

Quantitative Measurement of Rare Earth Elements in Brines: Isolation from the Charged Matrix Versus Direct LA-ICP-MS Measurements – A Comparative Study

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| 1 | Quantitative Measurement of Rare Earth Elements in Brines: Isolation from the |
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| 2 | Charged Matrix Versus Direct LA-ICP-MS Measurements, a Comparative Study |
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| 4 | Maria A. Kokh ^{a,b,*} , Béatrice Luais ^c , Laurent Truche ^b , Marie Christine Boiron ^a , |
| 5 | Chantal Peiffert ^a and Aimeryc Schumacher ^c |
| 6 | |
| 7 | ^a Université de Lorraine, CNRS, GeoRessources, F-54000 Nancy, France |
| 8 | marie-christine.boiron@univ-lorraine.fr, chantal.peiffert@univ-lorraine.fr |
| 9 | ^b Institut des Sciences de la Terre (ISTerre) - UMR 5275 of CNRS, Université Grenoble Alpes, 1381 |
| 10 | rue de la Piscine, B.P. 53, 38041 Grenoble, France <u>laurent.truche@univ-grenoble-alpes.fr</u> |
| 11 | ^c Université de Lorraine, CNRS, CRPG, F-54000 Nancy, France <u>luais@crpg.cnrs-nancy.fr</u> , |
| 12 | aimeryc@crpg.cnrs-nancy.fr |
| 13 | * Present address: Universität Potsdam, Institut für Geowissenschaften, Campus Golm, Haus 27, |
| 14 | Karl-Liebknecht-Str. 24-25, 14476 Potsdam, Germany <u>kokh@uni-potsdam.de</u> |
| 15 | |
| 16 | Corresponding author: laurent.truche@univ-grenoble-alpes.fr |
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Abstract

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The guantification of the REEs in natural rocks and fluids is important not only for the 26 27 sustainable use of geological resources but also for fundamental geological research. Due to significant improvements in experimental, modeling and analytical techniques, it was 28 29 discovered that the REEs could be efficiently mobilized and transported by hydrothermal fluids. 30 Thus, analyzing the REEs in fluids, especially in highly saline solutions, is a prerequisite to 31 understand their fate in hydrothermal fluids. An analytical methodology was established to determine the trace quantities of the REEs in aqueous solutions with compositions typical to 32 seawater and brines. The developed protocol was applied for the REEs quantification in 33 experimental sodium-rich carbonate-bearing hydrothermal solutions. For this purpose, we used 34 35 two analytical approaches: i) the REEs were chromatographically isolated from the charged matrix and introduced in a single collector quadrupole ICP-MS, and ii) direct analysis of the REEs 36 by LA-ICP-MS contents of experimental solutions loaded into glass capillaries without any 37 additional isolation procedure. Both LA-ICP-MS and solution nebulization ICP-MS analysis after 38 39 the REEs isolation provide identic data in terms of the REEs concentrations in brines and are consistent within analytical uncertainties. Being an express method, LA-ICP-MS could be 40 41 recommended for the REEs quantification in brines. At the same time, the REEs' isolation from 42 the charged matrix with subsequent ICP-MS analysis provides higher accuracy and precision, giving additional information for the REEs mass fractions that are below the detection limit of 43 the LA-ICP-MS that was estimated at ~10 ng g-1 level. 44

45

46 **Keywords**: rare earth elements, brine, cation-exchange resin, ICP-MS, LA-ICP-MS.

48 1. Introduction

49

Geological and industrial significance of precise quantification of the rare earth elements in hydrothermal brines

52

53 The rare earth elements (REEs) are crucial in many modern technologies, such as 54 mobile phones, screens, hard drives, electric vehicles, and play a key role in the development of 55 techniques related to the renewable carbon-free energy industry. The rise of demand for many items that require rare earth metals induced an increase in metal mining and stimulated 56 significant industrial efforts for their efficient recovery and recycling, as well as prospection of 57 new deposits. The quantification of the REEs in natural rocks and fluids is important not only for 58 59 the sustainable use of natural resources but also for the fundamental geological research: the rare earths are widely used as redox conditions proxies (e.g., Bau 1991, González-Álvarez and 60 Kerrich 2010), geothermobarometers (e.g., Gratz and Heinrich 1998, Heinrich et al. 1997, Pyle 61 et al. 2001), geochronometers using the ¹⁴⁷Sm-¹⁴³Nd and ¹⁴⁶Sm-¹⁴²Nd (e.g., Workman and Hart 62 2005, Boyet and Carlson 2006), ¹³⁸La-¹³⁸Ce (Tanaka and Masuda 1982) systems, and 63 geochemical tracers of various petrogenetic processes occurring in mantle, crust and seafloor 64 hydrothermal systems (e.g., Luais et al. 2009, Schmidt et al. 2010, Cole et al. 2014, Mercadier et 65 66 al. 2011, Doucelance et al. 2014, Chavagnac et al. 2018). For a long time, magmatism and sedimentary processes were considered as two main 67

forces controlling the distribution of the REEs on the Earth. However, during the last thirty
years, due to significant improvements in experimental, modeling and analytical techniques
(*e.g.*, Wood 1990, Bau 1991, Ghazy *et al.* 1993), it was found that the REEs could be efficiently
mobilized and transported by hydrothermal fluids as a result of both regional or contact

72 metamorphism (e.g., González-Álvarez and Kerrich 2010, Richard et al. 2013) and post-

73 magmatic hydrothermal events (Banks *et al.* 1994, Salvi and Williams-Jones 2006). These later

74 processes may lead to their redistribution, fractionation and/or focused concentration.

Geothermal brines were even considered as a potential source of the REEs for future industrial
extraction (Smith *et al.* 2017).

77 Hydrothermal mobilization and transport of the REEs (as well as of other metals) from 78 a parent magmatic body or metal-bearing rock is provided by water, preferentially in the form 79 of aqueous complexes with the most abundant ligands in the crust. The knowledge about this complexation is a key for the understanding of the transporting capacities of hydrothermal 80 fluids (e.g., Seward et al. 2014 and refs. therein). The first studies of the REEs solubilities in 81 hydrothermal fluids implied their complexation with chloride, fluoride and sulfate anions (e.g., 82 83 Banks et al. 1994, Salvi and Williams-Jones 2006), later acetate anions and hydrocarbons were found to be important for some specific geological environments (e.g., Lecumberri-Sanchez et 84 al. 2018); finally, there is growing evidence that carbonate ligands could be also involved into 85 the hydrothermal mobilization of the REEs (Fan et al. 2004, Liu et al. 2018, Vasyukova and 86 87 Williams-Jones 2018).

In our particular case, we study the role of carbonate and bicarbonate ligands in the 88 hydrothermal transport of the REEs. During this research, we carried out autoclave experiments 89 90 and faced the necessity of determining the trace quantities of the REEs in experimental products, including solutions with high mass fractions of sodium comparable with those in 91 92 seawater (up to 3.0 % m/m). Another area of our study is focused on monazite interaction with 93 hydrothermal brines with high concentrations of alkali and alkali earth metals originating from 94 seawater evaporites. These two types of experiments have led us to seek the best method for 95 the REEs quantification in solutions in terms of implementation time, cost of analyzes as well as

| 96 | accuracy, precision and limits of detection. Here, we compare two different strategies that |
|-----|--|
| 97 | could be applied for the REEs quantification using ICP-MS: direct liquid microvolume analysis |
| 98 | using the LA-ICP-MS versus the chromatographic separation of a sample matrix followed by |
| 99 | pneumatic nebulization ICP-MS analysis. |
| 100 | |
| 101 | 2. Overview of ICP-MS methods for REE quantifications in geological samples |
| 102 | |
| 103 | 2.1. ICP-MS for quantification of the rare earth elements in natural samples – alternatives, |
| 104 | advantages, and choice of mass-spectrometer configuration |
| 105 | |
| 106 | Since the development of inductively-coupled plasma-mass-spectrometry (ICP-MS) in |
| 107 | the early 1980s (Houk <i>et al.</i> 1980; Date and Gray 1981, 1983), this method is widely used to |
| 108 | determine the REEs in natural samples. As an alternative to the MS, the trace quantities of REE |
| 109 | could be also measured using inductively coupled plasma-atomic emission spectroscopy (ICP- |
| 110 | AES), instrumental neutron activation analysis (INAA) and high-performance ion |
| 111 | chromatography (HPIC). Another way of sample ionization, except for ICP that is commonly |
| 112 | used for the REEs analyses, is thermal ionization, in particular, isotope dilution-thermal |
| 113 | ionization mass spectrometry (ID-TIMS). Details about the advantages and negative aspects of |
| 114 | the listed techniques could be found elsewhere (<i>e.g.</i> , Ishikawa <i>et al</i> . 2003). Briefly, ICP-MS is a |
| 115 | method of choice for the REEs quantification if: i) a precise quantification of all the REEs |
| 116 | including the monoisotopic (Pr, Tb, Ho, and Tm) is needed, ii) the REEs mass fractions in samples |
| 117 | are expected to be at the level of 1 μg g-1 and below, iii) samples are unique and limited |
| 118 | quantities of material are available for analyses, and iv) the nuclear reactor facilities are not |

accessible: nowadays INAA is practically replaced by ICP-MS due to its complexity andassociated costs.

121 In our case, we used a single collector quadrupole based ICP-MS unit either coupled with a laser ablation unit, or equipped with traditional nebulization and spray chamber, in order 122 to compare the results of the REEs measurements with the two different ways of sample 123 124 solution introduction into ICP. Here, we compare the direct laser ablation of experimental 125 solutions loaded into glass capillaries and the classical injection of experimental solutions 126 aerosols after their matrix isolation using column chromatography with strong cation-exchange. Alternative ICP-MS systems, such as sector-field (sf-ICP-MS) or time of flight (TOF) systems that 127 both can have either single or multiple collector detection system (e.g., Sindern 2017) are out of 128 129 the scope of the present study. 130 By using single collector quadrupole ICP-MS we assume, that the ratio of stable isotope of the REEs in our samples have a chondritic isotope composition, thus, only one isotope of 131 each REE can be acquired to obtain the element concentration in a sample. However, we 132 measured a few isotopes of the same REE (where available) during ICP-MS analysis to verify any 133 134 biais due to uncorrected interferences. 135 136 2.2. Sample preparation of rocks, minerals and natural fluids applied for ICP-MS analyses 137 of the Rare Earth Elements 138 During previous studies of the REEs quantification by ICP-MS, two main analytical 139 140 issues were identified: 1) low abundances of the REEs in some natural objects that approach the

141 detection limits of ICP-MS method (*e.g.,* sub ng g-1 level in residual ultramafic rocks; Nakamura

and Chang 2007; Ulrich et al., 2012), and 2) a strong matrix effect. High concentrations of matrix

elements in solutions (essentially Si, F and NaCl-rich solutions obtained from digested quartz
after crush-leach procedures) induce an enhancement or a suppression of the analytical signal,
(Ghazy *et al.* 1993). For example, Eaton *et al.* (1992) detected a progressive signal suppression
as the salt (NaCl) concentration in the sample solution was increased. These findings
determined the main sample preparation procedures for the REEs analysis by ICP-MS: matrix
elimination and target elements preconcentration followed by an introduction of sample
solution into ICP.

150 The ion exchange resins are commonly used in chromatography and ICP-MS analysis for element separations. Conventional ion exchange resins consist of a cross-linked polymer 151 matrix with a relatively uniform distribution of ion-active sites. A cross-link is a bond that links 152 153 one polymer chain to another. The degree of cross-linking (DC), expressed quantitatively as the 154 mole percent, relates to the number of groups that interconnect the polymerization mixture. The DC affects the kinetics of the cation-exchange process (Michaud 2011, Kołodyńska et al. 155 2019). The resin AG 50W[®] is commonly used for REEs isolation from the matrix. It is composed 156 of sulfonic acid functional groups attached to a styrene-divinylbenzene (S/DVB) copolymer 157 158 lattice: the strands of polystyrene (S) are held together by DVB and X8 or X12 means DC = 8% or 159 DC = 12%, respectively (here, the mole percent of DVB in the (S/DVB) copolymer lattice). Luais et al. (1997) used a column of strong cation-exchange resin AG 50W®X12 in HCl acid media for 160 161 high-precision isotope ratios measurements of neodymium and samarium in basalt samples (Reunion Island, Indian Ocean) using a multiple collector ICP-MS (MC-ICP-MS). This method 162 163 allowed to isolate Heavy Rare Earth Elements and Yttrium (HREEs+Y) from the light and medium 164 REEs (LREEs+MREEs). In the subsequent studies the REEs separation protocol mentioned above 165 was combined with the other procedures of element separation to study the behavior of 166 multiple isotopic systems during fluid-related deformation zone of leucogranite from the French

167 Massif Central (Luais et al. 2009) and metamorphic dehydration-hydration in eclogite and amphibolite samples from the Western Gneiss Region in Norway (Martin et al. 2010). 168 169 Cation exchange resins methods before REE+Y measurements using ICP-MS techniques is 170 required in some specific cases for precisely and accurately determine REE+Y contents, such as 171 accurate measurements of geological reference materials (JGb2 leucogabbro and JR3 172 peralkaline rhyolite from the Geological Survey of Japan) (Lee et al. 2000). This also ensures a 173 pre-concentration of REE in highly depleted lithologies, such as ultra-depleted in REEs 174 peridotites (Ulrich et al. 2012). or REE separation from highly charged matrix such as salts (Steinman et al. 2001). Halizc et al. (1996) similarly, developed an on-line preconcentration 175 method for ICP-MS to quantify the REEs in seawaters and highly saline brines. 176 177 Here, on the basis of previous analytical developments acquired for solid samples in 178 CRPG (Nancy, France) and ISTerre (Grenoble, France) laboratories, we customize an offline isolation protocol for brines, with highly-charged matrix, typical to our experimental systems. 179 180 2.3. LA-ICP-MS as a tool for express quantification of the REEs in solutions 181 182 LA-ICP-MS analyses of the REEs in minerals and fluid inclusions have already been 183 performed successfully to tackle scientific issues such as abundance and fractionation of the 184 185 REEs in natural minerals and fluid inclusions (Leisen 2011), genesis of uranium deposits (Mercadier et al. 2011, Lach et al. 2013), geochronometric dating of rocks and minerals using 186 the radioactive REE isotopes ¹³⁸La, ¹⁴⁷Sm and ¹⁷⁶Lu (Sindern 2017), mineralogy of the REEs in 187 188 coal fly ash (Thompson et al. 2018), partial resettling of the U-Th-Pb geochronometers during 189 monazite recrystallization (Grand'Homme et al. 2018), adsorption of the REEs on aluminum and

iron (oxy)-hydroxides in bauxites (Mondillo *et al.* 2019) and variety of fluids origin in iron oxidecopper-gold ore deposits revealed by the REEs signatures in fluorites (Schlegel *et al.* 2020).

192 LA-ICP-MS is widely used in geosciences due to the unique combination of its spatial resolution, fast acquisition of results and low detection limits which are supported by 193 194 reasonable analytical costs. Spatial resolution is critical for some minerals and fluid inclusion 195 assemblages that have micrometer to millimeter scale, overgrowths and zoning of minerals 196 related to successive geological events. Thus, a spatially resolved analysis at an appropriate 197 scale is the only way to determine the evolution of the mineral composition through time and thereby decipher multiple processes related to geochemical evolution during single crystal 198 growth (*e.g.*, Lach *et al*. 2013). 199

200 The use of LA-ICP-MS for solutions historically originates from the challenging studies 201 of mineral fluid inclusions, that are complicated by very small quantities of solution in individual fluid inclusions typically ranging from 10⁻¹³ to 10⁻⁶ g (Roedder 1984). However, it was shown 202 that this method could be also used for a small quantity of any aqueous solution (e.g., standard 203 204 or experimental) loaded into pure silica capillary that could be then ablated by laser like a fluid 205 inclusion (Leisen et al. 2012a,b, Harlaux et al. 2015, Grand'Homme et al. 2018). The matrix 206 effects for elemental analyses using LA-ICP-MS were widely discussed in literature (e.g., Günther et al. 1998, Velásquez et al. 2012, Leisen et al. 2012a, Grand'Homme et al. 2018). 207 208 Liquid samples can be quantitatively analyzed by their introduction into glass capillaries without any additional chemical treatment using NIST SRM glasses as reference materials and precisely 209 210 quantified sodium concentrations as an internal standard (Günther et al. 1998). This procedure, 211 which uses solid reference materials to quantify the REEs mass fractions in aqueous solution, is 212 carefully validated in this study.

- **3.** Experimental

3.1. Experimental conditions and setup of hydrothermal reactors

| 218 | The solubilities of the REEs were studied in the following carbon-bearing and carbon- |
|-----|---|
| 219 | free systems at 100, 150, 200, and 250°C and at pressures ranging from 20 to 150 bar (Table 1). |
| 220 | Hydrothermal reactors were developed to combine injection and sampling of solutions in a |
| 221 | multi-phasic system at high temperatures and pressures with the monitoring of pH in-situ. The |
| 222 | detailed description of the experimental setup could be found elsewhere (e.g., Truche et al. |
| 223 | 2016). Briefly, we used commercial bolt-closure reactors (Parr Instrument \mathbb{G}) made of pure |
| 224 | titanium (grade 3). The body and the head of the reactors were sealed with a PTFE gasket. The |
| 225 | maximum working temperature and pressure of the reactors were 300°C and 200 bar, |
| 226 | respectively. The internal volumes of the vessels varied from 250 to 500 ml. The temperature |
| 227 | was monitored using an immersed thermocouple and was held constant within $\pm 1^\circ$ C by an |
| 228 | electric furnace. The vessel was equipped with several connections that enabled regular |
| 229 | sampling and injection of fluids, as well as adjustment of the working pressure. |
| 230 | |
| 231 | Table 1 . Summary of the experiments with the REE oxides conducted in this study. |
| 232 | |
| 233 | The reactors were filled to two-thirds of their volume with an experimental solution. |
| 234 | The solution and the pressure vessel were bubbled and flushed with argon for half an hour to |
| 235 | remove dissolved nitrogen and oxygen. After flushing with argon, the autoclaves were leak |
| 236 | checked at ambient temperature at a pressure of 60 bar. The experimental solutions and solids |
| 237 | were stirred at 200 rpm by an impeller driven by an external magnetic driver. Aliquots of |

solution (2-3 g) were regularly sampled during the experiments. The first sample during one
sampling session served to remove the dead volume from the sampling tubes and to measure
the pH at ambient conditions. The second sample was filtered through a 0.2 µm PTFE filter and
acidified with 2 wt. % HNO₃ solution (dilution factor 10) for the subsequent ICP-OES analyses of
sodium and determination of the REEs using LA-ICP-MS and ICP-MS.

243

3.2. Chemical reagents used for the hydrothermal experiments

245

The initial solid phases of the REEs used for the hydrothermal experiments were the 246 following oxide powders: yttrium oxide Y₂O₃ (Rare Earth Products Ltd., 99.999 % m/m, CAS № 247 1314-36-9), lanthanum oxide La₂O₃ (Sigma-Aldrich, 99.99 % m/m, CAS № 1312-81-8), cerium 248 249 oxide CeO₂ (Prolabo Rhône-Poulenc Ltd., 99.9 % m/m, CAS № 1306-38-3), praseodymium oxide Pr₂O₃ (Rhône-Poulenc Industries, 99.99%, CAS № 12036-32-7), neodymium oxide Nd₂O₃ (Sigma-250 251 Aldrich, 99.9 % m/m, CAS № 1313-97-9), samarium oxide Sm₂O₃ (Sigma-Aldrich, 99.9 % m/m, CAS № 12060-58-1), europium oxide Eu₂O₃ (Rhône-Poulenc Industries, 99.99 % m/m, CAS № 252 253 1308-96-9), gadolinium oxide Gd₂O₃ (Sigma-Aldrich, 99.9 % m/m, CAS № 12064-62-9), dysprosium oxide Dy₂O₃ (Sigma-Aldrich, 99.99 % m/m, CAS № 1308-87-8), erbium oxide Er₂O₃ 254 255 (Sigma-Aldrich, 99.99 % m/m, CAS № 12061-16-4), ytterbium oxide Yb₂O₃ (Sigma-Aldrich, 99.99 256 % m/m, CAS № 1314-37-0). The mass of each oxide powder loaded into the autoclave was ~100 mg in all the experiments. Experimental solutions were prepared using the following salts: 257 258 Na₂CO₃ (Roth, ≥99,5 % m/m, ACS, anhydrous, CAS № 497-19-8) and NaHCO₃ (Sigma Aldrich, 259 ≥99.7 % m/m, ACS, anhydrous, CAS № 144-55-8). An aqueous solution of HNO₃ (2 % m/m) was 260 prepared from a bi-distilled acid.

3.3. Certified reference materials (CRM) used for ICP-MS, LA-ICP-MS and ICP-OES analyses
 and standard solutions preparation

264

All chemical procedures were carried out in clean room conditions. Preparation of
reactants, standard solutions, and dilutions were performed by weight and using high purity
18.2 MΩ cm MQ water (Elga Chorus system[®]) was used through this study. HCl and HNO₃ acids
were purified by double distillation using a Savillex[™] DST-1000 sub-boiling distillation system
(Savillex, USA).

Calibration curves for the REEs analyses by ICP-MS were established relative to
standard solutions containing 1, 10, 25 and 50 ng g-1 of the REEs in HNO₃ matrix (2 % m/m).
These standard solutions were prepared by weighting the certified CRM standard solution REE
mix TraceCERT® Co. LLC Sigma-Aldrich, 16 elements, 50 mg l-1 in 2 % m/m nitric acid. Standard
solutions containing 100 and 500 ng g-1 of the REEs were also prepared and analyzed as
additional standards to check the linearity of the obtained calibration curves, which was useful
for analyses of the most concentrated samples.

277 Standard solutions with high contents of matrix elements (sodium, potassium, magnesium and calcium) were prepared to establish the off-line quantification protocol for the 278 279 REEs separation from a charged matrix using cation-exchange resin. The following salts were 280 weighted with the REE mix and yttrium CRM solutions and Milli-Q water: MgCl₂·6H₂O (Prolabo), CaCl₂·2H₂O (Merck), NaCl (99.5 % m/m, Acros Organics), KCl (99.5 % m/m, Roth pa ACS ISO 281 282 Reagent). The prepared standard solutions were analyzed for major cations in SARM (CRPG, 283 Nancy) using ICP-OES Thermo Fischer ICap 6500. An analytical signal calibration of sodium 284 during ICP-OES analyses was performed using the sodium CRM for ICP (TraceCERT[®] Co. LLC 285 Sigma-Aldrich, 1000 mg l-1 Na in 2 % m/m nitric acid).

| 286 | Standard solutions containing 10 ng g-1, 100 ng g-1, 1 μ g g-1 and 10 μ g g-1 of the REEs |
|-----|---|
| 287 | in the matrix of aqueous NaCl solution (1 mol l-1) were prepared to test the accuracy of LA-ICP- |
| 288 | MS analyses. These standard solutions were prepared from the diluted REEs CRM (TraceCERT® |
| 289 | 16 elements mentioned above) and NaCl salt (99.5 % m/m, Acros Organics). Sodium was added |
| 290 | into the standard solutions to be used as an internal standard for all LA-ICP-MS analyses. |
| 291 | Sodium concentrations in standard solutions and in experimental samples were measured by |
| 292 | ICP-OES in SARM (CRPG, Nancy). The standard solutions with known amount of Na were loaded |
| 293 | into glass capillaries. |
| 294 | The CRM glasses NIST SRM 610 and NIST SRM 612 (matrix composition SiO $_2$ 72 % m/m, |
| 295 | Na ₂ O 14 % m/m, CaO 12 % m/m, Al ₂ O ₃ 2 % m/m) were used as calibrators for LA-ICP-MS. These |
| 296 | glasses are certified for a limited number of elements by NIST. Thus, following ISO guidelines |
| 297 | and the protocol of the International Association of Geoanalysts, the reference values for the |
| 298 | REEs in NIST SRM 610 and NIST SRM 612 were taken from Jochum et al. (2011). |
| 299 | |
| 300 | 3.4. Chromatographic isolation and ICP-MS analyses of the REEs in experimental solutions |
| 301 | |
| 302 | Polypropylen columns (Triskem [®] 10 cm high and 1.5 cm internal diameter) were filled |
| 303 | with AG 50W $^{\circ}$ cation-exchange resin (200-400 mesh, hydrogen form) with a degree of cross- |
| 304 | linking of 8 mol % (or 12 mol % for some calibration tests). Considering the large amount of |
| 305 | matrix with respect to the REEs contents (~60 g l-1 of chloride salts vs ~60 mg l-1 of the REEs for |
| 306 | the calibration tests), a resin volume of 20 ml was used in order to avoid resin saturation. The |
| 307 | matrix-rich REE standard solutions and sampled experimental solutions were slowly evaporated |
| 308 | to dryness at 90°C in Savillex \degree PTFE vials. The residue was dissolved in 5 ml of HCl (2.0 mol l-1) |
| 309 | and loaded onto resin pre-cleaned with alternately HCl (6 mol l-1) and MQ water and |

conditioned by 20 ml of HCl (2 mol l-1). After complete elution of sodium (or complete elution
of Na, Mg, K, and ~75 % m/m of Ca for the case of calibration tests) with 90 ml HCl solution (2.0
mol l-1), the fraction containing the REEs was collected in Savilex[®] Teflon vials with 155 ml of
HCl solution (6.0 mol l-1). The REEs fraction was evaporated to dryness at 90°C and redissolved
in 10 ml HNO₃ 2 % m/m solution for the subsequent ICP-MS analyses. All chromatographic
isolation and evaporation steps were made in cleanroom conditions.

316 The REEs mass fractions in solutions were measured using the quadrupole ICP-MS X-Series 2 ThermoScientific at CRPG laboratory (Nancy, France) (Table 2). The selected isotopes 317 for REE quantification, and corresponding instrumental detection and quantification limits 318 obtained after ten parallel measurements of matrix solution HNO₃ (2 % m/m) are reported in 319 320 Table 3. Detection limits for all the REEs were determined as the mass fraction equivalent of 321 three times the standard deviation of the procedural blank. They range from 0.1 pg g-1 for Lu to 5 pg g-1 for La and 20 pg g-1 for Y, and are better than published values published, e.g., from 1 322 to 120 pg g-1 (Ulrich et al. 2012). Our detection limits are significantly lower relative to the REE 323 mass fractions of all the experimental solutions analyzed in this work and are systematically 324 325 lower than the REE contents determined in procedural blanks after the use of the chemical protocol that is described below (Table 6). 326 327

328 **Table 2.** Instrumental operating parameters for ICP-MS.

329

Table 3. The detection limits (DL, 3σ) and the quantification limits (QL, 10σ) for the ICP-MS
analyses of the REEs, reported in pg g-1.

332

333 3.5. LA-ICP-MS analyses of experimental solutions

| 335 | LA-ICP-MS analyses of experimental solutions were carried out at the GeoRessources |
|-----|--|
| 336 | laboratory (Nancy, France). The LA-ICP-MS setup is composed of a 193 nm GeoLas Excimer laser |
| 337 | (ArF, Microlas, Göttingen, Germany), with a microscope for sample observation and laser beam |
| 338 | focus onto the sample, and an Agilent 7500c quadrupole ICP-MS. The experimental solutions |
| 339 | were loaded into pure silica capillaries (320 μ m int Ø, 435 μ m ext Ø, [®] Polymicro-Technologies). |
| 340 | For each experiment, the lanthanides, ⁸⁹ Y, and ²³ Na were analyzed. Data were collected with an |
| 341 | integration time of 0.01 s per channel for ²³ Na, ²⁸ Si, ²⁹ Si, 0.02 s for ⁸⁹ Y, ¹³⁹ La, ¹⁴⁰ Ce, ¹⁴¹ Pr, ¹⁴⁶ Nd, |
| 342 | ¹⁴⁷ Sm, ¹⁵³ Eu, 0.03 s for ¹⁵⁷ Gd, ¹⁵⁹ Tb, ¹⁶³ Dy, ¹⁶⁵ Ho, ¹⁶⁶ Er, ¹⁶⁹ Tm, ¹⁷² Yb, and ¹⁷⁵ Lu. Data were |
| 343 | acquired as raw counts per second (cps) using the time-resolved mode. For each analysis |
| 344 | (reference material and sample), the background was measured for 30 s, and the overall signal |
| 345 | acquisition was recorded for 100 s. The glass reference materials NIST SRM 610 and NIST SRM |
| 346 | 612 (Jochum et al. 2011) were used as reference materials, whereas sodium concentrations in |
| 347 | solutions preliminary quantified using ICP-OES were used as an internal standard (for details see |
| 348 | section 2.3). The spot size of 60 μm was used. The data reduction (quantification and limits of |
| 349 | detection) was carried out using Iolite software (Paton et al. 2011) based on the equations |
| 350 | developed by Longerich et al. (1996). More details on instrumental operating parameters are |
| 351 | reported in Table 4. |

- **Table 4.** Instrumental operating parameters for LA-ICP-MS.

358 **4.**

Results

359

360 4.1. Calibration tests and choice of the cation-exchange resin, validation of analytical 361 protocol of off-line sample preparation for ICP-MS analysis of the REEs

362

363 Seven tests of column calibration on standard solutions were performed during this 364 study. The off-line protocol for the REEs separation from sodium-rich matrix, e.g., type of resin, concentrations and volumes of acids, was established from calibration tests with simulated Na-365 K-Mg-Ca-rich standard solutions (Table 3). Using these solutions, we selected the type of cation-366 exchange resin, as well as molarities and volumes of HCl acids used for subsequent elution of 367 368 matrix and the REEs. The cation exchange columns were calibrated to obtain the best yields 369 (100±7%) for the REEs using minimal volumes of acids during the elution process (Fig. 1). The two AG 50W[®] resins (200-400 mesh) with 8 and 12 mol % cross-linkage were tested for this 370 purpose. The AG50W-X8 resin (8 mol % in DVB) was chosen because the use of AG50W-X12 (12 371 mol % in DVB) required more than 225 mL of HCl 6.0 mol l-1 solution to quantitatively collect all 372 373 the REEs from the column, rising issues concerning procedural blanks, sample throughput and 374 reactive consumption.

Here we report only the result of the last calibration with complete isolation of the REEs. A volume of 5 ml of the standard solution in HCl (2.0 mol l-1), containing MgCl₂ 5.18 g l-1, CaCl₂ 10.01 g l-1, NaCl 15.93 g l-1, KCl 25.38 g l-1, lanthanides and Sc 3.85 mg l-1, and Y 8.27 mg l-1, was loaded into the column. This matrix composition was chosen to simulate a composition of typical natural brine that always contains high concentrations of the alkaline and alkaline earth elements (see chapter 1.1 for details). Sodium, magnesium, and potassium were completely removed from the resin after the elution with 90 ml of HCl solution (2.0 mol l-1),

| 382 | whereas calcium was removed incompletely (76%) (Table 5, Fig. 1c). In our experimental study |
|-----|--|
| 383 | with carbonates, only sodium needed to be carefully isolated from the REEs, because its |
| 384 | preferential ionization compared with other elements can disturb their ionization, thus, so we |
| 385 | have not further refined the presented extraction protocol. The elution of the REEs with HCl |
| 386 | solution (6.0 mol l-1, 155 cm ³) was complete during the final calibration, in the order HREEs |
| 387 | (~40 ml), MREEs (~25 ml), and finally LREEs (~90 ml), with the REEs yields in the uncertainty |
| 388 | range of 100±7% (Table 3). A complete Ca extraction of calcium could be reached by an |
| 389 | increasing the HCl 2.0 mol I-1 volume of matrix elution, but not by increasing HCl molarity to 2.5 |
| 390 | mol I-1 that would induce early elution of HREEs in the matrix fraction. |
| 391 | |
| 392 | Fig. 1. a) Photo of cation exchange resin; a) schematic illustration of a cation-exchange process |
| 393 | in a column; a) elution of Na, Mg, K, Ca matrix with HCl 2.0 mol I-1 solution; b) subsequent |
| 394 | removal of the REEs during elution with HCl 6.0 mol I-1. |
| 395 | |
| 396 | Table 5. Results of the final calibration of cation-exchange resin column AG 50W®X8 (200-400 |
| 397 | mesh). |
| 398 | |
| 399 | Blank solutions of high purity 18.2 M Ω cm water and aqueous solutions of NaCl (1 mol |
| 400 | l-1) were treated like experimental samples to quantify the procedural blanks. The obtained |
| 401 | mass fractions of the REEs in the blank solutions varied from 2 to 650 pg g-1 (Table 6) |
| 402 | depending on the REE and could not affect the outcome of our analytical results that are at the |
| 403 | 10 ng g-1 level, with the exception of the lowest mass fractions of Ce, La, and Nd analyzed |
| 404 | during this study. The obtained mass fractions of these elements were corrected for procedural |
| 405 | blank levels. |

| 406 | | |
|-----|-----------------|--|
| 407 | Table 6. The | REEs mass fractions in blank and standard solutions after the use of a cation |
| 408 | exchange res | in column, reported in pg g-1. |
| 409 | | |
| 410 | The | analytical protocol for REEs separation from sodium for the subsequent ICP-MS |
| 411 | analyses esta | blished during this study includes the following procedures: |
| 412 | i) | weighing of the sample solutions; |
| 413 | ii) | evaporation of solutions to dryness at 90°C and residue dissolution in 5 ml of HCl |
| 414 | | 2.0 mol l-1; |
| 415 | iii) | preparation of the cation exchange resin columns filled with 10 cm ³ volume resin |
| 416 | | (AG [®] 50W-X8 200-400 mesh); |
| 417 | iv) | conditionning of the resin with HCl 2.0 mol I-1 (20 ml) |
| 418 | v) | loading of the prepared samples |
| 419 | vi) | add 90 ml of HCl 2.0 mol l-1 in each column for matrix elution |
| 420 | vii) | add 155 ml of HCl 6.0 mol l-1, for complete REE elution, and collection into Teflon |
| 421 | | vials . |
| 422 | viii) | evaporation of the HCl 6.0 N REE-bearing fraction to dryness, at 90°C on a hot |
| 423 | | plate. |
| 424 | ix) | redissolution of the dry residue in 10 ml of HNO_3 2 % m/m (ultrasonic bath); |
| 425 | x) | weighing of the obtained HNO ₃ solutions and calculation of dilution factors. |
| 426 | | |
| 427 | In or | der to estimate the accuracy of our off-line analytical protocol for the REEs |
| 428 | separation fro | om saline matrix described above, REE-bearing standard solutions doped with high |
| 429 | contents of s | odium, potassium, magnesium, and calcium (Table 6) were treated in the same |
| 430 | manner as ou | ir experimental samples. The measured concentrations of the REEs are in a |
| 431 | reasonable a | greement with the REEs concentrations of prepared standard solutions, with an |
| 432 | accuracy esti | mated within $\pm 10\%$ (3 SD) for the mass fraction range between 25 and 100 ng g-1. |
| 433 | This is similar | to the calibration test performed for standard solution (\pm 7%) (Table 5) for ~1 to |

434 ~10 µg g-1 REEs mass fractions. Our analytical protocol is validated for the lanthanides and yttrium measurements with a ±10 % accuracy, within a mass fraction range from ~10 ng g-1 to 435 ~10 µg g-1. The reported accuracy estimation is comparable with the repeatability of the ICP-436 MS analysis for the same solution, whereas RSD for an individual analysis typically varied 437 438 between 3 and 5%. 439 440 4.2. Accuracy and repeatability of LA-ICP-MS analyses of the REEs in solutions with simulated matrix loaded in glass capillaries, reproducibility of NIST SRM glasses 441 442 analyses 443 444 The accuracy of LA-ICP-MS analyses was verified by using standard solutions prepared 445 by weighing (see section 2.3). Typical time resolved LA-ICP-MS signals and the REEs responses during capillary ablation are shown in Fig. 2. Limits of detection for each REE, as well as chosen 446 isotopes, are reported in Table 7. Typical relative standard deviations (RSD, %) for an analytical 447 448 signal obtained on standard solutions during LA-ICP-MS measurements were found to be below 449 20% for the mass fraction range from 100 ng g-1 to 10 µg g-1 for all the REEs and about 50-60% 450 for a standard solution containing 10 ng g-1 of the REEs. Accuracy and repeatability between measurements obtained by multiple ablations of the same standard solutions are illustrated in 451 452 Fig. 3 and are reported in the Electronic Supplementary Material. Analyses of standard solutions

453 with the REEs mass fractions lower than 1 μ g g-1 are less accurate, with deviation from the

454 regression line towards lower measured values.

| 456 | Table 7 | . Mean values of detection (DL) and quantification (QL) limits acquired on standard |
|-----|------------------|--|
| 457 | solutior | ns during one analytical LA-ICP-MS session, in ng g-1. Note that for individual analyses, |
| 458 | the bes | t DL values could rich ~10 ng g-1. |
| 459 | | |
| 460 | Fig. 2. L | A-ICP-MS analyses of the REEs in standard solutions. |
| 461 | | |
| 462 | Fig. 3. R | Results of the REEs analyses in standard solutions obtained using LA-ICP-MS (results for |
| 463 | other R | EEs are available in the Electronic Supplementary Material). Note the deviation to lower |
| 464 | values f | or some standard solutions with REE mass fractions lower than 1 μg g-1. |
| 465 | | |
| 466 | | The both reference materials - NIST SRM 610 and NIST SRM 612 - were measured twice |
| 467 | before a | and after each analytical session. The NIST SRM 612 glass was quantified for the REEs as a |
| 468 | sample | certified sodium concentration in NIST SRM 612 glass served as an internal standard and |
| 469 | NIST SR | M 610 glass served as a calibrator. The average error between the two quantifications of |
| 470 | the REE | s in NIST SRM 610 and NIST SRM 612 reference materials vary from 0,1 to 7% for all the |
| 471 | REEs in | both glasses (Electronic Supplementary Material). |
| 472 | | |
| 473 | 5. | Discussion |
| 474 | | |
| 475 | 5.1. | Comparison of the REEs quantification in experimental solutions with two |
| 476 | | introduction systems for quadrupole ICP-MS measurements: laser ablation and |
| 477 | | pneumatic nebulization after matrix purification |
| 478 | | |
| 479 | | The mass fractions of rare earth elements (including Y) in the experimental solutions |
| | | |

| 480 | obtained using direct LA-ICP-MS and ICP-MS with pneumatic nebulizer (hereafter, pn-ICP-MS) |
|-----|--|
| 481 | after matrix isolation are reported in Tables 8-11 and graphically presented in Fig. 4. |

As a whole, the mass fractions of the REEs measured in acidified experimental solutions (Fig. 4) are in a very good agreement with pn-ICP-MS for the case of Ce, Pr, Eu, Er and Yb within associated analytical uncertainties (~18-22%). Yttrium, Nd, Sm and Dy were well quantified within their typical RSD and reproducibility values (~25-30%, Tables 8-11), whereas the data obtained for La and Gd are much less accurate (~47 and ~41% of the pn-ICP-MS values, respectively).

There are however few outliers for Ce. Indeed, few low Ce-samples (10-70 ng g-1) 488 show LA-ICP-MS mass fractions systematically lower than those obtained by pn-ICP-MS, with, 489 490 however, similar typical analytical uncertainties in terms of 3RSD (%) for both methods (Tables 491 8-11). Such deviation towards low Ce mass fractions is also seen for LA-ICP-MS Ce analyses of standard solutions of same concentration range (Fig. 3). Then, it is improbable that for the low 492 concentrations, cerium mass fractions are overestimated by pn-ICP-MS analyses after matrix 493 494 separation of sample solutions and blank correction. The calculated Ce mass fractions of those 495 samples by LA-ICP-MS, using the regression line equation in Fig. 4, would be in the range of (12-496 85 ng g-1). Then, the observed discrepancy is more likely due to Ce concentrations being close to the detection limit of the LA-ICP-MS. 497

498

Table 8. Fluid compositions of experimental samples in the H₂O–NaHCO₃–REEs system. LA-ICPMS data. Uncertainty (measurement repeatability) is expressed in terms of 3RSD.

501

Table 9. Fluid composition of experimental samples in the H₂O–NaHCO₃–REEs system. ICP-MS
 after ion-exchange resin Na-isolation. Uncertainty (measurement repeatability) is expressed in

504 terms of 3RSD.

505

Table 10. Fluid composition in the $H_2O-Na_2CO_3-REEs$ system. LA-ICP-MS data. Uncertainty (measurement repeatability) is expressed in terms of 3RSD.

508

Table 11. Fluid composition in the H₂O–Na₂CO₃–REEs system. ICP-MS after ion-exchange resin
Na-isolation. Uncertainty (measurement repeatability) is expressed in terms of 3RSD.

511

512 5.2. Analyzing REEs in solutions by LA-ICP-MS

513

The current practice of LA-ICP-MS analyses of the REEs in solutions using NIST SRM 514 glasses as reference materials and sodium as an internal standard was further verified in the 515 present study. Three types of matrix were ablated for the REEs analyses: synthetic silicate RM 516 517 glasses NIST SRM 610 and NIST SRM 612, NaCl-rich neutral brines and NaNO₃-rich acidified experimental solutions. The obtained results show that the presence of matrix effects for the 518 REEs analyses in solutions could not be totally excluded for using NIST SRM glasses 519 520 standardization, at least for the MS configurations with nanosecond laser. In NaCl-rich neutral brines all the REEs mass fractions analyzed with NIST SRM glasses are systematically 521 522 underestimated by 4 to 40% compared to the REEs mass fractions in prepared standard 523 solutions (Fig. 3). At the same time, for the case of acidified NaNO₃-rich experimental samples the obtained results are systematically overestimated if we assume the data of nb-ICP-MS 524 preceding by off-line sample preparation as real values (Fig. 4, Tables 8-11). Thus, the 525 526 concentrations of the REEs in solutions could be both systematically overestimated and

underestimated by LA-ICP-MS depending on the nature of the sample (here, essentially
presence of Si, Cl, N matrix elements and pH of analyzed solution).

The matrix effects on some REEs observed in this study sometimes exceed the 529 analytical uncertainties reported both in terms of RSD (%, 3o confidence level), reproducibility 530 of multiple ablations of the same solution and cross-analysis of the two NIST SRM glasses using 531 532 sodium as an internal standard. For example, the analyzed mass fractions of Er in NaCl-rich 533 neutral brines are systematically underestimated by ~40% of the values known from the preparation procedure of the standard solutions (Fig. 3). Such a high discrepancy for Er exceeds 534 both the estimated values of RSD (~30%, 3o confidence level) and reproducibility (~30%) from 535 parallel LA-ICP-MS analyses, at least for the mass fractions above 100 ng g-1. However, for the 536 537 same analytical session, the matrix effects for Nd, Sm and Gd are undistinguishable within the 538 reported analytical uncertainties.

The solution matrix effects for the REEs vary in a very individual manner: the same REE 539 540 could be well analyzed in one type of matrix using NIST SRM reference materials and be subjected to the matrix effect in another type of matrix as could be seen for Gd and Er (Table 541 542 12). Thus, the NIST SRM calibration method works, but some systematic variations that were 543 observed in the present study encourage us to recommend an additional calibration using the 544 REEs standard solutions with adjusted matrix type, loaded in glass capillaries. The precision of 545 analyses could be further improved by adding an isotope spike of a single element (e.g., Sm or Nd) to sample solutions by weight and quantifying a single element with isotope dilution after 546 LA-ICP-MS. Then this element can be used as precisely known internal standard to correct all 547 548 other REEs for matrix effects.

549

| 551 | Fig. 4. Comparison of the mass fractions of the REEs in solutions obtained using LA-ICP-MS and |
|-----|--|
| 552 | ICP-MS after sodium isolation. |

554 **6.** Conclusions

555

556 The key points of this study are the following:

557 The established analytical protocol for the REEs quantification by ICP-MS in brines after 558 matrix isolation with cation-exchange resin provides robust data for the REEs mass fractions as 559 low as the 1 ng g-1 level.

LA-ICP-MS analyses of solutions loaded into capillaries and classical pneumatic
 nebulization ICP-MS analyses after saline matrix isolation with cation exchange resin provide
 consistent results for the REEs quantification in brines in a range of mass fractions between 10
 ng g-1 and 10 µg g-1 within reported analytical uncertainties (RSD from 15 to 30% of a value for
 LA-ICP-MS). The precision level acquired during this study by LA-ICP-MS technique is acceptable

and sufficient enough for the purposes of the experimental hydrothermal geochemistry.

566 LA-ICP-MS, being an express method, could be highly recommended for the REEs 567 quantification in brines: in a single day of LA-ICP-MS analyses it is possible to measure up to 100 capillaries with REE solution, while it takes one month of cleanroom REE isolations to produce 568 569 these analyses with classical ICP-MS with pneumatic nebulizer. At the same time, the REEs 570 isolation from the charged matrix with subsequent ICP-MS analysis provides more precise 571 results (±10% a value within 3SD confidential level), and is required for the REEs mass fractions 572 that are below the quantification limit of the LA-ICP-MS that was estimated at the ~100 ng g-1 573 level. These results provide a framework for the selection of analytical methods in future 574 studies, depending on the required accuracy of REE quantification.

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| |

Tables

| Run | Initial composition, | Temperature, | Pressure, | Calculated pH_{TP} | Total duration of |
|-----|----------------------|--------------|-----------|----------------------|-------------------|
| IN- | | 0 | bai | | experiment, days |
| 2 | Na₂CO₃ 0.55 m | 100 | 19-24 | 10.3 | 15 |
| | | 150 | 18-22 | 9.9 | |
| | | 200 | 23-25 | 9.7 | |
| | | 250 | 40-42 | 9.6 | |
| 3 | NaHCO₃ 0.63 m | 100 | 22-35 | 7.6 | 17 |
| | | 150 | 32-35 | 7.6 | |
| | | 200 | 66-70 | 7.8 | |
| | | 250 | 140-150 | 8.0 | |

Table 1. Summary of the experiments with the REE oxides conducted in this study.

Table 2. Instrumental operating parameters for ICP-MS.

| ICP-MS model | X-Series 2 ThermoScientific |
|------------------------|---|
| Туре | Quadrupole |
| RF Power | 1400W |
| Cool gas flow rate | 13.5 l min ⁻¹ Ar |
| Auxilary gas flow rate | 0.82 l min ⁻¹ Ar |
| Sample gas flow rate | 0.97 l min ⁻¹ Ar |
| CeO/Ce | <0.03 |
| Ba++/Ba | <0.035 |
| Nebuliser | Microflow PFA |
| Spray chamber | PC ³ ESI Peltier Chiller, cooled to 2°C |
| Torch | Quartz glass torch |
| Sample Cone | Ni 4450 |
| Skimmer Cone | Ni PS8R |
| Detector mode | Dual (pulse and analogue counting) |
| Dwell time / mass | Variable (10-15 ms) |
| Mass resolution | Standard |
| Solution uptake | peristaltic pump 12 rpm |
| Acquisition Mode | Peak Jumping |
| Number of sweeps | 55 |
| Channels | 5 |
| Number of repeats | 3 |
| Calibration | Indium as internal standard with on-line injection |
| | Standard solution control every five measurements |
| Isotopes | ⁸⁹ Y, ¹³⁹ La, ¹⁴⁰ Ce, ¹⁴¹ Pr, ¹⁴³ Nd, ¹⁴⁶ Nd, ¹⁴⁷ Sm, ¹⁴⁹ Sm, |
| - | ¹⁵¹ Eu, ¹⁵³ Eu, ¹⁵⁷ Gd, ¹⁵⁹ Tb, ¹⁶¹ Dy, ¹⁶³ Dy, ¹⁶⁵ Ho, ¹⁶⁶ Er, |
| | ¹⁶⁷ Er, ¹⁶⁹ Tm, ¹⁷¹ Yb, ¹⁷² Yb, ¹⁷⁵ Lu |

Table 3. The detection (DL) and quantification (QL) limits for the ICP-MS analyses, in pg g-1.

| ⁸⁹ Y | ¹³⁹ La | ¹⁴⁰ Ce | ¹⁴¹ Pr | ¹⁴³ Nd | ¹⁴⁶ Nd | ¹⁴⁷ Sm | ¹⁴⁹ Sm | ¹⁵¹ Eu | ¹⁵³ Eu | ¹⁵⁷ Gd |
|-----------------|-----------------------------|--|--|--|--|--|--|--|--|--|
| 21 | 4.5 | 0.6 | 1.0 | 4.7 | 1.9 | 1.7 | 28 | 0.6 | 0.6 | 1.2 |
| 37 | 11.4 | 1.3 | 1.8 | 9.6 | 4.5 | 3.6 | 43 | 1.5 | 1.3 | 2.8 |
| | | | | | | | | | | |
| | ¹⁵⁹ Tb | ¹⁶¹ Dy | ¹⁶³ Dy | ¹⁶⁵ Ho | ¹⁶⁶ Er | ¹⁶⁷ Er | ¹⁶⁹ Tm | ¹⁷¹ Yb | ¹⁷² Yb | ¹⁷⁵ Lu |
| | 0.21 | 0.9 | 0.6 | 0.12 | 0.15 | 1.5 | 0.16 | 0.9 | 0.7 | 0.08 |
| | 0.48 | 1.8 | 1.3 | 0.28 | 0.38 | 2.9 | 0.36 | 2.2 | 1.8 | 0.21 |
| | ⁸⁹ Y 21 37 | ⁸⁹Y ¹³⁹La 21 4.5 37 11.4 ¹⁵⁹Tb 0.21 0.48 | 89Y 139La 140Ce 21 4.5 0.6 37 11.4 1.3 159Tb 161Dy 0.21 0.9 0.48 1.8 | 89Y 139La 140Ce 141Pr 21 4.5 0.6 1.0 37 11.4 1.3 1.8 159Tb 161Dy 163Dy 0.21 0.9 0.6 0.48 1.8 | 89Y 139La 140Ce 141Pr 143Nd 21 4.5 0.6 1.0 4.7 37 11.4 1.3 1.8 9.6 159Tb 161Dy 163Dy 165Ho 0.21 0.9 0.6 0.12 0.48 1.3 0.28 | 89Y 139La 140Ce 141Pr 143Nd 146Nd 21 4.5 0.6 1.0 4.7 1.9 37 11.4 1.3 1.8 9.6 4.5 159Tb 161Dy 163Dy 165Ho 166Er 0.21 0.9 0.6 0.12 0.15 0.48 1.8 1.3 0.28 0.38 | 89Y 139La 140Ce 141Pr 143Nd 146Nd 147Sm 21 4.5 0.6 1.0 4.7 1.9 1.7 37 11.4 1.3 1.8 9.6 4.5 3.6 159Tb 161Dy 163Dy 165Ho 166Er 167Er 0.21 0.9 0.6 0.12 0.15 1.5 0.48 1.8 1.3 0.28 0.38 2.9 | 89Y 139La 140Ce 141Pr 143Nd 146Nd 147Sm 149Sm 21 4.5 0.6 1.0 4.7 1.9 1.7 28 37 11.4 1.3 1.8 9.6 4.5 3.6 43 159Tb 161Dy 163Dy 165Ho 166Er 167Er 169Tm 0.21 0.9 0.6 0.12 0.15 1.5 0.16 0.48 1.8 1.3 0.28 0.38 2.9 0.36 | 89Y 139La 140Ce 141Pr 143Nd 146Nd 147Sm 149Sm 151Eu 21 4.5 0.6 1.0 4.7 1.9 1.7 28 0.6 37 11.4 1.3 1.8 9.6 4.5 3.6 43 1.5 159Tb 161Dy 163Dy 165Ho 166Er 167Er 169Tm 171Yb 0.21 0.9 0.6 0.12 0.15 1.5 0.16 0.9 0.48 1.8 1.3 0.28 0.38 2.9 0.36 2.2 | 89Y 139La 140Ce 141Pr 143Nd 146Nd 147Sm 149Sm 151Eu 153Eu 21 4.5 0.6 1.0 4.7 1.9 1.7 28 0.6 0.6 37 11.4 1.3 1.8 9.6 4.5 3.6 43 1.5 1.3 159Tb 161Dy 163Dy 165Ho 166Er 167Er 169Tm 171Yb 172Yb 0.21 0.9 0.6 0.12 0.15 1.5 0.16 0.9 0.7 0.48 1.8 1.3 0.28 0.38 2.9 0.36 2.2 1.8 |

881 Table 4. Instrumental operating parameters for LA-ICP-MS.

| Laser type | GeoLas Excimer |
|-------------------------|--|
| Wavelength | 193 nm |
| Laser frequency | 5Hz |
| Fluence | 7 J cm ⁻² |
| Ablation cell | Internal diameter 5.5 cm, useful volume 24 cm ³ |
| Ablation spot size | 60 μm |
| ICP-MS model | Agilent 7500cs |
| Туре | Quadrupole |
| RF Power | 1550W |
| Carrier gas flow rate | 0.92 l min ⁻¹ Ar |
| Vector gas flow rate | 0.50 l min ⁻¹ He |
| ThO/Th | <0.3 |
| Th/U | 0.98-1 |
| Torch | Quartz glass torch |
| Sample Cone | Ni G1820-65238 made for Agilent by Spectron, Inc. |
| Skimmer Cone | Ni G3270-65024 for 7500ce/cx, Agilent Technologies |
| Sampling depth | 5 mm |
| Detector mode | Dual (pulse and analogue counting) |
| Dwell time / mass | depending on the mass, see text |
| Background | 30 s |
| Acquisition time | 40 s (200 pulses) |
| Total time (1 analysis) | 100 s |
| Number of repeats | 6-7 |
| Calibration | Calibrator (NIST SRM 610,), internal (²³ Na) |
| Isotopes | ²³ Na, ⁸⁹ Y, ¹³⁹ La, ¹⁴⁰ Ce, ¹⁴¹ Pr, ¹⁴⁶ Nd, ¹⁴⁷ Sm, ¹⁵³ Eu, ¹⁵⁷ Gd, ¹⁵⁹ Tb, |
| | ¹⁶³ Dy, ¹⁶⁵ Ho, ¹⁶⁶ Er, ¹⁶⁹ Tm, ¹⁷² Yb, ¹⁷⁵ Lu |

882

883 Table 5. Results of the cation-exchange resin column calibration, chemical yield (%).

| Element | Υ | La | Ce | Pr | Nd | Sm | Eu | Gd | Tb | Dy | Но | Er | Tm | Yb | Lu | Mg | (| Са | N | а | K |
|--------------------------------|-------------|----------------------------|--------------|--------------|----------------|----------------|------|--------|------|-------|-------|------|-------|------|-------|---------|-------|--------|-------|-------|--------|
| Loaded, µg | 43 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 20 | 6 879 | 18 | 810 | 32 (| 608 | 69 246 |
| Measured, µg | 42 | 19 | 19 | 19 | 21 | 20 | 19 | 20 | 19 | 19 | 20 | 19 | 20 | 19 | 20 | 6 6 4 0 | 14 | 217 | 37 4 | 416 | 72 133 |
| Yield, % | 99 | 97 | 96 | 93 | 106 | 102 | 93 | 101 | 97 | 97 | 99 | 97 | 99 | 95 | 99 | 97 | 7 | 76 | 11 | 15 | 104 |
| and salts): M mg I-1, Y 8.2 | ИgĊ 27 n | l ₂ 5. ng l- | 18 g 1. V | l-1, olum | CaCl le: 5. | ₂ 10. 2 ml. | 01 g | I-1, I | NaCl | 15.9 | 93 g | I-1, | ŘCI 2 | 25.3 | 8 g l | -1, lar | ithan | nides | and | Sc 3 | 8.85 |
| Table 6. The I | REE | Es n | nass | s fra | ction | is in | the | blanl | k so | lutio | n (ir | ר pg | ı g-1 |) an | d st | andaı | rd so | olutic | ons (| (in n | g g-1) |
| the chemical t | rea | tme | nt u | sing | the | catio | on e | xcha | nge | resi | in A | G® (| 50W | -X8 | (20 | 0-400 |) me | esh) (| colu | mn. | |
| Sample/RE | E | | Y | La | С | e l | Pr | Nd | Sm | n E | u | Gd | Tb | D | y F | Ho E | r 1 | Tm | Yb | Lu | Sc* |
| Blank MQ-0, p | g g- | -1 | 32 | 446 | 64 | 8 1 | 04 | 258 | 39 | 17 | 79 | 19 | N/A | 8 | Ν | I/A 6 | 5 N | N/A | 15 | N/A | N/A |
| Blank MQ-1 n | a a- | .1 | 51 | 426 | 3 47 | 'A 6 | 31 | 234 | 44 | 10 | 76 | 48 | 52 | 32 | > 7 | 162 | 6 6 | 52 | 19 | 53 | 591 |

| Sample/REE | Y | La | Ce | Pr | INC | Sm | Eu | Ga | dl | Dy | но | Er | Im | YD | Lu | SC" |
|----------------------|-----|-----|-----|-----|-----|----|-----|----|-----|----|-----|----|-----|----|-----|-----|
| Blank MQ-0, pg g-1 | 32 | 446 | 648 | 104 | 258 | 39 | 179 | 19 | N/A | 8 | N/A | 6 | N/A | 15 | N/A | N/A |
| Blank MQ-1, pg g-1 | 51 | 426 | 474 | 61 | 234 | 44 | 106 | 48 | 5.2 | 32 | 7.6 | 26 | 5.2 | 19 | 5.3 | 591 |
| Blank MQ-2, pg g-1 | 164 | 89 | 278 | 41 | 195 | 41 | 18 | 42 | 4.3 | 38 | 8.3 | 24 | 4.3 | 24 | 5.2 | 500 |
| Blank salt-1, pg g-1 | 47 | 427 | 469 | 52 | 195 | 52 | 88 | 41 | 2.2 | 26 | 2.9 | 33 | 2.2 | 65 | 2.4 | 684 |
| Blank salt-2, pg g-1 | 167 | 114 | 313 | 41 | 221 | 43 | 22 | 45 | 3.6 | 41 | 8.2 | 28 | 3.6 | 25 | 4.9 | 454 |
| Standard 1, ng g-1 | 48 | 24 | 24 | 23 | 25 | 24 | 24 | 24 | 23 | 24 | 23 | 24 | 23 | 24 | 23 | 19 |
| Standard 2, ng g-1 | 106 | 47 | 48 | 46 | 54 | 50 | 48 | 50 | 51 | 50 | 51 | 48 | 51 | 50 | 52 | 39 |
| | | | | | | | | | | | | | | | | |

⁸⁹¹

- 892 Blank MQ - REE-free Milli-Q solution treated like a sample;
- 893 Blank salt – REE-free 1 mol I-1 NaCl solution treated like a sample;
- 894 Standard 1, reference mass fractions: REEs (lanthanides and Sc) 25.1 ng g-1, Y 50.9 ng g-1;
- 895 896 Standard 2, reference mass fractions: REEs (lanthanides and Sc) 51.0 ng g-1, Y 103.3 ng g-1. N/A – not analyzed; *Sc mass fractions were tentatively checked in blanks and standard solutions (yield 75-77%).
- 897

898 Table 7. Mean values of detection and quantification limits (DL and QL) acquired on standard solutions

899 during one LA-ICP-MS measurement session, in ng g-1.

| | ⁸⁹ Y | ¹³⁹ La | ¹⁴⁰ Ce | ¹⁴¹ Pr | ¹⁴³ Nd | ¹⁴⁶ Nd | ¹⁴⁷ Sm | ¹⁴⁹ Sm | ¹⁵¹ Eu | ¹⁵³ Eu | ¹⁵⁷ Gd |
|-------------------|-----------------|--------------------------------|---------------------------------|--------------------------------|--------------------------------|--------------------------------|---------------------------------|-------------------------------|---------------------------------|--------------------------------|-------------------------------|
| DL, 3σ | N/A | 28 | 32 | 24 | N/A | 140 | 250 | N/A | N/A | 37 | 180 |
| QL, 10σ | N/A | 93 | 110 | 78 | N/A | 470 | 820 | N/A | N/A | 120 | 590 |
| | | | | | | | | | | | |
| | | | | | | | | | | | |
| | | ¹⁵⁹ Tb | ¹⁶¹ Dy | ¹⁶³ Dy | ¹⁶⁵ Ho | ¹⁶⁶ Er | ¹⁶⁷ Er | ¹⁶⁹ Tm | ¹⁷¹ Yb | ¹⁷² Yb | ¹⁷⁵ Lu |
| DL, 3σ | | ¹⁵⁹ Tb 49 | ¹⁶¹ Dy N/A | ¹⁶³ Dy 75 | ¹⁶⁵ Ho 31 | ¹⁶⁶ Er 55 | ¹⁶⁷ Er N/A | ¹⁶⁹ Tm 24 | ¹⁷¹ Yb N/A | ¹⁷² Yb 88 | ¹⁷⁵ Lu 23 |
| DL, 3σ QL, 10σ | | ¹⁵⁹ Tb 49 160 | ¹⁶¹ Dy N/A N/A | ¹⁶³ Dy 75 250 | ¹⁶⁵ Ho 31 100 | ¹⁶⁶ Er 55 182 | ¹⁶⁷ Er N/A N/A | ¹⁶⁹ Tm 24 80 | ¹⁷¹ Yb N/A N/A | ¹⁷² Yb 88 293 | ¹⁷⁵ Lu 23 78 |

900

901 Table 8. Fluid compositions of experimental samples in the H2O-NaHCO3-REEs system. LA-ICP-MS

902 data. Uncertainty (measurement repeatability) is expressed in terms of 3RSD.

| Sample | Na, | Y | La | Ce | Pr | Nd | Sm | Eu | Gd | Dy | Er | Yb | |
|--------|-------------|--------|--------|-----|-----|-----|-------|-------|--------|--------|--------|---------|--|
| number | mol/kg soln | | ng g-1 | | | | | | | | | | |
| 3-0 | 0.63 | 6 845 | - | 3.6 | 43 | 421 | 1 827 | 4 447 | 17 695 | 2 460 | 325 | 1 523 | |
| 3-1 | 0.62 | 15 525 | - | 5.7 | 29 | 147 | 958 | 2 982 | 10 080 | 9 500 | 960 | 4 160 | |
| 3-2 | 0.65 | 53 945 | - | 12 | 132 | 228 | 706 | 3 412 | 6 315 | 37 360 | 13 865 | 55 650 | |
| 3-3 | 0.65 | 59 250 | - | 16 | 466 | 505 | 1 747 | 3 300 | 7 400 | 43 345 | 32 965 | 97 750 | |
| 3-4 | 0.63 | 55 510 | - | 12 | 240 | 344 | 1 472 | 3 323 | 7 665 | 40 905 | 33 605 | 101 750 | |
| 3-5 | 0.65 | 19 135 | - | 8.4 | 419 | 142 | 490 | 980 | 2 245 | 8 850 | 26 300 | 155 000 | |
| 3-6 | 0.67 | 14 380 | - | bdl | 287 | 90 | 361 | 852 | 2 059 | 8 620 | 21 620 | 104 067 | |
| 3-7 | 0.67 | 14 270 | - | bdl | 186 | 66 | 296 | 814 | 1 896 | 9 180 | 17 635 | 86 100 | |
| 3-8 | 0.60 | | | | | | N/A | | | | | | |
| 3-9 | 0.53 | 6 240 | - | bdl | 40 | - | 174 | 371 | 915 | 3 790 | 6 640 | 33 000 | |
| 3-10 | 0.52 | 5 800 | - | bdl | 68 | 56 | 194 | 389 | 947 | 3 740 | 6 400 | 27 520 | |
| 3-11 | 0.48 | 7 507 | - | 3.5 | 77 | 88 | 256 | 557 | 1 316 | 5 043 | 8 570 | 38 830 | |
| 3-15 | 0.71 | 7 640 | - | 28 | 539 | 557 | 1 433 | 1 883 | 3 505 | 7 575 | 7 395 | 23 650 | |
| 3RSD, | ±15 | ±25 | - | ±20 | ±20 | ±20 | ±25 | ±20 | ±25 | ±25 | ±25 | ±25 | |
| % | | | | | | | | | | | | | |

903

"bdl" – below detection limit; N/A – not analyzed. The samples N 3-12, 3-13, and 3-14 were not analyzed

904 905 906 because of the low concentration of sodium that was used as an internal standard for LA-ICP-MS. The

results of measurements with an unexploitable signal during laser ablation are not reported (-).

Table 9. Fluid composition of experimental samples in the H₂O–NaHCO₃–REEs system. ICP-MS after

| Sample | Na, | Y | La | Се | Pr | Nd | Sm | Eu | Gd | Dy | Er | Yb |
|------------|-------------|--------|-----|-----|-----|-----|-------|-------|--------|--------|--------|---------|
| number | mol/kg soln | | | | | | n | g g-1 | | , | | |
| 3-0 | 0.63 | 4 760 | 16 | 13 | 31 | 345 | 1 391 | 3 472 | 11 530 | 1 902 | 257 | 1 297 |
| 3-1 | 0.62 | 11 199 | 32 | 48 | 28 | 150 | 890 | 2 758 | 7 073 | 8 059 | 841 | 3 620 |
| 3-2 | 0.65 | 41 722 | 31 | 22 | 136 | 216 | 669 | 3 241 | 4 630 | 32 623 | 12 856 | 49 461 |
| 3-3 | 0.65 | 43 725 | 121 | 26 | 448 | 479 | 1 560 | 3 038 | 5 479 | 36 748 | 29 437 | 82 370 |
| 3-4 | 0.63 | 41 003 | 36 | 20 | 212 | 298 | 1 203 | 2 794 | 5 416 | 33 000 | 28 155 | 80 257 |
| 3-5 | 0.65 | 15 180 | 65 | 11 | 397 | 136 | 434 | 895 | 1 653 | 8 012 | 22 954 | 131 423 |
| 3-6 | 0.67 | 12 174 | 57 | 12 | 270 | 99 | 342 | 816 | 1 479 | 7 423 | 18 559 | 86 114 |
| 3-7 | 0.67 | 11 972 | 38 | 10 | 182 | 69 | 279 | 750 | 1 350 | 7 660 | 14 707 | 68 387 |
| 3-8 | 0.60 | 11 665 | 36 | 10 | 200 | 80 | 414 | 863 | 1 650 | 7 576 | 12 558 | 58 247 |
| 3-9 | 0.53 | 6 168 | 16 | 9.5 | 42 | 41 | 164 | 337 | 671 | 3 091 | 5 697 | 26 684 |
| 3-10 | 0.52 | 5 913 | 26 | 11 | 78 | 58 | 192 | 395 | 745 | 3 225 | 5 560 | 22 027 |
| 3-11 | 0.48 | 6 803 | 27 | 13 | 80 | 93 | 240 | 518 | 901 | 3 800 | 6 527 | 28 166 |
| 3-12 | 0.0051 | 946 | 22 | 19 | 22 | 22 | 43 | 96 | 188 | 643 | 508 | 997 |
| 3-13 | 0.0004 | 1 415 | 51 | 12 | 85 | 66 | 185 | 384 | 778 | 1 668 | 791 | 1 174 |
| 3-14 | 0.0058 | 149 | 13 | 15 | 2.7 | 11 | 12 | 24 | 41 | 73 | 37 | 58 |
| 3-15 | 0.71 | 5 102 | 86 | 32 | 383 | 393 | 869 | 1 292 | 2 158 | 5 394 | 5 440 | 16 476 |
| 3RSD, % | ±15 | ±20 | ±10 | ±10 | ±10 | ±10 | ±10 | ±10 | ±15 | ±15 | ±15 | ±15 |

910 ion-exchange resin Na-isolation. Uncertainty (measurement repeatability) is expressed in terms of 3RSD.

Table 10. Fluid composition in the H₂O–Na₂CO₃–REEs system. LA-ICP-MS data. Uncertainty

| Sample | Na, | Y | La | Ce | Pr | Nd | Sm | Eu | Gd | Dy | Er | Yb |
|---------|-------------|---------|-------|-----|-------|-------|--------|-------|---------|--------|--------|--------|
| name | mol/kg soln | | | | | | ng g- | 1 | | | | |
| 2-0 | 1.1 | 119 733 | 7 715 | 387 | 2 603 | 2 215 | 4 060 | 325 | 136 400 | 73 300 | 13 970 | 6 020 |
| 2-1 | 1.2 | 183 133 | 4 425 | 312 | 1 813 | 2 907 | 17 237 | 1 625 | 207 333 | 81 400 | 16 260 | 5 430 |
| 2-2 | 1.2 | 199 133 | 1 808 | 267 | 1 064 | 2 328 | 15 307 | 1 587 | 181 533 | 76 000 | 15 573 | 5 447 |
| 2-3 | 1.2 | 230 150 | 1 384 | 211 | 861 | 1 980 | 13 045 | 1 371 | 160 650 | 78 700 | 16 915 | 5 850 |
| 2-4 | 1.2 | 126 200 | 311 | 14 | 208 | 430 | 2 085 | 211 | 36 900 | 69 000 | 90 900 | 86 300 |
| 2-5 | 1.2 | 36 570 | 481 | 11 | 567 | 1 705 | 4 519 | 100 | 16 165 | 21 800 | 36 825 | 39 470 |
| 2-6 | 1.2 | 19 700 | - | bdl | 360 | - | 7 300 | - | 10 710 | 10 450 | 21 400 | 22 400 |
| 2-7 | 1.2 | 33 495 | 770 | 12 | 783 | 1 437 | 9 050 | 157 | 19 250 | 20 260 | 35 435 | 44 425 |
| 2-8 | 1.1 | 29 390 | 723 | 11 | 943 | 1 520 | 6 240 | 157 | 16 695 | 17 925 | 32 765 | 47 335 |
| 2-9 | 1.2 | 24 770 | 450 | 7.4 | 1 012 | 1 143 | 3 358 | 101 | 10 060 | 13 715 | 30 015 | 52 100 |
| 2-10 | 1.2 | 24 430 | - | bdl | 608 | - | - | - | 6 740 | 10 870 | 31 400 | 56 200 |
| 2-11 | 1.2 | 11 800 | - | bdl | 214 | 175 | 384 | 30 | 1 383 | 4 050 | 13 945 | 26 700 |
| 2-12 | 0.7 | 4 865 | - | bdl | 125 | 128 | 446 | 24 | 1 515 | 1 949 | 3 620 | 4 435 |
| 2-13 | 1.3 | 14 910 | - | bdl | 32 | 86 | 390 | 81 | 1 625 | 5 305 | 12 115 | 12 445 |
| 3RSD, % | ±15 | ±25 | ±45 | ±20 | ±20 | ±20 | ±25 | ±20 | ±25 | ±25 | ±25 | ±25 |

912 (measurement repeatability) is expressed in terms of 3RSD.

913 The results of measurements with an unstable signal during the laser ablation are not reported (-).

914 Table 11. Fluid composition in the H₂O–Na₂CO₃–REEs system. ICP-MS after ion-exchange resin Na-

| 915 | isolation. | Uncertainty | (measurement | repeatability) is | s expressed in | terms of 3RSD. |
|-----|------------|-------------|--------------|-------------------|----------------|----------------|
| | | | ` | 1 3/ | | |

| Sample | Na, | Y | La | Ce | Pr | Nd | Sm | Eu | Gd | Dy | Er | Yb |
|---------|-------------|---------|--------|-----|-------|-------|--------|-------|---------|--------|--------|--------|
| name | mol/kg soln | | ng g-1 | | | | | | | | | |
| 2-0 | 1.1 | 88 822 | 5 354 | 293 | 2 035 | 1 527 | 2 574 | 262 | 86 202 | 52 391 | 10 635 | 5 649 |
| 2-1 | 1.2 | 146 901 | 2 649 | 268 | 1 627 | 2 287 | 13 812 | 1 329 | 154 630 | 63 625 | 13 355 | 6 708 |
| 2-2 | 1.2 | 159 758 | 1 431 | 223 | 940 | 1 800 | 11 543 | 1 277 | 129 638 | 55 862 | 12 359 | 6 033 |
| 2-3 | 1.2 | 175 187 | 1 067 | 173 | 705 | 1 471 | 9 590 | 1 061 | 108 748 | 55 473 | 12 923 | 5 876 |
| 2-4 | 1.2 | 111 903 | 221 | 16 | 168 | 365 | 1 690 | 172 | 26 350 | 55 531 | 74 528 | 65 916 |
| 2-5 | 1.2 | 30 144 | 391 | 11 | 548 | 1 383 | 3 773 | 87 | 13 543 | 18 363 | 29 911 | 30 458 |
| 2-6 | 1.2 | 20 157 | 269 | 12 | 362 | 593 | 7 712 | 85 | 10 900 | 12 370 | 24 360 | 27 326 |
| 2-7 | 1.2 | 25 312 | 492 | 11 | 685 | 1 137 | 7 097 | 135 | 15 367 | 16 468 | 27 925 | 32 613 |
| 2-8 | 1.1 | 21 505 | 430 | 9.4 | 800 | 1 125 | 4 576 | 123 | 12 594 | 13 506 | 25 760 | 32 363 |
| 2-9 | 1.2 | 19 782 | 322 | 6.6 | 939 | 893 | 2 623 | 87 | 7 919 | 11 283 | 25 041 | 37 734 |
| 2-10 | 1.2 | 19 700 | 128 | 5.3 | 559 | 659 | 1 712 | 67 | 5 424 | 9 446 | 26 070 | 42 697 |
| 2-11 | 1.2 | 11 772 | 45 | 9.2 | 202 | 155 | 376 | 37 | 1 068 | 4 341 | 15 432 | 29 245 |
| 2-12 | 0.7 | 3 496 | 66 | 6.6 | 87 | 90 | 301 | 18 | 1 009 | 1 277 | 2 500 | 2 915 |
| 2-13 | 1.3 | 16 478 | 6.2 | 8.5 | 29 | 78 | 370 | 68 | 1 360 | 5 986 | 13 694 | 13 121 |
| 3RSD, % | ±15 | ±20 | ±10 | ±10 | ±10 | ±10 | ±10 | ±10 | ±15 | ±15 | ±15 | ±15 |

Table 12. Quantitative estimation of matrix effects observed on solutions during LA-ICP-MS.

| Matrix effect, % | Matrix salt | La | Ce | Pr | Nd | Sm | Eu | Gd | Dy | Er Yb |
|----------------------------|--|----|----|----|----|----|----|----|----|-------|
| Underestimated vs prepared | NaCl 3.0 mol I-1 pH = 5.5 | 4 | 13 | 17 | 3 | 2 | 26 | 6 | 36 | 41 30 |
| Overestimated vs pn-ICP-MS | NaNO ₃ 0.06-0.11 mol I-1 pH = 1.0 | 47 | 22 | 18 | 30 | 26 | 16 | 41 | 30 | 19 22 |

Figures



920



922 the cation-exchange process in a column during elution with acid; c) elution of sodium,

923 magnesium, potassium and calcium-bearing matrix with HCl 2.0 mol l-1 solution; d) subsequent

924 removal of the REEs during elution with HCl 6.0 mol l-1.



Fig. 2. ICP-MS profiles of the REEs during the ablation of standard solutions in glass capillaries.



Fig. 3. REEs analyses of standard solutions obtained using LA-ICP-MS (for other REEs see Electronic Supplementary Material). Analytical uncertainties (2SD) are expressed in the symbols' sizes. Solid lines and corresponding equations represent the linear fits of the analytical results obtained for the prepared by weight standard solutions, whereas analyses of the NIST SRM glasses (610 and 612) used as calibrators are plotted for information.



Fig. 4. Comparison of the mass fractions of the REEs in experimental solutions obtained using LA-ICP-MS and ICP-MS after sodium isolation (regression lines for elements are calculated taking into account the complete set of obtained analytical results, including the few outliers for Ce).