalyzed by NADPH-dependent ketolacid

reductoisomerase (*IlvC*), while the conversion of isobutyraldehyde to isobutanol can be catalyzed by NAD(P)H-dependent aldehyde/alcohol dehydrogenases such as *YqhD* (Figure 1). Since *YqhD* is NADPH-dependent, the engineered isobutanol pathway should consume 2 mol NADPH per mol isobutanol. *P. putida* possesses a membrane-bound and a soluble transhydrogenase. The latter is encoded by the *sthA* gene [42] and has in *E. coli* been reported to favor the re-oxidation of NADPH to  $\text{NADP}^+$  under reduction of  $\text{NAD}^+$  to NADH [43,44].

To test whether the inactivation of the soluble transhydrogenase is beneficial for isobutanol production, we deleted the *sthA* gene in *P. putida* EP1, yielding *P. putida* EP2 which was transformed with the plasmid pIP02. The resulting strain *P. putida* Iso2 showed in minimal medium containing 5.4 g/L glucose, a growth rate of  $0.26 \pm 0.01 \text{ h}^{-1}$ , a  $Y_{X/S}$  of  $0.13 \pm 0.01 \text{ g/g}$ , and produced  $438 \pm 20 \text{ mg/g}_{\text{GLC}}$  2-KG and for the first time isobutanol with a  $Y_{\text{Iso/S}}$  of  $22 \pm 2 \text{ mg/g}_{\text{GLC}}$  (Table 2, Figure 3). We also replaced the aldehyde reductase gene *yqhD* on the overexpression plasmid pIP02 with the *adhA* genes encoding NADH-dependent alcohol dehydrogenase variants from *Lactococcus lactis* and *Corynebacterium glutamicum*, respectively. The plasmids pIP03 and pIP04 were used to transform *P. putida* EP2, yielding *P. putida* Iso3 and Iso4,



**FIGURE 3** Growth (black circles), glucose consumption (grey circles), isobutanol production (white bars) and 2-KG formation (grey bars) of *P. putida* Iso2 in DeBont minimal medium containing glucose. Experiments were performed in triplicates and error bars represent the corresponding standard deviation

which were characterized in minimal medium with glucose (Table 2). Both strains showed reduced growth rates and about 40% lower product yields compared to *P. putida* Iso2. All engineered strains with deletion of *sthA* converted 40% to 83% of the available glucose into 2-KG that was secreted into the culture broth (Table 2). To avoid 2-KG secretion and to improve isobutanol production, we constructed *P. putida* EP3 by deletion of the *gcd* gene encoding periplasmic glucose dehydrogenase in *P. putida* EP2. To construct *P. putida* Iso5, *P. putida* EP3 was transformed with the plasmid pIP02. In fact, *P. putida* Iso5 did not secrete any 2-KG, however, inactivation of GCD also abolished isobutanol production completely (Table 2).

### 3.3 | Ketoacid decarboxylase from *Carnobacterium maltaromaticum* is suitable for isobutanol production

The key enzyme for isobutanol production via the Ehrlich pathway is ketoacid decarboxylase (KDC) converting 2-KIV to isobutyraldehyde (Figure 1). So far, only KDC from *L. lactis* has been proven as suitable variant that efficiently catalyzes this reaction [2,5]. Recently, the genome sequence of *Carnobacterium maltaromaticum* LMA28 [28] was published that harbors a gene encoding a putative KDC. KDC from *C. maltaromaticum* shows 54% identity to the KDC enzyme from *L. lactis*. To test the suitability of KDC from *C. maltaromaticum* for isobutanol production, we replaced the *kivD* gene on plasmid pIP02 with the respective gene from *C. maltaromaticum*. Plasmid pIP05 was transformed into *P. putida* EP2 and the resulting strain *P. putida* Iso6 was characterized. *P. putida* Iso6 showed a growth rate of  $0.28 \pm 0.01 \text{ h}^{-1}$ , a  $Y_{X/S}$  of  $0.10 \pm 0.01 \text{ g/g}$ , and produced  $633 \pm 39 \text{ mg/g}_{\text{GLC}}$  2-KG. Furthermore, *P. putida* Iso6 secreted

**TABLE 3** Overview of engineered *P. putida* strains cultivated under oxygen deprivation conditions

Strain	$Y_{\text{Iso/S}}$ [mg/g <sub>GLC</sub> ]	$q_s$ [g g <sup>-1</sup> h <sup>-1</sup> ]	$Y_{2\text{-KG/S}}$ [mg/g <sub>GLC</sub> ]
KT2440	0	$0.11 \pm 0.01$	$42 \pm 32$
Iso2	$9 \pm 1$	$0.14 \pm 0.01$	$120 \pm 9$
Iso3	$5 \pm 2$	$0.12 \pm 0.01$	$183 \pm 8$
Iso4	$4 \pm 1$	$0.13 \pm 0.01$	$193 \pm 12$
Iso5	0	$0.01 \pm 0.00$	0
Iso6	$19 \pm 2$	$0.07 \pm 0.01$	$397 \pm 14$

as much isobutanol as *P. putida* Iso2 with a  $Y_{\text{Iso/S}}$  of  $21 \pm 1 \text{ mg/g}_{\text{GLC}}$  (Table 2) showing that KDC from *C. maltaromaticum* LMA28 is a useful alternative to KDC from *L. lactis*.

### 3.4 | Microaerobic isobutanol production in *P. putida*

*P. putida* is regarded as an obligate aerobic bacterium [18]. However, since the implementation of the synthetic isobutanol pathway theoretically enables a closed redox balance, we tested the capabilities of our engineered *P. putida* strains to produce isobutanol from glucose in a zero-growth bioprocess under oxygen deprivation conditions [45]. Therefore, we inoculated *P. putida* WT and Iso2–6 to an OD<sub>600</sub> of 15–20 in closed bottles filled with minimal medium containing 5.4 g/L glucose and characterized substrate consumption and (by-) product formation (Table 3). In the micro-aerobic environment *P. putida* WT showed no growth, but remained metabolically active and consumed the glucose that was converted to 2-KG. With the exception of *P. putida* Iso5, all other engineered strains consumed glucose and produced isobutanol. *P. putida* Iso6 showed the best performance under oxygen deprivation conditions. Compared to the WT the  $q_s$  was reduced by 37% and *P. putida* Iso6 produced about 10% less isobutanol compared to the aerobic shaking flask experiments (Table 3).

## 4 | DISCUSSION

*P. putida* is an emerging host for industrial biotechnology [46–48]. However, this bacterium is also known to efficiently metabolize a broad range of substrates including amino and organic acids and alcohols [21]. As shown here, *P. putida* grows rapidly on isobutanol as well as on its precursor 2-KIV. Although *P. putida* KT2440 possesses four aldehyde dehydrogenases and about 10 alcohol dehydrogenases, Simon et al. [30] showed that deletion of the two alcohol dehydrogenase genes *pedE* and *pedH* and the two aldehyde dehydrogenases genes *pedI* and *aldB-I* is sufficient to prevent *n*-butanol degradation. Accordingly, we found that this strain background also prevents growth on

the branched-chain alcohol isobutanol. *P. putida* possesses a branched chain ketoacid dehydrogenase complex that converts 2-ketoacids to the respective decarboxylated CoA-derivatives [40,41]. As expected and also observed for *P. taiwanensis* VLB120 [41], inactivation of the BCKDH abolished growth on 2-ketoisovalerate. To avoid auxotrophies, we relinquished the inactivation of the L-valine forming transaminase IlvE, the 2-isopropylmalate synthase LeuA and the 2-ketoisovalerate hydroxymethyltransferase PanB as has been applied to improve isobutyric acid production with *P. taiwanensis* strain VLB120 [41].

Since AHAIIR is usually NADPH-dependent, the synthesis of one molecule of isobutanol either requires two molecules of NADPH or one NADH plus one NADPH molecule depending on the applied alcohol/aldehyde dehydrogenase variant for the reduction of isobutyraldehyde to isobutanol. Optimization of NAD(P)H availability has already been shown to be a crucial factor for isobutanol production with other hosts such as *E. coli* and *C. glutamicum* [6,49]. Recently, Nikel et al. [16] showed that *P. putida* cells growing on glucose exhibit a slight catabolic overproduction of reducing power and run a biochemical cycle that favors NADPH formation. Therefore, we applied in our experiments the broad-substrate range NADPH-dependent aldehyde reductase YqhD [50], which has also been successfully applied for isobutanol production with *E. coli* [8]. However, expression of the synthetic pathway in *P. putida* Iso1 to channel pyruvate toward isobutanol did not result in isobutanol production from glucose. Similar to *E. coli*, *P. putida* possesses a membrane bound (PntAB) and a soluble transhydrogenase (SthA) to balance the overall redox state of the cell (Figure 1). SthA has in *E. coli* been reported to favor the oxidation of NADPH to NADP<sup>+</sup>, accompanied with the reduction of NAD<sup>+</sup> to NADH [43,44]. To improve NADPH availability, we inactivated SthA that resulted in isobutanol formation in *P. putida* Iso2 under aerobic conditions. Accordingly, expression of two *adhA* genes encoding NADH-dependent alcohol dehydrogenases from *L. lactis* and *C. glutamicum*, which have previously been shown to be suitable for isobutanol production [7,8], instead of YqhD, led to significantly reduced isobutanol yields in the  $\Delta$ *sthA* background (Table 2).

Inactivation of SthA resulted in isobutanol production, however, also in the secretion of significant amounts of 2-KG. In *P. putida* a majority of the glucose is converted in the periplasm by glucose dehydrogenase (Gcd) to gluconate, which is transported to the cytoplasm and activated by the gluconate kinase to feed the Entner–Doudoroff pathway with 6-phosphogluconate. Usually, only a small fraction of gluconate is converted in the periplasm by gluconate dehydrogenase to 2-KG, which is subsequently transported into the cytoplasm to finally form 6-phosphogluconate via 2-KG kinase and 2-ketogluconate-6-P reductase [16]. Since deletion of *gcd* abolished 2-KG production completely, the synthesis of this

molecule occurs solely in the periplasm via the described route. The accumulation of 2-KG in the culture broth indicates a transport inhibition of gluconate and/or 2-KG from the periplasm to the cytoplasm by an unknown mechanism and/or an inhibition or limitation of the ATP-dependent conversion to the phosphorylated derivatives. The latter might result as consequence of a perturbed redox state due to the inactivated transhydrogenase SthA.

*P. putida* is an obligate aerobic bacterium, however, in a bioelectrochemical system *P. putida* was metabolically active under anoxic conditions when an electron mediator was applied for redox balancing in a high-yield 2-KG production system [51,52]. Since isobutanol synthesis enables regeneration of NAD(P)<sup>+</sup>, we cultivated *P. putida* WT and the engineered derivatives under microaerobic conditions. All strains showed no growth (data not shown) but with the exception of *P. putida* Iso5, remained metabolically active and *P. putida* Iso2-4 and 6 also secreted 2-KG, isobutanol, and further unidentified products. However, according to the zero-growth the  $q_S$  values are low compared to aerobic conditions (e.g. for *P. putida* WT 0.11 vs. 1.55 g g<sup>-1</sup> h<sup>-1</sup>). The capability of *P. putida* to remain metabolically active opens the possibility to develop dual-phase production processes that comprise an aerobic growth phase for rapid biomass formation and a micro-aerobic or anaerobic production phase [45].

This study paths the way to construct more efficient *P. putida* strains for isobutanol production in future studies. The overall isobutanol yield is significantly higher compared to other engineered *P. putida* strains [41], however, rather low compared to tailored *E. coli* and *C. glutamicum* strains [49]. Product and precursor degradation can be prevented by the presented deletions in this study, however, improving NAD(P)H and pyruvate availability [49] will be crucial to achieve high-yield isobutanol production strains.

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## CONFLICT OF INTEREST

The authors have declared no conflicts of interest.

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