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### 1 Basis for a new process for producing REE oxides from *Dicranopteris linearis*

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13

#### 14 Abstract

15

16 Dicranopteris linearis (also known as Dicranopteris dichotoma) is a fern that grows 17 naturally on the tailings of former rare earth element (REE) mines in southern China. It 18 accumulates these elements in its aerial parts at concentrations that classify this plant 19 as hyperaccumulator. The main objective of this research is to design a process to 20 recover REEs directly from the dry biomass, without burning it. This process involves 21 three steps: (i) extraction of REEs from the dry plant by leaching with a solution 22 containing acid and/or complexing agents, (ii) precipitation of REEs with oxalate and (iii) 23 calcination to prepare REE oxide. A methodology based on speciation calculation made 24 it possible to determine operating conditions for precipitation by taking into account the 25 influence of the species present according to the extraction method. Simulations were 26 carried out using Chess software, allowing the calculation of the composition of a 27 multicomponent solution at thermodynamic equilibrium. These operating conditions were 28 then validated experimentally. REE extraction from the dry plant was significantly 29 improved under acid conditions (with  $H_2SO_4$ ) or with a strong complexing agent such as 30 ethylenediaminetetraacetic acid (H<sub>4</sub>EDTA). After optimization of the precipitation 31 conditions, REE precipitation yields were above 90%. After calcination, the final solid 32 (REE oxide) contained ca 69% of the REEs present in the initial biomass. The use of 33 sulphuric acid as an extracting agent resulted in the recovery of a solid composed of 52 34 wt % of REEs (based on metal content). Thus, this study proposes a new recovery route 35 for these strategic elements with a REE concentration factor from soil to final solid of 36 2500.

37

#### 38 Keywords

39 Agromining; hyperaccumulator plant; hydrometallurgy; leaching; EDTA; REE speciation

40

#### 41 **1. Introduction**

42 Rare earth elements (REEs), including the 15 lanthanides as well as yttrium and 43 scandium, share similar chemical properties. In recent years, they have been widely 44 used in functional materials (especially in green and electronic technologies), as 45 catalysts or as diagnostic reagents for magnetic resonance imaging, etc. [1]. The 46 disparity of their resource around the world requires increasing extraction yields from 47 primary ores, but also increasing their recovery from secondary materials (e.g. 48 phosphogypsum from phosphoric acid production or red mud from bauxite) [2, 3]. 49 Among these secondary resources, low grade soils and tailings from old mines could be 50 suitable for REE recovery. However, new concentration processes have to be developed 51 to be economically and environmentally competitive. Agromining represents a promising 52 means of achieving this goal.

Agromining is an emerging phytotechnology, which uses natural and local hyperaccumulating plants to "harvest" metals [4]. The plants, which accumulate metals in their aerial parts at very high levels, are already being used to produce nickel from ultramafic soils, which are not concentrated enough for conventional mining industry [5, 6]. The extraction yield is over 100 kg of nickel per hectare in Mediterranean climate (with *Alyssum murale* [7] and is very promising in tropical areas (with *Phyllanthus rufuschaneyi*) [8]. The metal is then generally recovered by hydrometallurgical 60 processes from biomass ashes after burning [6]. The combustion stage removes organic 61 matter and concentrates the metal. It also generates energy, which can be wittingly 62 used. The direct extraction of the metal from the plants, without burning, has also been 63 reported [9, 10]. The main advantages of this route are to limit the issue of gas treatment 64 after metal contamination, and to use biomass after metal extraction for other 65 applications (e.g. composting, anaerobic digestion, etc.). However, part of the soluble 66 organic matter is co-extracted, which implies the development of specific processes to 67 recover the metal [10].

The general objective of this work is to develop the agromining of REEs. Several hyperaccumulating plants have been identified worldwide [11], the concentration threshold being set at 0.1wt% of the dry biomass. One of them, *Dicranopteris linearis* (also known as *Dicranopteris dichotoma*) is a fern growing in southeastern China (subtropical climate). It is found naturally on the tailings of former mines (ionic adsorption clay deposits), at a plant density that makes large-scale cultivation possible.

74 While the interactions soil nutrients/D. linearis and metal translocation in the fern are 75 fairly well documented [12-14], only one process for the extraction and purification of 76 REEs has been reported. Recently, it has been demonstrated that the recovery of REE 77 mixtures is possible by direct extraction, without burning the plant [15]. The recovery 78 step consists of nitric acid extraction followed by adsorption of the elements on an ion 79 exchange resin. Selective elution with nitric acid solutions of several concentrations 80 allows recovery of aqueous REEs with a yield of ca 75% and a purity close to 80%. The 81 main impurity is aluminium (AI), which the fern contains in large quantities; actually, this 82 fern is also an hyperaccumulator of Al. Other strategies should therefore be applied to avoid this contamination, and among them, the precipitation of REE-oxalate ( $C_2O_4^2$ -) 83 84 seems to be relevant. These intermediates are often studied to produce concentrated 85 solids (REE-oxides) after calcination [16]. One of the main advantages of this strategy is 86 the high efficiency of selective REE precipitation. In addition, solids after combustion are 87 relatively pure, due to the transformation of oxalate with few residues (a molecule 88 composed only of carbon and oxygen).

89 The aim of this research is therefore to synthetize REE-oxides after direct extraction of 90 REE from the hyperaccumulator D. linearis and precipitation as oxalate. It consists of 91 three steps: (i) a leaching step using extracting agents (organic or inorganic) followed by 92 (ii) a selective precipitation step with oxalate, and (iii) calcination of the intermediate (Fig. 93 1). Considering that REEs are complexed by carboxylic acids in the plant tissues, the 94 strategy is either to break these complexes using an acid solution or to shift the 95 complexing equilibrium using a more powerful complexing agent than the one present in 96 the plant. Sulphuric and citric acids have been selected for their acidic properties, and 97 H<sub>4</sub>EDTA and citrate for their complexing properties. H<sub>4</sub>EDTA is a well-known complexing 98 agent for REEs [17,18], as is citrate, which naturally transports REEs in plants through 99 the xylem [13]. During precipitation with oxalate, competitive reactions must be taken 100 into account. They are due to the presence of extracting agents introduced in the 101 previous step, and to soluble organic matter (SOM) of the plant, which may interfere. No 102 data are available on the precipitation of REEs by oxalic acid  $C_2H_2O_4$  in the presence of 103 high concentrations of sulphate, H<sub>4</sub>EDTA and citrate. The proposed methodology relies 104 on post-extraction stage speciation studies to investigate the feasibility of downstream 105 unit operations. Hence to determine the optimal precipitation conditions (pH and molar 106 ratio), simulations of REE speciation were first performed. In order to validate the model, 107 an experiment was prepared with a synthetic solution of lanthanum (La) (chosen as 108 representative REEs). The simulations were performed under the assumption that SOM 109 plays no role during precipitation. Tests with a real extract allow us to evaluate its role 110 during REEs recovery. The whole process is finally evaluated according to the 111 composition of the REE oxide obtained after calcination.



- 112
- 113Fig. 1: Flowsheet of the proposed process to recover REEs from *D. linearis*
- 114

#### 115 2. Materials and methods

#### 116 2.1 Plant material

Samples of *Dicranopteris linearis* were collected in a former REE mine in Jiangxi Province (China), and dried for 24 h, at 70 °C. The concentration of REEs in the soil was in the range 300 to 700 mg per kg of dry soil [19]. The entire biomass (leaves and stem) was crushed with a blade grinder (SEB type 8100) before use. The median particle size, determined by laser diffraction (Malvern Mastersizer hydro 2000SM), was 369 µm.

#### 122 2.2 Chemicals

123 Nitric acid (67-69%, Sigma Aldrich) was used for digestion prior to elemental analysis. 124 Dehydrated disodium salt of ethylenediaminetetraacetic acid (99%), ACS reagent citric 125 acid monohydrate ( $\geq$  99.0%) and L(+)-glutamic acid monosodium salt monohydrate 126 (99%) (supplied by Fisher Chemicals) and sulfuric acid (95-98%, Sigma Aldrich) were 127 tested in the leaching step. Oxalic acid dihydrate ACS ( $\geq$  99.5%, Sigma Aldrich) was 128 used for the precipitation step, and synthetic solutions were prepared with lanthanum nitrate hexahydrate (> 98%, ACROS Organics). Deionized (DI) water (Elga - Purelab
Option) was used for all experiments.

#### 131 2.3 Elemental and structural analysis

132 Prior to elemental analysis, the solid and liquid samples were digested in a microwave 133 oven (UltraWave, Milestone) for 40 min with a set-point of 200 °C. Biomass samples or 134 precipitates (ca 0.1 g) were randomly collected and digested by adding 5 mL of 135 concentrated nitric acid (5 repetitions each time). For liquids, 2 mL of nitric acid was 136 added to 2 mL of sample. After digestion, the leachate was introduced into a 50 mL flask 137 with deionized water, then filtered by syringes equipped with filters (Phenomenex, pore 138 size: 0.45 µm) and analysed by ICP-AES (Thermo ICAP6000). Calibration was 139 performed with the custom standard PlasmaCAL (SCP Science) in a 5% vol. HNO<sub>3</sub> 140 matrix. Two ranges of standard solutions were used, one for the REEs and one for the 141 other elements. The major elements were analysed by radial measurement while the 142 minor elements were analysed by axial measurement. Three replicates were made for 143 each sample and a quality test was performed every 10 measurements with a control 144 solution (PlasmaCAL, SCP Science).

X-ray diffraction analyses (XRD) were performed with a D8-Discover diffractometer
(Brucker) (cobalt probe). The crystals were identified with PDF-02 databank of ICDD
(International Centre for Diffraction Data). EDX (Energy dispersive X-ray spectrometry)
analyses were performed with a JSM-6490LV microscope (Jeol).

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#### 150 2.4 **REE extraction and recovery from biomass**

151 To study the effect of each agent on the efficiency of REE extraction, the fern biomass 152 was leached with six aqueous solutions (solid/liquid ratios S/L = 3 g/100 mL, 2 h, 153 magnetic stirring): deionized water,  $H_2SO_4$  (0.25 M), citric acid (1 M) (pH = 1.6 and 4.0, 154 adjusted with NaOH), citrate (1M) (pH = 8.0, adjusted with NaOH) and EDTA (0.05 M) 155 the following EDTA written in italic refer (in text, to the generic 156 ethylenediaminetetraacetic acid, whatever its chemical form). The latter complexing 157 agent was added in the disodic form Na<sub>2</sub>H<sub>2</sub>EDTA (to obtain an extraction at pH 4.5 (predominance area of H<sub>2</sub>EDTA<sup>2-</sup>). This agent can create a complex with REE, without 158

generating insoluble REE hydroxides (at pH higher than 7-8). Solid particles were
separated from solutions using 0.45 µm porosity cellulose filter papers (Whatman).

161 REE speciation was investigated and simulated at 25 °C using CHESS (Chemical 162 Equilibrium with Species and Surfaces, version 5.0.8) software [20]. The stability 163 constants of La with *EDTA* and oxalate were implemented in the default database EQ3/6 164 (already containing data on La and sulphate interactions) [21].

165 A synthetic solution (*ca* 300 mL) was prepared, containing lanthanum (La(NO<sub>3</sub>)<sub>3</sub>, 2.3  $10^{-3}$ 166 mol) and Na<sub>2</sub>H<sub>2</sub>EDTA (4  $10^{-3}$  mol). The pH adjustment (between 2 and 8.5) was carried 167 out with 1 M HNO<sub>3</sub> solution and oxalic acid was added in a molar ratio of 1 with 168 Na<sub>2</sub>H<sub>2</sub>EDTA (4  $10^{-3}$  mol). The final pH was measured. After one hour of mixing, 169 precipitates were filtered and dried. The liquid phases were analysed by ICP.

After simulation and determination of optimal precipitation conditions, pH was adjusted to 2.6 (with 1 M NaOH solution) and to 2.3 (with 1 M HNO<sub>3</sub> solution) for H<sub>2</sub>SO<sub>4</sub> and Na<sub>2</sub> H<sub>2</sub>EDTA extracts respectively. In the latter case, this caused the formation of a solid S1, which was separated and analysed (ICP and XRD). Oxalic acid was then added (molar ratio of 0.16 and 1.56, respectively) and the solutions were stirred during 1 h at 20 °C. Precipitates (S2 and S3 respectively) were filtered, washed with deionized water and dried at 100 °C. They were calcined at 700 °C for 2 h to obtain S4 and S5.

177

#### 178 **3. Theory**

179 The addition of oxalic acid to a solution containing REEs and *EDTA* will affect the 180 speciation

181 . The distribution of the chemical forms of the three species as a function of pH is
182 exhibited in Fig. 2. In this environment, many competitive reactions can take place,
183 including those represented by Equations 1-3.

- 184
- 185



- Deprotonation of oxalate:  $HOx^{-} \xrightarrow{1}_{2} Ox^{2-} + H^{+} Eq. 3$
- 189
- 190

191 Ideally, to promote precipitation, the REE<sup>3+</sup> and  $Ox^{2-} (C_2O_4^{2-})$  forms should be preferred 192 to obtain the insoluble complex REE<sub>2</sub>Ox<sub>3</sub>. However, in the presence of *EDTA*, the REEs 193 are theoretically complexed at pH higher than 1.9 (Fig. 2). Two other aspects must also 194 be taken into account:

- 195 *(i)* the very low solubility of the precipitate  $REE_2Ox_3$  (Ks between 5  $10^{-30}$  and 4.5  $10^{-32}$  [22]). This implies that the presence of a very small quantity of deprotonated 197 oxalic acid can lead to the formation of the precipitate. The latter inevitably leads 198 to the displacement of reaction 3 in the forward direction;
- (*ii*) only stability constants between EDTA<sup>4-</sup> and HEDTA<sup>3-</sup> are available. However,
   due to the *EDTA* excess introduced during extractions, REEs are complexed in
   totality with these two forms below pH 2.

Only modelling that takes all these data into account can enable optimum precipitation conditions to be found. Due to the large number of species involved, the Chess software was used [20], after having completed its database with the stability constants given in Table 1. Conventionally, the stability constant of the formation reaction of a species (Eq. 4) is given by Eq. 5. The species present on the left side of the equilibrium, called
"basis species", are usually written in their deprotonated form.

208

$$xA^{a+} + yB^{b-} \rightleftharpoons A_x B_y^{(ax-by)}$$
 Eq. 4

$$K = \frac{\left[A_{x}B_{y}^{(ax-by)}\right]}{[A^{a+}]^{x} \cdot [B^{b-}]^{y}}$$
Eq. 5

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210

211

#### Table 1: Stability constants used for speciation determination

Basis species		Species formed	Log K (25 °C)*
Ox <sup>2-</sup> + H <sup>+</sup>	$\Rightarrow$	HOx	4.27
Ox <sup>2-</sup> + 2 H <sup>+</sup>	$\rightleftharpoons$	H <sub>2</sub> Ox	5.52
Ox <sup>2-</sup> + La <sup>3+</sup>	$\rightleftharpoons$	LaOx+	6.00
2 Ox <sup>2-</sup> + La <sup>3+</sup>	$\rightleftharpoons$	LaOx <sub>2</sub> <sup>-</sup>	9.55
3 Ox <sup>2-</sup> + La <sup>3+</sup>	⇒	LaOx <sub>3</sub> <sup>3-</sup>	11.59
3 Ox <sup>2-</sup> + 2 La <sup>3+</sup>	⇒	La <sub>2</sub> Ox <sub>3</sub>	28.23
$EDTA^{4-} + H^+$	⇒	HEDTA <sup>3-</sup>	10.95
$EDTA^{4-}+2H^{+}$	$\rightleftharpoons$	H <sub>2</sub> EDTA <sup>2-</sup>	17.22
$EDTA^{4-} + 3 H^{+}$	⇒	H <sub>3</sub> EDTA <sup>-</sup>	20.34
$EDTA^{4-} + 4 H^{+}$	⇒	H₄EDTA	22.56
$EDTA^{4-} + 5 H^{+}$	$\rightleftharpoons$	H₅EDTA⁺	24.06
EDTA <sup>4-</sup> + La <sup>3+</sup>	≓	LaEDTA	17.94
$EDTA^{4-} + H^+ + La^{3+}$	$\rightleftharpoons$	LaHEDTA	20.40



\*The data is corrected to zero ionic strength using Davies equation.

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#### 215 4. Results and discussion

#### 216 4.1 Elemental composition of Dicranopteris linearis

The results of the *D. linearis* elemental analysis are presented in Table 2. Like all plants, the fern is rich in K, Ca, Mg. However, it contains high levels of Si and Al, which is a common characteristic of this plant [23]. The threshold for Al hyperaccumulation is set at 220 0.1%, indicating that this plant can be considered as AI - hyperaccumulator. Concerning 221 REEs, La, Nd and Ce were the main elements identified. Pr, Sm, Y and Gd were also 222 present but at lower concentrations. The analysis shows that *D. linearis* can accumulate 223 more than 3 g of REEs per kg of dry biomass (0.30%). This means that this plant batch 224 can be considered as a REE hyperaccumulator (threshold fixed at 1 g kg<sup>-1</sup>).

225 In the literature, concentration in plant tissues are generally measured between 0.023 226 and 0.26% [15]. The plant studied is therefore very rich. It mainly accumulates light 227 REEs.

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- 229

Table 2: Composition in major cations and rare earths in *Dicranopteris linearis* (wt%) 230 (confidence ranges are determined with 3 replicates)

Element		Dry biomass
REEs	La	0.128 ± 0.010
	Ce	0.113 ± 0.013
	Nd	0.088 ± 0.014
	Pr	0.027 ± 0.007
	Sm	0.014 ± 0.002
	Y	0.012 ± 0.002
	Gd	0.007 ± 0.001
	Total	0.389 ± 0.045
	Si	1.65 ± 0.05
Other	K	0.254 ± 0.031
	AI	0.205 ± 0.043
elements	Ca	0.174 ± 0.036
	Mn	0.051 ± 0.012
	Mg	0.048 ± 0.007

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232

#### 233 4.2 Extraction of REEs from *D. linearis*

234 The extraction yield of REEs is defined by Eq. 6:

235 
$$Extraction yield of REEs = \frac{mass of REEs in the extract}{intial mass of REEs in the plant} x 100 Eq. 6$$

236 Fig. 3 compares the extraction yield of REEs (Eq. 6) from biomass with the six extracting solutions: deionized water (blank sample), H<sub>2</sub>SO<sub>4</sub> (0.25 M) (strong mineral acid, pH < 237 238 0.5), citric acid (1 M) (weak carboxylic acid) at two pHs (1.6 and 4), citrate (1 M) 239 (complexing agent) at pH 8, and EDTA (0.05 M) (strong chelating agent) at pH 4.5. 240 Experiments were carried out under optimal conditions because the biomass had been

finely ground (therefore the diffusion limitations were low), and the solid/liquid ratio waslow.





250 The results show, as a first observation, that the major REEs contained by the plant 251 behaved in a very similar way during their extraction. With water, an extraction yield 252 between 40 and 60% was obtained. Na<sub>2</sub>EDTA and H<sub>2</sub>SO<sub>4</sub> were the most efficient 253 extracting agents with an almost similar yield > 80%. Citric acid had less effect, but its 254 efficiency increased as the pH decreased. At pH 8, no difference with water was 255 recorded. The results indicate that REEs can be extracted with very good yields under 256 acidic conditions, or with a strong complexing agent. Indeed, the only acidity of the 257 ETDA solution cannot explain the results obtained with this molecule (better extraction 258 yields at high pH compared to citric acid).

259 The stability constants of REEs with the EDTA ligand are relatively high, with a log K 260 between 15 and 20 [17, 18]. They are certainly higher than those of the various REE 261 related compounds in biomass. The formation of REE-EDTA complexes can thus make 262 it possible to displace the reactions that are at the origin of the interactions between 263 REE and organic matter. Insofar as these interactions help to maintain the REEs within 264 the biomass, this leads to better extraction. In the case of citrates, the stability constants 265 of the REE-citrate complexes are lower. The fact that the equilibria involved do not allow 266 sufficient displacement of the reactions at the origin of these interactions could explain a 267 low extraction. For the other elements, yields in water were low (< 20%), except for K 268 which had a very high extraction yield (> 80%). This was also the case for all the other 269 solutions used. K is known as a very extractable element from hyperaccumulators [10]. 270 The extraction of divalent and trivalent elements was improved by the addition of an acid 271 or a complexing molecule. However, lowering the pH is to be preferred to increase their 272 extraction yields.

In conclusion, even if REEs are directly extractable from biomass with water, higher
yields (> 80%) are obtained either with strong complexing agents (*EDTA*) or with strong
acid. These two options were selected for the following.

#### 277 4.3 **Precipitation of REEs in EDTA medium with oxalate**

#### 278 4.3.1 **REE speciation in presence of** *EDTA* **and oxalic acid**

279 To precipitate REEs, oxalic acid was added to the EDTA extract. The co-existence of 280 EDTA and oxalic acid leads to the formation of a large number of chemical species. As 281 the former is a tetraacid and the latter a diacid, the concentrations of the different 282 chemical forms depend on the pH. These different forms react with the REEs to form 283 soluble species such as REEEDTA<sup>-</sup> or insoluble species such as REE<sub>2</sub>Ox<sub>3</sub>, this last compound being the desired one. The concentrations of EDTA<sup>4-</sup> and Ox<sup>2-</sup> increase with 284 increasing pH, but the precipitation of insoluble REE hydroxides increases at the same 285 time. Therefore, the search for a pH high enough to obtain the  $Ox^{2}$  form that allows the 286 287 precipitation of REEs (Eq. 1) under the oxalate form is a compromise to avoid high solubilization of REEs due to the complex formed with EDTA<sup>4-</sup> (Eq. 2). 288

289 Lanthanum was chosen as a representative REE, and the composition of a solution 290 containing La, oxalate and EDTA was simulated as a function of pH, using Chess 291 software. Results are represented by lines in Fig. 4. The concentration term on the y-292 axis does not correspond to the conventional definition of concentration. In the Chess 293 nomenclature, for a solid, it is the number of moles of precipitates per litre of solution. 294 Only the main species are shown in this figure. Under very acid conditions (pH < 1), La is a trivalent cation (La<sup>3+</sup>). As the pH increases, it reacts with oxalate to form insoluble 295 296 La oxalate  $(La_2(C_2O_4)_3)$ , then it is complexed with *EDTA* (to give soluble LaEDTA). 297 Finally, at very high pH (> 12), La hydroxide precipitates. At pH between 1 and 2, La is 298 totally precipitated as oxalate. Above this pH range, LaEDTA appears to become 299 predominant at pH 4.5, and the only species at pH > 5.3.



In order to validate the results obtained in the previous part, a synthetic solution containing La, *EDTA* and oxalic acid was prepared at different pH values (2 to 7). Precipitate masses and concentrations of La were measured after reaction (confidence intervals were estimated at 10%). The experimental results are plotted in Fig. 4. Overall, a very good agreement was found between the experimental results and the simulations.

Based on these results, it can be concluded that at a pH between 1.2 and 2.3, REEs precipitate as oxalate, even in the presence of a high concentration of *EDTA* in solution. The simulations also indicate that as  $EDTA/C_2H_2O_4$  molar ratio increases, the pH corresponding to the total complexation of La with *EDTA* decreases (from 5.2 to 4.6, when the *EDTA* concentration is multiplied by 5). Conversely, when the amount of oxalic acid is multiplied by 5, this pH increases to 6. In addition, for a given concentration of La,
 increasing the addition of oxalic acid limits the formation of La<sup>3+</sup> and increases the pH
 range (especially at low pH) at which lanthanum can be precipitated.

324

# 325 4.3.3 Precipitation of REEs in biomass extract

## 326 a) pH adjustment

327 Firstly, the extract at pH 4.1 was acidified in order to reach the previously defined pH (ca 328 2.3), resulting in the formation of a first precipitate  $S_1$  (6.5 g L<sup>-1</sup>) from the protonation of 329 EDTA in the H<sub>4</sub>EDTA form, which has a low aqueous solubility. This precipitation was 330 confirmed by XRD analysis (Fig. 5). All identified peaks corresponded to EDTA. No other 331 molecules have been identified. During extraction, the molar ratio EDTA/REEs was 332 approximately 21 so that competitive reactions between REEs and the other elements 333 present in solution did not influence their extraction. This molar ratio could be optimized, 334 however, even in the present conditions, half of the EDTA (45%) can be recovered and 335 reused after filtration.





337

Fig. 5: Diffractogram of the solid  $S_1$  obtain after pH adjustment for extraction with *EDTA*.

All the peaks correspond to *EDTA* (except the two marked with an arrow)

Analysis of this solid showed that less than 2% of the REEs were lost at this stage. This percentage was calculated from the amount of initial biomass. Other elements are also identified: 6.5% and 1.5% for P and Ca respectively. For the other elements, only 0.4 to 0.6% of the initial biomass content is found in  $S_1$ . This step could be a way to recycle.

344

# b) REE precipitation using oxalic acid

345 The addition of oxalic acid caused a decrease in pH. To ensure that the pH is far from 346 the lower limit of precipitation, a large excess was introduced (molar ratio oxalate/REE = 347 70), giving a final pH of 1.3. The large amount of reagent was also chosen to reduce 348 competition from other cations and SOM (organic carbon concentration: 7.0  $\pm$  0.2 g L<sup>-1</sup>). 349 The formation of the oxalate precipitate  $(S_2)$  was instantaneous. After filtration, analysis 350 of the extract showed that REE precipitation was almost complete. Concentrations of 351 Ce, Gd, Pr and Sm were below the detection limit (10  $\mu$ g L<sup>-1</sup>) and less than 1% remained in solution for the other three main REEs (La, Nd, Y). Thus, this process can recover 352 353 65.9 to 72.2% of the REEs present in the initial biomass. The organic matter had no 354 influence on the operations implemented.

The recovered solid (2.56 g  $L^{-1}$  of extract, 42.6 mg g<sup>-1</sup> of raw biomass) was washed with deionized water, which removed most of the sodium, coming from the addition of *EDTA* during extraction. Finally, the precipitate contained 4.62% (27.3 mg) of REEs, which was 13 times more concentrated than the starting plant (Table 3). Calcium was the major impurity in the solid.

360

Table 3: Elemental composition of the precipitate  $S_2$  (%), obtained after oxalic acid addition in the extract with *EDTA* (confidence ranges are determined with 3 replicates)

E la ma		
Element		Wt%
REEs	Total	4.621 ± 0.489
	Ca	0.456 ± 0.012
	Р	0.117 ± 0.003
Other	K	0.043 ± 0.001
elements	AI	0.029 ± 0.001
	Pb	0.028 ± 0.001
	Na	0.020 ± 0.001
	S	0.015 ± 0.001

364 Stricter pH control and optimization of the amounts of *EDTA* and oxalic acid used during 365 the extraction would certainly improve the REE content of the S<sub>2</sub> precipitate. Due to the 366 low content (REEs and impurities), it appears that an excessive amount of *EDTA* has 367 still precipitated. This was confirmed by an XRD analysis of the precipitate, where only 368 *EDTA* was identified.

Even in choosing better conditions, it seems difficult to avoid completely this coprecipitation of EDTA, which will be lost during the calcination step. As it is an expensive reactant, precipitation in  $H_2SO_4$  medium was also tested.

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#### 373 4.4 Precipitation of REEs in H<sub>2</sub>SO<sub>4</sub> with oxalate

### 4.4.1 REE speciation in the presence of $H_2SO_4$ and oxalic acid

375 As with the previous approach, the objective is to precipitate REEs as oxalate. The 376 insoluble form referred to is  $La_2(C_2O_4)_3$ . However, in the presence of sulphuric acid, REE 377 sulphate complexes are formed, with  $REE(SO_4)^+$  being the predominant species 378  $(REE(SO_4)_2)$  is negligible all the time). A typical simulation of the equilibria is exhibited in 379 Fig. 6. The precipitation of REEs as oxalate depends on pH. At low pH, sulphate forms 380 solubilize REEs (pH is below the pKa of oxalic acid (1.2), C<sub>2</sub>H<sub>2</sub>O<sub>4</sub> is predominant), while 381 hydroxides are formed under alkaline conditions. When the sulphate concentration is 382 divided by 2, the pH corresponding to the formation of  $REE(SO_4)^+$  decreases from 2.5 to 383 2. Around pH 3.5 - 4 and above, two species of oxalate coexist,  $La_2(C_2O_4)_3$  and  $La(C_2O_4)_3^{3-}$ . Of course, the higher the oxalate/REEs ratio, the higher the concentration of 384  $La(C_2O_4)_3^{3-}$  (this species is very low when the oxalate/REE molar ratio is less than about 385 386 30). Therefore, there are two advantages to not raising the pH of the solution too much: 387 less reagent used and no risk of formation of an ionic REE oxalate compound. Based on 388 these results, the pH of the solution should be set between 2.5 and 3.0 to optimize REE 389 oxalate precipitation.

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#### 398 4.4.2 Precipitation of REEs in biomass extract

Following the recommendations determined by simulation, the pH of the extract obtained with D. linearis was first adjusted to 2.6 by NaOH addition (the initial pH was 0.8). After addition of oxalic acid (8 mmol for 0.37 mmol of REE), a precipitate S<sub>3</sub> appeared directly. After filtration, analysis of the aqueous solutions showed that 90% of REEs precipitated. The 10% remaining in solution were mainly La and Y. Fig. 7 shows the percentage of REE oxalate precipitation in relation to the initial content of the plant, as well as what remained in solution. The addition of oxalic acid resulted in the precipitation of 68.6% of the initial REEs (present in the initial biomass).









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412 The other elements were also analysed to assess the purity of the REE-oxalate 413 precipitate (Table 4). The mass ratio of REEs in this solid was 52.35%. Identified 414 impurities represent 3.13% by mass. Considering that REEs are precipitated as 415 Ox<sub>3</sub>REE<sub>2</sub> the remainder of this mass corresponded to the oxalate. The main impurity 416 was Na, from the sodium hydroxide solution used to neutralize the extract. A better 417 washing of the solid could certainly reduce this impurity.

418 These results prove that SOM did not inhibit precipitation under these conditions. The 419 REE content is more than 10 times higher in  $S_3$  (H<sub>2</sub>SO<sub>4</sub>) than in  $S_2$  (EDTA) (52.4 and 420 4.62% respectively), while the recovery yields are similar (68.6 and 69.3% respectively). 421 This is due to the presence of *EDTA* in  $S_2$ .

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Element  $S_3$  (wt%)

	La	21.00 ± 4.86
	Ce	7.42 ± 2.29
	Nd	14.66 ± 2.70
REES	Pr	4.08 ± 1.88
	Sm	2.31 ± 0.89
	Y	1.84 ± 0.45
	Gd	0.99 ± 0.37
	Na	2.00± 0.69
Other	K	0.22 ± 0.10
elements	Al	0.31 ± 0.17
	Ca	0.39 ± 0.13
	S	0.11 ± 0.05

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#### 428 4.5 Calcination of the oxalate precipitate

To obtain REE-oxides,  $S_2$  and  $S_3$ , obtained from *EDTA* and sulphuric acid extractions respectively, were burnt. Many parameters can influence the final composition of the solids; to compare the two processes, a calcination step at 700 °C for 2 h was chosen in a first approach. Fig. 8 shows the mass ratios of the various elements identified in the two precipitates after calcination (determined by ICP analyses using the method described in paragraph 2.3.).



In the case of *EDTA* leaching, REEs represented 79.8% of the mass of the calcination residue. The identified impurities were Ca (2.30%) and Al (1.94%) coming from the plant and Na (1.34%) mainly from *EDTA*. Given that the elements were in the form of oxides (REE<sub>2</sub>O<sub>3</sub>, CaO, etc.) the remainder of this mass mainly corresponded to oxygen.

In the case of  $H_2SO_4$  leaching, REEs represented 78.9% of the calcination residue mass while the mass ratio of impurities was 5.2% of which 3.8% is Na. It can be noted that the calculated uncertainties are high. They take into account the different stages from the acid leaching to calcination. It is important to note that experiments are carried out with very low masses and that the mass of the final product obtained is 300 times smaller than the initial mass of plant.

The two leaching methods led to close purities with a slightly different impurity profile. For example, Ca content was slightly higher when *EDTA* leaching was used (2.3% against 0.4%). This can be explained by the stability of the Ca-EDTA complex.

453 XRD and EDX analyses showed a better crystallized solid when extraction was 454 performed with sulphuric acid. For example, cerium oxide ( $Ce_7O_{12}$ ) was well identified. In 455 the case of an extraction with EDTA, the final solid contains carbon, coming from EDTA 456 calcination.

#### 457 **5. Conclusion and perspectives**

458 A three-step process involving extraction, oxalate precipitation and calcination has been 459 developed to produce REE oxides from hyperaccumulator D. linearis fern. Proof of 460 concept has been established. Experimental study of the extraction showed that REEs 461 can be extracted from the plant using either EDTA or sulphuric acid. A methodology based on speciation studies was successful in determining the operating conditions for 462 463 precipitating REEs in oxalate form. The pH range to be maintained, close to pH 2 for 464 both extraction methods, was found to be critical. The experimental study showed that 465 the soluble organic matter was also extracted from the plant but did not inhibit the 466 precipitation of REE oxalate.

467 Overall, REE purity was similar in solids after calcination (impurity content close to 5 468 wt%). However, even if extraction yields were higher with *EDTA* (87.5% against 79. %),

- the yield for the whole process was higher with  $H_2SO_4$  (72. % against 60.5%). The final solid contained 0.7 g of REES per g.
- The proposed process has a major advantage not to precipitate AI with oxalate (even if more than 50% is extracted). Here, and especially after  $H_2SO_4$  extraction, the final solid contained only 0.4% AI.
- 474 Experiments have shown that the choice of  $H_2SO_4$  as the extraction agent was relevant 475 to EDTA. Yields are better, it is cheaper, and a part of *EDTA* is burnt (lost) during 476 calcination. Further studies are required to optimize this process (amount of  $H_2SO_4$  and 477 oxalic acid, combustion temperature *etc.*). Reducing the quantity of reactants and 478 studying the possibility of recycling certain solutions is necessary to design and scale up 479 a process, which will allow the development of agromining development to recover 480 REEs from secondary resources with more environmentally friendly techniques.
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## 489 **References**

- 490
- 491 [1] G. Charalampides, K. I. Vatalis, B. Apostoplos, B. Ploutarch-Nikolas, Rare Earth
  492 Elements: Industrial Applications and Economic Dependency of Europe, Procedia
  493 Economics and Finance, 24 (2015) 126–135. DOI: 10.1016/S2212-5671(15)00630494 9.
- 495 [2] K. Binnemans, P.T. Jones, B. Blanpain, T. Van Gerven, Y. Yang, A. Waltone, M.
  496 Buchert, Recycling of rare earths: a critical review, Journal of Cleaner Production,
  497 51 (2013) 1–22. DOI: 10.1016/j.jclepro.2012.12.037.
- 498 [3] M.K. Jha, A. Kumari, R. Panda, J.R. Kumar, K. Yoo, J.Y. Lee, Review on
  499 hydrometallurgical recovery of rare earth metals, Hydrometallurgy, 161 (2016) 77–
  500 101. DOI: 10.1016/j.hydromet.2016.01.035.

- 501 [4] A. Van der Ent, G. Echevarria, J.L. Morel, A. Baker, Agromining: farming for metals,
  502 first ed., Springer International Publishing, Cham. 2018.
- 503 [5] A. Bani, G. Echevarria, D. Pavlova, S. Shallari, J.L. Morel, S. Sulçe, Element Case
  504 Studies: Nickel, in: A. Van der Ent, G. Echevarria, J.L. Morel, A. Baker, Agromining:
  505 farming for metals, first ed., Springer International Publishing, Cham, 2018, pp.
  506 221–232.
- 507 [6] M.-O. Simonnot, J. Vaughan, B. Laubie, Processing of bio-ore to products, in: A. Van
  508 der Ent, G. Echevarria, J.L. Morel, A. Baker, Agromining: farming for metals, first
  509 ed., Springer International Publishing, Cham, 2018, pp. 39–52
- [7] A. Bani, G. Echevarria, X. Zhang, E. Benizri, B. Laubie, J.L. Morel, M.-O.Simonnot,
  The effect of plant density in nickel-phytomining field experiments with Alyssum
  murale in Albania, Australian Journal of Botany, 68(2) (2015) 72-77. DOI:
  10.1071/BT14285.
- [8] P. Nti Nkrumah, R. Tisserand, R.L. Chaney, A.J.M. Baker, J.L. Morel, R. Goudon, P.
  D. Erskine, G. Echevarria, A. van der Ent, The first tropical 'metal farm': Some perspectives from field and pot experiments, Journal of Geochemical Exploration 198 (2019) 114-122. DOI: 10.1016/j.gexplo.2018.12.003.
- 518 [9] J. Vaughan, J. Riggio, J. Chen, H. Peng, H.H. Harris, A. van der Ent, 519 Characterisation and hydrometallurgical processing of nickel from tropical 520 agromined bio-ore, Hydrometallurgy 169 (2017) 346-355. DOI: 521 10.1016/j.hydromet.2017.01.012.
- 522 [10] M. Guilpain, B. Laubie, X. Zhang, J.L. Morel, M.-O. Simonnot, Speciation of nickel
  523 extracted from hyperaccumulator plants by water leaching, Hydrometallurgy 180
  524 (2018) 192-200. DOI: 10.1016/j.hydromet.2018.07.024.
- [11] C. Liu, M. Yuan, W.-S. Liu, M.-N. Guo, H. Huot, Y.-T. Tang, B. Laubie, M.-O.
  Simonnot, J.L. Morel, R.L. Qiu, Element case studies: rare earth elements, in: A.
  Van der Ent, G. Echevarria, J.L. Morel, A. Baker, Agromining: farming for metals,
  first ed., Springer International Publishing, Cham, 2018, pp. 297-308.
- [12] T. Liang, S. Ding, W. Song, Z. Chong, C. Zhang, H. Li, A review of fractionations of
  rare earth elements in plants, Journal of Rare Earths 26 (2008) 7-15. DOI:
  10.1016/S1002-0721(08)60027-7.

- [13] X. Shan, H. Wang, S. Zhang, H. Zhou, Y. Zheng, H. Yu, B. Wen, Accumulation and
  uptake of light rare earth elements in a hyperaccumulator Dicranopteris linearis,
  Plant Science 165 (2003) 1343-1353. DOI: 10.1016/S0168-9452(03)00361-3.
- [14] W.-S. Liu, H.-X. Zheng, M.-N. Guo, C. Liu, H. Huot, J.L. Morel, A. van der Ent, Y.T.
  Tang, R.-L. Qiu, Co-deposition of silicon with rare earth elements (REEs) and
  aluminium in the fern Dicranopteris linearis from China, Plant and Soil 437 (2019)
  427-437. DOI: 10.1007/s11104-019-04005-0.
- [15] Z. Chour, B. Laubie, J.L. Morel, Y.T. Tang, R.L. Qiu, M.-O. Simonnot, L. Muhr,
  Recovery of Rare Earth Elements from Dicranopteris linearis by an enhanced ion
  exchange leaching process, Chemical Engineering and Processing-Process
  Intensification 130 (2018) 208-213. DOI: 10.1016/j.cep.2018.06.007.
- 543 [16] D. Qi, Hydrometallurgy of Rare Earths, Extraction and Separation, first ed., Elsevier
  544 Science, Amsterdam, 2018.
- 545 [17] T. Moeller, R. Ferrus, Observations on the rare earths—LXXIII the heat and entropy
  546 of formation of the 1: 1 chelates of N-hydroxyethylethylenediaminetriacetic acid with
  547 the tripositive cations, Journal of Inorganic and Nuclear Chemistry 20 (1961) 261548 273. DOI: 10.1016/0022-1902(61)80275-3.
- 549 [18] S.A. Wood, The aqueous geochemistry of the rare-earth elements: Critical stability
  550 constants for complexes with simple carboxylic acids at 25° C and 1 bar and their
  551 application to nuclear waste management, Engineering Geology 34 (1993) 229552 259. DOI: 10.1016/0013-7952(93)90092-Q.
- [19] M. Yuan, C. Liu, W.-S. Liu, M. -N. Guo, J.L. Morel, H. Huot, H.-J. Yu, Y.-T. Tang, R.L. Qiu, Accumulation and fractionation of rare earth elements (REEs) in the
  naturally grown *Phytolacca americana L.* in southern China, International Journal of
  Phytoremediation 20(5) (2018) 415-423. DOI:10.1080/15226514.2017.1365336.
- [20] J. van der Lee, L. De Windt, Chess Tutorial and Cookbook. Updated for Version
  3.0. Ecole Nationale Supérieure des Mines de Paris, Fontainebleau, France,
  Technical Report n LHM/RD/02/13. (2002) http://chess.geosciences.minesparistech.fr
- 561 21] National Institute of Standards and Technology (NIST), Database of Critically
   562 Selected Stability Constants of Metal Complexes, Version 8.0.

563 [22] D.-Y. Chung, E.-H. Kim, E.-H. Lee, J.-H. Yoo, Solubility of Rare Earth Oxalate in
564 Oxalic and Nitric Acid Media, Journal of Industrial and Engineering Chemistry 4(4)
565 (1998) 277-284.

- [23] M.N. Nguyen, A.A. Meharg, M. Carey, S. Dultz, F. Marone, S.B. Cichy, C.T. Tran,
  G.H. Le, N.T. Mai, T.T.H. Nguyen, Fern, Dicranopteris linearis, derived phytoliths in
  soil: Morphotypes, solubility and content in relation to soil properties, European
  Journal of Soil Science 70(3) (2019) 507-517. DOI: 10.1111/ejss.12754.