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1 2	Taxonomic revision of the Sylvarum group of bumblebees using an integrative approach
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24 Abstract

25 Recent improvements in taxonomy consider multiple operational criteria. Integrative taxonomy provides a methodological framework merging these multisource approaches. 26 Bumblebees are considered as a uniform group where their taxonomy remains one of the most 27 28 difficult. Here, we investigate the taxonomic statuses inside a monophyletic group including six taxa (B. inexspectatus, B. mlokosievitzii, B. ruderarius, B. sylvarum, B. velox and B. 29 veteranus) in the most diverse subgenus of bumblebees: Thoracobombus. We used an 30 31 integrative approach based on mitochondrial and nuclear genetic markers and eco-chemical traits commonly used in bumblebee taxonomy. For all species, our study shows a clear 32 differentiation in DNA and eco-chemical traits. However, we conserve the subspecies status 33 of B. ruderarius simulatilis and B. sylvarum daghestanicus from the east of Turkey and Iran 34 according to their degree of genetic and eco-chemical differentiation. 35

36 <u>Keywords:</u> Bumblebees, eco-chemical traits, integrative taxonomy, mitochondrial genetic
 37 maker, nuclear genetic maker, *Thoracobombus*.

38 Introduction

Species are considered as the fundamental unit in biology and their accurate delimitation is 39 essential respectively in biodiversity classification and in evolutionary and conservation 40 biology (Moraes, 1987; Paquin, Dupérré, Cokendolpher, White & Hedin, 2008; Wheeler, 41 Raven & Wilson, 2004). Taxonomy inside invertebrate groups is mainly based on 42 morphological approach. However, the common strong homogeneity in morphology may be 43 an obstacle for identification, especially in the case of cryptic species (Williams et al., 2012). 44 Moreover, many species exhibit a high intra-specific variability and cases of convergent color 45 pattern were recorded in sympatry (Françoso, Freitas de Oliveira & Arias, 2016; Lecocq et al., 46 2015a). Therefore, many alternative features and methods such as DNA-based approaches 47

(e.g. pairwise distance thresholds, bootstrap support or reciprocal monophyly), geometric 48 49 morphometrics of wing shape or semiochemical traits have been used to delineate species (Lecocq et al., 2015b; Martin, Helantera & Drijfhout, 2008; Oleksa & Tofilski, 2015; White, 50 Pilgrim, Boykin, Stein & Mazor, 2014). In order to draw a strongly supported taxonomic 51 hypothesis, recent betterment in taxonomy consider multiple operational criteria from the 52 same sampling (Gibbs, 2009; Lecocq et al., 2011; Martinet et al., 2018). The integrative 53 54 taxonomy based on the Unified Species Concept (USC) provides a methodological framework merging these multisource approaches (De Queiroz, 2007; Padial, Miralles, De la Riva & 55 Vences, 2010; Schlick-Steiner et al., 2010). The USC argues that all species concepts agree 56 57 that species exist as separately evolving metapopulation lineages but diverge in their criterion for delimiting species (De Queiroz, 2007). Therefore, ecological differentiation, phylogenetic 58 divergence, morphological characteristic or reproductive isolation could highlight a 59 60 metapopulation lineage separation by avoiding false taxonomic conclusions driven by a single trait. Moreover, by considering subspecies as a step in the process of allopatric speciation 61 (Mayr, 1942), and because of its important value in conservation and evolutionary biology 62 (Coppée, Terzo, Valterová & Rasmont, 2008; Phillimore & Owens, 2006; Rasmont, Coppée, 63 Michez & De Meulemeester, 2008; Sackett et al., 2014; Terzo, Urbanová, Valterová & 64 65 Rasmont, 2005), subspecies status has been proposed as solution when differentiation is highlighted by a subset of operational criteria. This proposal aims to reduce the under-66 estimate risk of a congruence approach (Hawlitschek, Nagy & Glaw, 2012; Padial et al., 67 68 2010).

Bumblebees are considered as a complex group where their classification remains one of the most difficult in bees (Moure & Sakagami, 1962; Murray, Fitzpatrick, Brown & Paxton, 2008). In this group, integrative taxonomy has considerably improved taxonomic studies and has solved many doubtful cases thanks to operational criteria as DNA (mitochondrial and

nuclear) and semio-chemicals (Bertsch, Schweer, Titze & Tanaka, 2005; Lecocq et al., 2015a;
Martinet et al. 2018).

In this paper, we apply an integrative approach to assess the taxonomic status of a group of phylogenetically closely related species in the *Thoracobombus* Dalla Torre, 1880 subgenus: *B. mlokosievitzii* Radoszkowski, 1877, *B. ruderarius* (Müller, 1776), *B. sylvarum* (L., 1761), *B. velox* (Skorikov, 1914), *B. veteranus* (Fabricius, 1793) and *Bombus inexspectatus* (Tkalců, 1963). We revise the taxonomy of this group through a decision framework based on differentiation of three operational and independent criteria commonly used in bumblebee taxonomy: a mtDNA marker (COI), a nuDNA marker (PEPCK) and an eco-chemical trait.

82

83 Materials and methods

84 Studied taxa

B. ruderarius (Müller, 1776) is a widespread species distributed from Tunisia in the south to 85 the Arctic Circle in the north (Rasmont et al., 2015a). Four subspecies and one form have 86 been described (Rasmont, 1983): (i) B. ruderarius ruderarius (Müller, 1776) is characterized 87 by a reddish tail and the rest of the body is black intermixed with more or less greyish hairs 88 89 (Fig. 1C). They occurred throughout Europe except the Mediterranean region; (ii) B. 90 ruderarius simulatilis Radoszkowski, 1888 with a wide white collare and scutellare bordering 91 a black interalar band, the tergite 1 and 2 completely white and with the tergite 3 black, the 92 tail is red (Fig. 1D). This subspecies occurred in mountainous regions of Anatolia as well as in the Caucasus region in north Iran; (iii) B. ruderarius tunensis (Tkalců, 1974) is 93 characterized by three yellow bands and a white tail and is endemic to the east of Algeria and 94 north of Tunisia; (iv) B. ruderarius sartus (Skorikov, 1922) with a wide yellowish collare and 95 scutellare bordering a black interalar band, the tergite 1 and 2 completely yellowish and with 96 97 the tergite 3 black, the tail is red. The eastern limits of the species were known to be the mountains of east Turkestan but other specimens were found in Mongolia (Skorikov, 1922);
(v) *B. ruderarius* form *montanus* is characterized by large yellowish bands compared to the
dark nominal subspecies (Fig. 1C). They occur from the Cantabrian Mountains to the
Pyrenees where they coexist with the nominal subspecies *B. ruderarius ruderarius*.

B. sylvarum (L., 1761) is also a widespread species distributed in the west-palaearctic region 102 but hardly reaches Mongolia (Rasmont et al., 2015a). We considered four subspecies and one 103 104 form (Rasmont, 1983): (i) B. sylvarum sylvarum (L., 1761) with three large greyish bands (thorax and abdomen) and thin grey bands (abdomen) and a reddish tail intermixed with grey 105 (Fig. 1F). They occur in the west-Palaearctic region; (ii) B. sylvarum citrinofasciatus Vogt, 106 107 1909 is characterized by grey yellowish bands compared to the nominal subspecies. Moreover, the thin grey bands on the abdomen are often absent. They occur in the west of 108 Turkey; (iii) B. sylvarum daghestanicus Radoszkowski, 1877 is characterized by white bands 109 110 and a reddish tail (Fig. 1E). This subspecies is distributed in the east of Turkey, the Caucasus region as well as in Iran; (iv) B. sylvarum rogenhoferi Dalla Torre, 1882 with white yellowish 111 bands, a red tail and without thin grey bands on the abdomen is restricted to south Italy and 112 Sicily; (v) B. sylvarum form nigrescens Pérez, 1879 is considered as the melanic form inside 113 B. sylvarum species. They are completely black with a reddish tail and with or without 114 115 intermixed grey hairs on the thorax (Fig. 1G). They occur in the south-west of France, the Atlantic regions as well as the south of Scandinavia. Currently, all these subspecies and form 116 are considered as *B. sylvarum* (Rasmont et al., 2015a; Williams, 1998). 117

118 All other species included in this paper form a monophyletic group with *B. ruderarius* and *B.*

- 119 *sylvarum* (Cameron, Hines & Williams, 2007) and will be treated as comparison group.
- 120 Bombus inexspectatus (Tkalců, 1963) (Fig. 1A) is a sub-alpine uncommon European species
- restricted in the Cantabrian Mountains and the Alps from 1200m to 2100m in altitude (Amiet,
- 122 1996; Rasmont et al., 2015a; Tkalců, 1963, 1965; Yarrow, 1970). It has been described as a

species adapted to parasitic life (Müller, 2006; Yarrow, 1970) with *B. ruderarius* as its only
host (Müller, 2006). There is no doubt about its specific status.

B. mlokosievitzii Radoszkowski,1877 is a ponto-mediterranean species whose distribution 125 extends from the Balkan Mountains to Caucasian region via the south of the Black Sea 126 (Rasmont & Iserbyt, 2014). Three subspecies diagnosable by their coat color variation has 127 been defined: (i) B. mlokosievitzii mlokosievitzii Radoszkowski, 1877 with a wide greyish 128 129 white collare and scutellare bordering a black interalar band. This subspecies is distributed in the north of Anatolia; (ii) B. mlokosievitzii convergens Skorikov, 1908 characterized by a 130 thorax completely greyish white is distributed in the Caucasian region and (iii) B. 131 132 mlokosievitzii vogtiellus diagnosable by their thorax completely black (Fig. 1B) and distributed in Greece, Macedonia, Bulgaria and west of Anatolia. The presence of a reddish 133 band on tergite 2 is common to all subspecies. Unfortunately we were not able to collect all 134 subspecies inside this taxon and only the subspecies *mlokosievitzii* was considered in this 135 136 study.

B. velox (Skorikov, 1914) is restricted to the north east of Anatolia and the Caucasus region. It
is an uncommon species with two greyish bands on the thorax and one on the tergite 1, the
rest of the abdomen is yellowish (Fig. 1H). There is no doubt about its specific status.

B. veteranus (Fabricius, 1793) lives in sparse population in the plains of northern Europe
(Rasmont & Iserbyt, 2014). This species presents very little color variation and is characterize
by a yellowish coat color on all the body except the presence of a black interalar band (Fig.
11). There is no doubt about its specific status.

144 *Sampling*

145 150 male and female specimens were collected between 2001 and 2017 in the Palaearctic 146 region (supplementary file 1, Tab. 1). All specimens were determined based on their 147 morphology. We attributed a name to the specimens without taxonomic a priori on the species

status. The ingroup comprises: ruderarius from Estonia (n=5), France (n=16), Macedonia 148 149 (n=1), Serbia (n=1), Slovakia (n=4), Sweden (n=1) and Switzerland (n=5), montanus from Andorra (n=3), Spain (n=2) and France (n=5), *simulatilis* from Iran (n=3) and Turkey (n=9), 150 sartus from Kyrgyzstan (n=2) and Mongolia (n=2), sylvarum from Belgium (n=3), France 151 (=13), Germany (n=4), Italy (n=3), Poland (n=6), Serbia (n=2), Slovakia (n=1), Spain (n=2) 152 and Sweden (n=4), *daghestanicus* from Iran (n=1) and Turkey (n=8), *nigrescens* from Spain 153 154 (n=1) and Sweden (n=5), rogenhoferi from Italy (n=2). The comparison group comprises: *inexspectatus* from Italy (n=9); *mlokosievitzii* from Turkey (n=11), *velox* from Turkey (n=1) 155 and veteranus from Belgium (n=4), France (n=4) and the Netherlands (n=4). The 156 157 phylogenetically related species *B. mucidus* (n=3) was used as outgroup.

158 *Genetic trait analyses*

Two genes commonly used in bumblebee phylogeny were sequenced (Cameron et al., 2007; Martinet et al., 2018; Williams et al., 2012): the "barcode" region of the cytochrome oxidase 1 (COI) mitochondrial gene was sequenced from 88 specimens, and the nuclear protein-coding gene phosphoenolpyruvate carboxykinase (PEPCK) was sequenced from 83 specimens (supplementary file 1).

164 We carried out polymerase chain reaction amplifications with new design primer pairs (5'-TGATCAGGAATAATTGGATCTTCA-3') 165 CSylG-F/CSylG-R: (5'-/ TGATCAGGAATAATTGGATCTTCA-3') PSylG-F/PSylG-R: 166 for COI and (5'-CTTTAAAGGCAATTGTATCAAAGTT-3') / (5'-CTCGCCAATCGGTGATCTAT-3') for 167 PEPCK. We performed sequencing procedures described in Lecocq et al. (2013). Sequences 168 were aligned with BioEdit version 7.2.5 (Hall, 1999). The resulting COI (624bp) and PEPCK 169 170 (894bp) sequences are available on GenBank (accession numbers in supplementary file 1). We investigated the potential genetic differentiation for each gene individually within the six 171

taxa through haplotype networks analyses. We used the median-joining method with Network

4.6.1.1 software (www.fluxus-engineering.com) to produce haplotype networks. We weighted
transversions twice as high as transitions to reconstruct the network (Lecocq et al., 2015a;
Martinet et al., 2018). In phylogenetic analyses, gaps (deletion or insertion) are usually treated
as missing data. Because of the presence of gaps sites in our PEPCK genes, two haplotype
networks considering or not the gaps sites will be performed in order to be aware of
phylogenetic information provide by these two methods.

179 In phylogenetic analyses, we analyzed each gene independently with maximum likelihood (ML) and Bayesian (MB) methods to explore the genetic divergence and define lineages 180 within the different groups. For ML methods, we partitioned each gene to explore the best 181 182 substitution model: i) PEPCK into two introns and one exon; ii) COI and PEPCK exons by base position (1st, 2nd and 3rd) and choose the best fitting substitution models with JModelTest 183 Server 2.0 (Posada, 2008) using the Akaike information criterion corrected for small sample 184 185 sizes (Hurvich & Tsai, 1989). The chosen models were: i) For COI: TIM1 + I (1st), HKY (2^{nd}) , and TPM3uf + I (3^{rd}) ; ii) for PEPCK, exon: F81 (1^{st}) , JC (2^{nd}) , K80 (3^{rd}) ; intron 1: 186 TMP3uf + I; intron 2: HKY. Selected models which are not implemented in MrBayes were 187 substituted by the closest over-parameterized model. For each gene, ML analyses were 188 189 performed with ten independent runs in Garli 2.0 (Zwickl, 2006); the topology and -ln L was 190 the same among replicates. The run with the highest likelihood was retained. Statistical significance in nodes was evaluated with 10000 non-parametric bootstrap replicates. 191 Topologies with bootstrap values $\geq 85\%$ were considered as well supported (Hillis & Bull, 192 193 1993). For MB method, we performed Bayesian inference analyses with mrbayes 3.1.2 (Ronquist & Huelsenbeck, 2003). Ten independent analyses were achieved for each gene (100 194 195 million generations, four chains with mixed models, default priors, saving trees every 1000 generations). Then the first 10 million generations were removed as burn-in procedure. A 196 majority-rule 50% consensus tree was constructed. We considered as statistically significant 197

only branch supports (topologies) with high posterior probabilities (≥ 0.95) (Wilcox, Zwickl, Heath & Hillis, 2002). The phylogenetic analysis based on PEPCK gene fragment considered gaps as missing data. All trees were rooted with the taxon *B. mucidus*.

In order to recognize species threshold, we applied the Bayesian implementation of the 201 general mixed Yule-coalescent (bGMYC) method to the COI dataset by using R Package 202 "bGMYC" (Reid & Carstens, 2012). A range of probabilities < 0.05 was considered as strong 203 204 evidence that taxa was heterospecific while a range of probabilities > 0.95 suggested that taxa was conspecific (Reid & Carstens 2012). BEAST 1.7.4 (Drummond, Suchard, Xie & 205 Ranbaut, 2012) was used to generate ultrametric trees with a phylogenetic clock model in 206 207 order to calculate a posterior distribution of trees (length of the MCMC chain: 1 billion generations). The first million sampled trees were burned-in using the maximum clade 208 209 credibility method and setting the posterior probability limit to 0. The bGMYC analysis was 210 based on 1000 trees sampled every 10 000 generations. For each of these 1000 trees, the MCMC was made of 100 000 generations, discarding the first 90 000 as burn-in and sampling 211 every 100 generations. 212

213 Eco-chemical trait analyses

214 We focused on the cephalic labial gland secretions (CLGS) of males, the most commonly 215 reproductive trait involved in the bumblebee pre-mating recognition (Ayasse & Jarau, 2014). Currently, the composition of the CLGS is known for 15 species of *Thoracobombus* (Ayasse 216 & Jarau, 2014; Brasero et al., 2015; Brasero et al., 2018a,b) and this study will increase the 217 218 knowledge inside this group to 17 species. These secretions are species-specific (Calam, 1969) and provide an efficient diagnostic characters for species delimitation (Brasero et al., 219 220 2015). They are complex mixtures of mainly aliphatic compounds synthesized de novo (Coppée et al., 2008; Žáček et al., 2013) in the head of bumblebee males. Main components 221 were defined as those with the highest relative amount in at least one individual of the taxon. 222

All specimens were killed by freezing at -20°C and the CLGS were extracted with 400 μl of
heptane (method described in De Meulemeester, Gerbaux, Boulvin, Coppée, & Rasmont,
2011). Samples were stored at -40°C prior to analysis. We were able the sample 111
specimens belonging to the six studied taxa (Tab. 1).

We determined the CLGS composition by gas chromatography-mass spectrometry (GC/MS) 227 using a Focus GC (Thermo Scientific) with a non-polar DB-5 ms capillary column [5% 228 229 phenyl (methyl) polysiloxane stationary phase; column length 30 m; inner diameter 0.25 mm; film thickness 0.25 µm] coupled to DSQ II quadrupole mass analyzer (Thermo Scientific) 230 with 70 eV electron impact ionization. We used a splitless injection mode (220°C) and 231 232 helium as a carrier gas (one ml/min). The temperature program of the oven was set to 70°C for two minutes and then heated up at a rate of 10°C/min to 320°C. The temperature was then 233 held at 320°C for five minutes. Compounds were identified in XcaliburTM using the retention 234 235 times (t_r) and mass spectra of each peak, in comparison to those at National Institute of Standards and Technology library (NIST, U.S.A). Double bond positions (C=C) were 236 determined by dimethyl disulfide (DMDS) derivatization (Cvacka et al., 2008). 237

The CLGS compounds of all samples were quantified by a gas chromatograph-flame 238 239 ionization detector Shimadzu GC-2010 with a SLB-5ms non-polar capillary column (5% 240 phenyl (methyl) polysiloxane stationary phase; 30-m column length; 0.25-mm inner diameter; 0.25-µm film thickness) with the same chromatographic conditions as in GC/MS. Peak areas 241 of compounds were detected in GCsolution Postrun (Shimadzu Corporation) with automatic 242 243 peak detection and noise measurement. We calculated relative amounts (RA in %) of compounds in each sample by dividing the peak areas of compounds by the total area of 244 compounds. We discarded all compounds for which RA were recorded as less than 0.1% for 245 all specimens (De Meulemeester et al., 2011). The data matrix (supplementary file 2) for each 246

taxon was based on the alignment of each relative proportion of compound between allsamples performed with GCAligner 1.0 (Dellicour & Lecocq, 2013).

Before each samples injection, a standard (Kovats) containing a mix of hydrocarbons (alkanes) from C10 (decane) to C40 (tetracontane) was injected in order to facilitate the alignment and the identification of compounds. Kovats indices were calculated with GC Kovats 1.0 (Dellicour & Lecocq, 2013).

253 Clustering method were performed using R 3.3.2 (R Development Core Team, 2016) to detected CLGS differentiations between taxa. A Canberra distance matrix based on CLGS 254 (RA of each compound) was computed. An unweighted pair group method with arithmetic 255 256 mean (UPGMA) was used as clustering method (R-package ape, Suzuki & Shimodaira, 2011). We assessed the uncertainty in hierarchical cluster analysis using *p*-values calculated 257 via multiscale bootstrap resampling with 50 000 bootstrap replications (significant branch 258 259 support > 0.85) (R-package pvclust, Suzuki & Shimodaira, 2011). We assessed CLGS differentiations between taxa by performing a permutation multivariate analysis of variance 260 using distance matrix (perMANOVA) (R package vegan; Oksanen et al., 2011). When a 261 significant difference was detected, we performed a pairwise multiple comparison with an 262 263 adjustment of *P*-values (Bonferroni correction) to avoid type I errors.

To determine the indicator compounds (IC) of each taxon, we used the indicator value (IndVal) method (Claudet, Pelletier, Jouvenel, Bachet & Galzin, 2006; Dufrêne & Legendre, 1997). The value given is the product of relative abundance and relative frequency of occurrence of a compound within a group. We evaluated the statistical significance of a compound as an indicator at the 0.01 level with a randomization procedure.

269 Decision framework of taxonomic status

We used an approach that corresponds to the strictest commonly used framework called "integration by congruence" by Padial et al. (2010). We thus assigned the specific status to a

taxon (i) genetically differentiated in all genetic markers (i.e. unique haplotypes), (ii) which 272 273 constitute a monophyletic group with high branch support, and (iii) with a significant differentiation in CLGS composition (significance differentiation in perMANOVA and high 274 bootstrap values > 0.85). All criteria used must be convergent to assign specific status. This 275 approach can lead to underestimation of species differentiation but reduces the taxonomic 276 277 inflation. We assigned the subspecies status to phenotypically distinct allopatric population 278 that does not diverge in all lines of evidence. This decision reduces the 'underestimate's risk' of our strict integrative approach by naming them as a subspecies (Ennen et al., 2014; 279 Hawlitschek et al., 2012). 280

281

282 <u>Results</u>

283 *Genetic trait analyses*

284 All phylogenetic analyses (ML and MB) showed the expected differentiation between outgroup and ingroup (Fig. 2A, 3A). Seven strongly supported (bootstrap>0.95) monophyletic 285 groups are detected in our COI phylogenetic analyses (Fig. 2A): (i) one group with 286 inexspectatus specimens; (ii) one group with ruderarius, montanus, sartus and simulatilis 287 288 specimens; (iii) one with velox specimens; (iv) one group with veteranus specimens; (v) one 289 group including all sylvarum, nigrescens, daghestanicus and rogenhoferi specimens; (vi) one group with *mlokosievitzii* specimens and (vii) the outgroup *B. mucidus*. Within "*ruderarius* 290 group", two unexpected lineages were revealed (Fig. 2A): one main lineage strongly 291 292 supported including the widespread ruderarius taxa, montanus and sartus and one another lineage not strongly supported (bootstrap<0.95) including simulatilis taxon from Turkey and 293 294 Iran. As expected, we found a less structured tree in the PEPCK gene fragment for all taxa. PEPCK gene analyses showed little phylogenetic information and failed to reconstruct the 295 phylogenetic affinities between all taxa (Fig. 3A). However, we found some strongly 296

supported groups as *inexspectatus* and *mlokosievitzii*. As for COI gene, no geographical
entities were underlined by our PEPCK phylogenetic analyses expected for "*ruderarius*group" where the unexpected lineage *simulatilis* was revealed and strongly supported in our
MB (Fig. 3A) but not in our ML analysis.

15 private haplotypes were highlighted for COI gene while 11 and 10 haplotypes were 301 302 highlighted in PEPCK genes respectively considering or not the gaps sites by our network 303 analysis (Fig. 2B, 3B, C). Within the "ruderarius and sylvarum groups", only simulatilis displayed specific haplotypes in COI and PEPCK, while daghestanicus had specific COI 304 haplotype (Fig. 2B, 3B, C). Considering all taxa, COI revealed 72/624 nucleotide sites 305 306 substitution corresponding to 11.54% of phylogenetic information. Compared to the taxa inside their groups, simulatilis and daghestanicus revealed respectively 0.96% (6/624 307 diagnostic nucleotide sites) and 0.64% (4/624 diagnostic nucleotide sites) of phylogenetic 308 309 information in COI gene (Fig. 2B). Considering all taxa, PEPCK gene revealed 143/894 (considering gaps) and 23/894 (not considering gaps) nucleotide sites respectively 310 corresponding to 16% and 2.6% of phylogenetic information. Compared to the taxa inside 311 their groups, simulatilis revealed 1.79% (16/894 diagnostic nucleotide sites) of phylogenetic 312 313 information when gaps are considered (Fig. 3C) and 0.34% (3/894 diagnostic nucleotide sites) 314 when gaps are not considered (Fig. 3B).

The tree generated by bGMYC analysis highlighted several entities with low probabilities (<0.05) to be conspecific with the other ones (Fig. 4). It suggested the delimitation of 7 prospective species (P<0.05) (Fig. 4) which match with those obtained in the COI tree (ML and MB, Fig. 2A): (i); one group with *inexspectatus* specimens (P=1); (ii); one group with *velox* specimens (P=1); (iii) one group with *ruderarius, sartus, montanus* and *simulatilis* specimens (P>0.16-1); (iv) one group with *veteranus* specimens (P=1); (v) one group including *sylvarum, daghestanicus, nigrescens, rogenhoferi* specimens (P>0.34-1); (vi) one

- group with *mlokosievitzii* specimens (P=1) and (vii) one group with the outgroup *B. mucidus*(P=1). The pairwise matrix shows a non significant heterospecificity threshold between *simulatilis* and the other taxa from *ruderarius* group (P>0.05).
- 325 Eco-chemical trait analyses

The chemical analyses highlighted 7 strongly supported groups (bootstrap > 85%) (Fig. 5). A: 326 velox (Turkey); B: inexspectatus (Italy); C: mlokosievitzii (Turkey), D: sylvarum + 327 328 daghestanicus + nigrescens + rogenhoferi (Belgium, France, Germany, Iran, Italy, Poland, Serbia, Slovakia, Spain, Sweden and Turkey); E: veteranus (Belgium, France and Holland); 329 F: montanus + ruderarius (Andorra, Estonia, France, Macedonia, Serbia, Slovakia, Spain, 330 331 Sweden and Switzerland) and G: simulatilis (Turkey). 95 compounds were detected (respectively 17 from A, 36 from B, 34 from C, 55 from D, 36 from E, 49 from F and 44 332 from G; see supplementary file 2). For each group, we detected main compounds, A: 333 334 octadecen-1-ol (88.2%); B: hexadec-7-en-1-ol (17.2-57.8%), octadec-11-en-1-ol (12.2-32.6%) and octadec-11-envl acetate (10-23.4%); C: hexadec-7-en-1-ol (14-46.7%) and hexadec-7-335 enyl acetate (19.6-45.9%); D: hexadec-7-en-1-ol (2.2-57.5%) and hexadec-7-enyl acetate 336 (20.1-66.6%); E: hexadec-9-en-1-ol (1.1-57.8%) and ethyl octadec-11-enoate (1.5-31.4%); F: 337 338 hexadec-9-en-1-ol (0-62.3%), hexadec-9-enyl acetate (0-32.7%), tricos-9-ene + tricos-7-ene 339 (2.1-59.2%) and hexadecenyl hexadecenoate2 (0.1-25.1%) and G: hexadec-9-en-1-ol (4.5-59.9%), octadecadien-1-ol (0.4-22%), and tricos-9-ene + tricos-7-ene (5.7-54.2%) and 340 tricosane (2-20%). Moreover, the IndVal method highlighted several significantly indicator 341 342 compounds (IC) (IndVal value>70) for the group A (IC=4), B (IC=11), C (IC=3), D (IC=7), **E** (IC= 5), **F** (IC=1) and **G** (IC=0) (supplementary file 2). Despite the separation we observed 343 between montanus + ruderarius and simulatilis our IndVal method did not find indicator 344 compounds for simulatilis, however, our statistical analysis significantly supports this 345 differentiation (bootstrap > 85%). perMANOVA test confirmed the differentiations between 346

all groups (see supplementary file 3) and particularly between *montanus* + *ruderarius* and *simulatilis* (p-value: 0.041; F: 2.9986).

349 Decision framework of taxonomic status

Within the "*ruderarius* group" (*ruderarius*, *sartus*, *montanus* and *simulatilis*), operational criteria did not reflect the current subspecific differentiation (Tab. 2). CLGS and haplotype networks analyses detected differentiation between *simulatilis* and the other taxa. We then considered *B. ruderarius simulatilis* as a subspecies inside the "*ruderarius group*". This subspecies occurs in mountainous regions of Anatolia, the Caucasus region as well in north Iran. We also assigned the subspecific status *B. ruderarius ruderarius* (Müller, 1776) to the other taxa (Tab. 2).

The genetic structure observed in COI, PEPCK and CLGS did not reflect the current 357 subspecific status encountered in the "sylvarum group" (sylvarum, daghestanicus, nigrescens 358 359 and rogenhoferi). Based on its unique haplotype in COI we decided to consider B. sylvarum daghestanicus as a subspecies inside the "sylvarum group" as described by Radoszkowski 360 (1877). This subspecies occurred in mountainous regions of Anatolia, the Caucasus region as 361 well in Iran. We then assigned the subspecific status B. sylvarum sylvarum (Müller, 1776) to 362 the other taxa inside which we considered the forms sylvarum, nigrescens and rogenhoferi 363 364 based on their color patterns (Tab. 2). The specific status of B. inexspectatus, B. mlokosievitzii, B. velox and B. veteranus were confirmed by our results. 365

366

367 Discussion

Genetic character along with CLGS can confirm with a high degree of certainty the speciation processes between taxa. We established the species status of our different taxa according to their divergences in genetic markers (mitochondrial and nuclear) and their CLGS differentiation (Tab. 2).

Our integrative approach support the specific status of all closely related consubgeneric six 372 373 species studied here (Cameron et al., 2007; Williams, 1998). Similar interspecific differentiations were recorded independently based on DNA (Cameron et al., 2007, Hines & 374 Cameron, 2010) and CLGS composition (Appelgren, Bergstrom, Svensson & Cederberg, 375 1991; Brasero et al., 2018a; Terzo et al., 2005). However, the analyses in this study were 376 carried out on a larger sampling covering the distribution of the species. Within "ruderarius 377 378 group" and "sylvarum group" and despite some phenotypic differentiations, all studied taxa are regarded as conspecific (Tab. 2). ruderarius and sylvarum groups appear respectively as 379 single species with high geographic phenotypic differentiations. In the light of our results, we 380 381 decide for the "sylvarum group" to use the oldest available nominal species name as the valid name for all of them: B. sylvarum sylvarum except for daghestanicus where we maintain the 382 subspecific status B. sylvarum daghestanicus from east of Turkey and Iran because of its 383 384 private haplotype in COI. Inside the "ruderarius group" we decide to conserve the subspecific status B. ruderarius simulatilis for the population in the east of Turkey and Iran based on their 385 CLGS differentiation and private haplotype (COI and PEPCK) and raised the other 386 populations to the oldest available nominal species name B. ruderarius ruderarius (see 387 388 decision framework Tab. 2). For the comparison groups B. inexspectatus, B. mlokosievitzii, B. 389 veteranus and B. velox, our analyses (Fig 2A, 3A, 4 & 5) confirm clear differentiation between these species (Cameron et al., 2007; Hines & Cameron, 2010). Inside B. 390 *mlokosievitzii*, we have not been able to analyze the subspecies *convergens* and *vogtiellus*. 391

392 Intraspecific differentiation of B. sylvarum and B. ruderarius

B. sylvarum is a widespread species and can be found in Central Spain, Sicily, southern Italy, Greece and Southern Turkey of its distribution. It reaches Ireland and northem Portugal to the West and Mongolia to the East. To the north, this species nearly reaches the Arctic Circle (Rasmont et al., 2015a). Divergence of color patterns (sometimes very pronounced: see the

melanic form *nigrescens*), seems to be a simple intraspecific variation as encountered in other 397 398 bumblebees species such as B. pascuorum (Lecocq, Brasero, Martinet, Valterova & Rasmont, 399 2015c). However, even if unique haplotypes (Fig. 2B) are found in *daghestanicus* (COI gene), there were not strongly supported as reciprocally monophyletic group in our phylogeny (Fig. 400 2A). No private haplotype was found in PEPCK gene for this taxon (Fig. 3B, C). Moreover, 401 inside the "sylvarum group", high probabilities (>0.05) to be conspecific is showed by the 402 403 bGMYC analyses (Fig. 4). On the other side, the CLGS did not highlight any divergence between the different populations which would imply that all of them could recognize each 404 405 other as sexual partners.

406 B. ruderarius is also a widespread species and its distribution is almost identical to that of B. sylvarum. However, B. ruderarius is absent from Sicily and the southern Italy but the 407 subspecies B. ruderarius tunensis occurs in the east of Algeria and Tunisia (Rasmont et al., 408 409 2015a). The same configuration as in B. sylvarum has been highlighted. Indeed, the divergence in color patterns seems to be a simple intraspecific variation and no diagnostic 410 character was found to discriminate significantly the different population inside *B. ruderarius*. 411 The unique haplotypes found in COI and PEPCK (Fig. 2B, 3B, C) for simulatilis were not 412 413 strongly supported as reciprocally monophyletic group in our phylogenies (Fig. 2A, 3A) as 414 well in our bGMYC analyses (Fig. 4). However, the CLGS of this Turkish population differ 415 from other populations and it is well supported in our cluster analysis (Fig. 5) but share main compounds (hexadec-9-en-1-ol, tricos-9-ene + tricos-7-ene) with the other B. ruderarius taxa 416 417 (supplementary file 2). We can therefore hypothesize that this divergence involved in the recognition system is not sufficient to induce a pre-mating reproductive barrier. This 418 419 geographic differentiation sharing main compounds could suggest only a dialect between populations as already described in moths (Toth et al., 1992) and solitary bees (Vereecken, 420 Mant & Schiestl, 2007). Although the taxon simulatilis seems more advanced on the path of 421

speciation than *daghestanicus*, complementary tests based on ethological behavior are needed.
We then hypothesize that these results could reflect the recent divergence of these taxa.
Unfortunately, we have not been able to analyze the subspecies *citrinofasciatus* and *tunensis*respectively inside the "*sylvarum* group" and the "*ruderarius* group".

Turkey is divided by a diagonal belt which is a massive structure formed by continuous 426 mountains (Miocene-Pliocene) from the north-east towards the south-west. These mountains 427 428 isolated populations from east and west in many species of insects or plants (Ciplak, Demisroy & Bozcuk, 1993; Ciplak, 2003; Ekim & Güner, 1986). Moreover, the distribution 429 of taxa from the West of this barrier generally spread through Europe while the taxa from the 430 431 east spread through Iran and Asia. In addition with this geographical particularities, climatic 432 change during the ice ages pushed many northern populations especially those found in the Balkans and Caucasus to the south and led to their isolation and speciation at higher altitudes 433 434 (Hewitt, 1996, 2000). These two taxa (B. ruderarius simulatilis and B. sylvarum daghestanicus) are known to occur in the east part of the diagonal belt and may have 435 colonized Turkey from Caucasus and Asia while the taxa to the west of this barrier may have 436 colonized Turkey from Balkans. It is also interesting to note that B. sylvarum daghestanicus 437 and B. ruderarius simulatilis are characterized by the same color pattern (white bands and red 438 439 tail) (Fig. 1D, E) which could be explained by a local Müllerian mimicry processes (Hines & Williams, 2012). Other species from other subgenus also posses the same color pattern in this 440 region and could confirm this process: B. cullumanus apollineus (Cullumanobombus), B. 441 442 sicheli cazuroi and B. incertus (Melanobombus), B. handlirschianus (Mendacibombus), B. brodmannicus (Pyrobombus) and B. niveatus (Sibiricobombus) (Rasmont & Iserbyt, 2014). 443

444 *Consider GAPS or not as phylogenetic information?*

445 Our PEPCK gene fragment contains gaps sites (representing an insertion or deletion and also

446 called "indels") and our statistical method for phylogenetic estimation (ML and MB) consider

the gap as missing data. Indeed, in most of taxonomic studies, the indels are treated as missing 447 448 data as this is the case for birds (Alström et al., 2015; McKay et al., 2014), frogs (Glaw, Kohler, De la Riva, Vieites & Vences, 2010), mammals (Ndiaye et al., 2014), spiders (Rezac, 449 Gasparo, Kral & Heneberg, 2014). Considered indels will increase the phylogenetic 450 information even if they are located in introns (in our case, all indels are located in the first 451 introns of the PEPCK gene fragment). These gaps reflect the evolutionary history of the taxa 452 453 and thus contain phylogenetic information which could be very informative particularly for simulatilis where the divergence estimation with all other ruderarius populations remains 454 unclear. It's seems therefore that the treatment of indels has not been fully investigated in 455 456 taxonomic studies and delimitaion of species. Moreover, in our study, haplotype network analysis considering or not the indels do not bring us any additional information except for *B*. 457 velox which not differs from B. ruderarius if we not considered indels in our analysis (Fig. 458 459 3B). Indeed, only gap sites allow to separate B. velox from B. ruderarius. However, both species are respectively strongly differenciated by the genitalia and other morphological 460 characters. 461

462 *Conservation implication*

Our integrative approach brings to attention the relevance of geographically isolated groups 463 464 differentiated in genetic and reproductive traits. This strict approach allows defining taxonomic status (species or subspecies) and providing a decision framework for policy-465 makers and conservation organizations. Except for B. inexspectatus (Rasmont et al., 2015b) 466 467 considered as endangered species in the International Union for Conservation of Nature (IUCN) Red List, B. mlokosievitzii as data deficient (Nieto et al., 2014) and B. velox who is 468 not yet assessed, the other taxa (B. ruderarius, B. sylvarum and B. veteranus) are all 469 considered as least concern by Nieto et al. (2014). Our new taxonomical hypotheses taking 470 into account two subspecies (B. ruderarius simulatilis and B. sylvarum daghestanicus) have 471

472 no implications for the red list assessments. However, the conservation of subspecies with
473 unique haplotypes as well as CLGS differentiation should be seen as source of diversity
474 within respective species and then should be considered and integrated in future management
475 plan.

476

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493 **<u>References</u>**

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760	Figures



Figure 1. Photos of some studied bumblebee species: A) *Bombus inexspectatus* male from
Alps, Italy; B) *B. mlokosievitzii vogtiellus* male from Turkey ; C) *B. ruderarius ruderarius*and *B. ruderarius montanus* from Pyrenees, France; D) *B. ruderarius simulatilis* female from
Turkey; E) *B. sylvarum daghestanicus* male from Turkey; F) *B. sylvarum sylvarum* female
from Pyrenees, France; G) *B. sylvarum nigrescens* male from Sweden; H) *B. velox* male from
Turkey and I) *B. veteranus* from the Netherlands. All black lines correspond to a scale of 1

dahlbomii

cm. All photographs are by P. Rasmont. J) Phylogeny of *Thoracobombus sensu stricto*. The
red box shows the monophyletic group of all species included in this study (according to
Hines & Cameron 2010).



Figure 2. A: Majority rule (50%) consensus tree based on maximum likelihood analyses of
COI. Values above tree branches are maximum likelihood bootstrap values/Bayesian posterior

- probabilities. **B**: Median-joining network of haplotypes based on cytochrome oxidase 1 (COI)
- sequences. Numbers on the lines correspond to the number of mutation between haplotypes.



Figure 3. A: Majority rule (50%) consensus tree based on maximum likelihood analyses of
PEPCK. Values above tree branches are maximum likelihood bootstrap values/Bayesian

Median-joining network 781 posterior probabilities. **B**: of haplotypes based on phosphoenolpyruvate carboxykinase (PEPCK) sequences. This network do not consider the 782 Gap sites. C: Median-joining network of haplotypes based on phosphoenolpyruvate 783 carboxykinase (PEPCK) sequences. This network consider the Gap sites. Numbers on the 784 lines correspond to the number of mutation between haplotypes. 785



Figure 4. bGMYC results based on COI phylogenetic tree. The vertical and horizontal lines correspond to the eight different groups shown on the tree. The scale corresponds to the probability to be conspecific.

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Figure 5. Unweighted pair group method with arithmetic mean (UPGMA) cluster based on a
Canberra distance matrix calculated from the cephalic labial gland secretions matrix of *velox*, *veteranus, inexspectatus, ruderarius, montanus, simulatilis, mlokosievitzii, sylvarum, daghestanicus, nigrescens and rogenhoferi*. The values near the node are multiscale bootstrap
resampling.