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A Comparison of Alkaline and Organosolv Lignin Extraction Methods from Coconut Husks as an Alternative Material for Green Applications

Nur Hanis Abd Latif,^a Nicolas Brosse,^{b,*} Isabelle Ziegler-Devin,^b Laurent Chrusiel,^b Rokiah Hashim,^c and M. Hazwan Hussin^{a,*}

Effects of alkaline (kraft and soda) and organosolv pulping were evaluated relative to the structural properties of lignin isolated from coconut husk (CH) biomass. The various types of functional groups within the isolated lignin samples were characterized and compared using a variety of complementary analyses including Fourier-transform infrared (FTIR) spectroscopy, nuclear magnetic resonance (NMR) spectroscopy, gel permeation chromatography (GPC), and high-performance liquid chromatography (HPLC). All isolated CH lignin samples contained a significant quantity of non-condensed guaiacyl-type (G) and syringyl-type (S) units but a lesser amount of p-hydroxyphenyl (H) units, as demonstrated and supported by previous research. The alkaline CH lignin produced lignin with higher molecular weight ($M_{w\ SL}: 959\text{ g mol}^{-1} > M_{w\ KL}: 769\text{ g mol}^{-1}$) than the organosolv lignin ($M_{w\ OL}: 606\text{ g mol}^{-1}$) sample, resulting in smaller fragments and a higher degree of solubility in water or other solvents. Because of significant differences in the physicochemical characteristics of the various lignin polymers, their properties and structure were improved with respect to alternative approaches in lignin-based applications.

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Keywords: Coconut husk; Alkaline lignin; Organosolv lignin

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INTRODUCTION

Many researchers have become interested in turning abundantly available biomass waste into bio-based products as a way to help mitigate the rise in global warming induced by non-renewable petroleum/fossil-based products. Biomass wastes are typically made from agricultural wastes, such as livestock feed, which is left to decompose naturally, and still, others are left unutilized (Johari *et al.* 2016; Zakaria *et al.* 2017; Ram and Mondal 2018). In Malaysia, coconut trees (*Cocos nucifera* L.) are widely cultivated and known for their important use based on their various parts (Kumar 2011; Johari *et al.* 2016; Saari *et al.* 2016). According to the Department of Statistics Malaysia in 2020, the highest fruit in terms of self-sufficiency ratio (SSR) and per capita consumption (PCC) from 2015 to 2019 was coconut, which is consumed at a rate of 22.7 kg per year, followed by other fruits in selected crops (Mahidin 2020). The outer layer of the coconut fruit, known as coconut husk

(CH) is a major agro-industrial waste produced in developing countries every year that is biodegradable, low in cost, and readily available in large quantities (Kaewdang and Benjakul 2015; Johari *et al.* 2016; Saari *et al.* 2016; Ram and Mondal 2018). Coconut husk is composed of 30 wt% fibre and 70 wt% pith, which is primarily made up of cellulose (34.9 to 42.3%), hemicellulose (1.1 to 21.9%), and a rich amount of lignin (29.8 to 53.5%) (van Dam *et al.* 2004; Israel *et al.* 2011; Jincy *et al.* 2015; Saari *et al.* 2016). Furthermore, coconut husk fibre has been investigated for its ethanolic extract effect relative to the gel properties of gelatin (Kaewdang and Benjakul 2015), fuel production (Ram and Mondal 2018), cement material for building applications (Lertwattanaruk and Suntijitto 2015), and performance as adsorbents for the removal of heavy metal ions (Hg) (Johari *et al.* 2016).

Lignin is a well-researched natural feedstock for industrial applications that is widely studied such as an emulsifiers of catalytic cracking heavy oil (Xu *et al.* 2011; Upton and Kasko 2016), low-cost lignin carbon fibers manufacture (Baker and Rials 2013), bioplastic (Spiridon *et al.* 2015), biocomposite (Privas and Navard 2013), polyurethane foam (Luo *et al.* 2018), and in the production of value-added chemicals. In plants, lignin acts as an adhesive by filling the gaps between cellulose and hemicellulose and binding the lignocellulose matrix together as well as providing strength and stiffness to the cell walls (Qingqing *et al.* 2019). It is a complex three-dimensional biopolymer composed of three phenylpropanoid monomers: p-coumaryl alcohol (p-hydroxyphenyl, H units), coniferyl alcohol (guaiacyl, G units), and sinapyl alcohol (syringyl, S units) (Fig. 1) (Jincy *et al.* 2015; Aziz *et al.* 2019; Hussin *et al.* 2019; Latif *et al.* 2019; Qingqing *et al.* 2019). Recently, in a cellulose-based polymer electrolyte, CH lignin was shown to play an important part as a natural plasticizer by improving the ionic conductivity of the solid polymer electrolyte along with the presence of cellulose derivatives (Chua *et al.* 2020), and it has also been used for dye removal in carbon cryogel beads (Ngadi *et al.* 2015). Because CH lignin has a similar phenolic structure to phenol, with potential to cross-link with resins, it has shown a good performance especially in mechanical properties of exterior-grade plywood sheets used in industrial manufacturing (Saari *et al.* 2016; Aziz *et al.* 2019; Hussin *et al.* 2019).

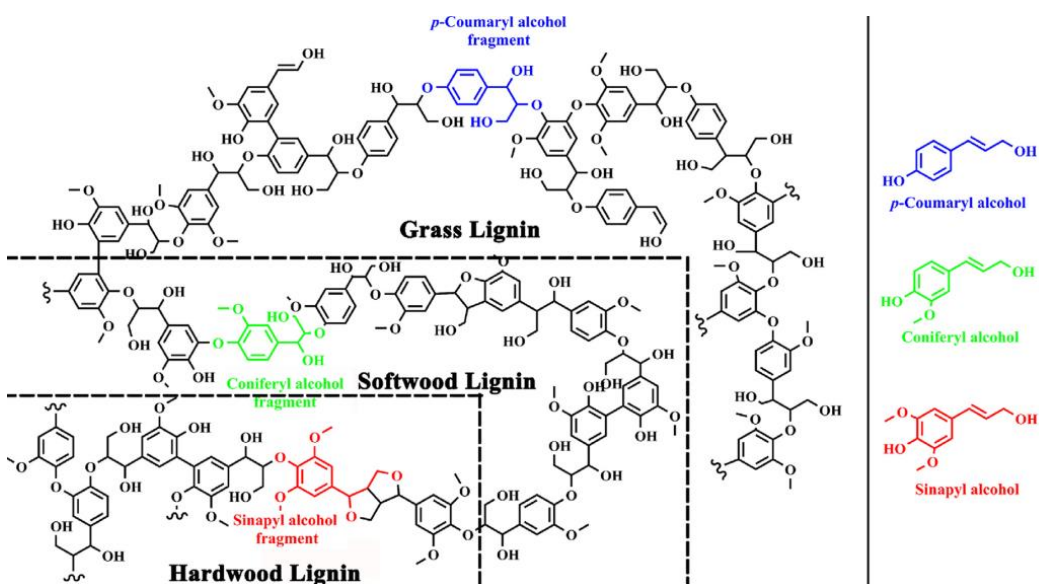


Fig. 1. Lignin structure with the main inter-subunit linkages (Qingqing *et al.* 2019)

A separation or pulping process is needed to utilize lignin and process it into high-value products. The physicochemical characteristics of lignin obtained from alkaline pulping (kraft and soda) and organosolv pulping processes were different (Aziz *et al.* 2019; Latif *et al.* 2019). According to the previous studies, the alkaline processes are used to isolate lignin from wood content in the pulp and paper industry (96% of the market). The breaking down of aryl-ether linkages occurred during alkaline pulping, resulting in the liberation of the lignin fragments as well as their dissolution and condensation reaction. During the condensation procedure, the molecular size of lignin fragments increased because of the conjugate addition of carbanions to quinone methide intermediates (Hussin *et al.* 2013). Due to the absence of sulfur contents, the soda lignin obtained was much purer than kraft lignin, which also improved its reactivity (Tribot *et al.* 2019). Previously, organosolv pulping produced more reactive lignin with higher purity, resulting in a lower molecular weight that is mostly soluble in common solvents (Hussin *et al.* 2013; Latif *et al.* 2019). Moreover, their structure contains a high proportion of oxidized groups and phenolic hydroxyl groups (such as Hibbert ketones), which facilitates chemical modification and incorporation into polymer formulations (Hussin *et al.* 2013).

Consequently, the current research aimed to study the structural and physicochemical characteristics of lignin isolated from coconut husk (CH) using various pulping techniques (kraft, soda, and organosolv pulping), as well as its potential as a phenolic substitution for material applications. There have not been several studies contrasting the functionalization of alkaline and organosolv lignin isolated from CH biomass. The isolated kraft lignin (KL), soda lignin (SL), and organosolv lignin (OL) were characterized using Fourier transform infrared (FTIR) spectroscopy, gel permeation chromatography (GPC) analysis, and ^{13}C - and 2D-heteronuclear single quantum coherence (HSQC) nuclear magnetic resonance (NMR) spectroscopy to investigate any changes in the structure of lignin after undergoing the pulping process. Additionally, thermal analyses were used to analyze the thermal activity of extracted lignins. Simultaneously, the phenolic acids, aldehydes, and sugar analysis of lignin were determined using high-performance liquid chromatography (HPLC) and high-performance anion-exchange chromatography with pulsed amperometric detection (HPAEC-PAD). To optimize the proper applications of lignin, the physicochemical characteristic of lignin isolated from CH were compared with several previous works.

EXPERIMENTAL

Materials

Coconut husks (CH) were collected in Sungai Petani, Kedah, Malaysia (5°39'08.5"N, 100°31'17.5"E) in August 2019. The husks were sun-dried for 2 days to remove any excess moisture before being ground into powder using a stainless steel laboratory blender (Waring Commercial, Torrington, CT, USA). Then, the ground CH was sieved into small particles (1 to 3 mm) and further dried in an oven at 50 °C for 24 h. The chemical composition (% w/w) of the CH biomass in term of moisture content ($9.26 \pm 0.15\%$), ash content ($2.50 \pm 0.002\%$), extractives content ($9.16 \pm 0.14\%$), cellulose content ($27.19 \pm 0.10\%$), hemicellulose content ($14.64 \pm 0.70\%$), and Klason lignin ($37.93 \pm 0.91\%$) was determined based on the National Renewable Energy Laboratory (NREL) laboratory analytical procedure (LAP) (Sluiter *et al.* 2005a,b, 2008a,b) and TAPPI T203 cm-09 (2009) standard. The obtained chemical composition in CH biomass was compared

with previous studies and is summarized in Table 1. For this study, the chemicals used such as nitrobenzene, *p*-hydroxybenzaldehyde, syringic acid, syringaldehyde, vanillic acid, *p*-hydroxybenzoic acid, vanillin, ferulic acid, and *p*-coumaric acid, were obtained from Sigma-Aldrich (St. Louis, MO, USA). Toluene (grade AR), ethanol (99.7%), sulfuric acid (98%), sodium hydroxide (97%), hydrochloric acid (37% v/v), glacial acetic acid (99.8%), dichloromethane, and acetone were purchased from QReC (Rawang, Malaysia). Acetic anhydride ($\geq 98.5\%$), acetonitrile (HPLC grade), and dimethyl sulfoxide-d₆ (d₆-DMSO) were obtained from Merck (Darmstadt, Germany). Chloroform (99.8%) and tetrahydrofuran (HPLC grade 99.99%) were from Fisher Chemicals (Loughborough, UK). Sodium sulfide (95%) was purchased from R&M Chemical (Essex, UK), as well as pyridine ($\geq 99.5\%$) was purchased from Bendosen (KL, Malaysia). Sodium chlorite (80% pure) was obtained from Acros Organics (Morris Plains, NJ, USA). In all experiments, distilled water was used, except that ultrapure water was used for running HPLC and HPAEC-PAD analysis.

Table 1. Comparison of Chemical Composition of Coconut Husk Biomass with Previous Studies (Israel *et al.* 2011; Lertwattanakruk and Suntijitto 2015; Ram and Mondal 2018)

Proximate Analysis	Chemical Composition (% w/w)			
	This Work*	Lertwattanakruk and Suntijitto (2015)	Ram and Mondal (2018)	Israel <i>et al.</i> (2011)
Moisture content	9.26 ± 0.15	NA	3.40	25.50
Ash content	2.50 ± 0.01	NA	5.20	9.00
Klason lignin	37.93 ± 0.91	29.80	47.50	53.50
Cellulose	27.19 ± 0.10	34.90	42.30	35.90
Hemicellulose	14.64 ± 0.70	21.90	1.10	NA
Extractives content	9.16 ± 0.14	NA	9.10	NA

* All samples were run in triplicates and the average values as well as the standard deviation values were reported (n = 3)

Extraction of Lignin Using Pulping Process

A 0.2-L TGYF-A stainless steel small high pressure chemical reactor (Henan Lanphan Industry Co. Ltd., Henan, China) was used when lignin was isolated under an alkaline (kraft and soda) and organosolv pulping process. In the alkaline pulping process, all of the conditions were the same as described in the authors' previous works (Hussin *et al.* 2013; Aziz *et al.* 2019) with minor changes. Kraft pulping process was initiated with the addition of active alkali (20%) and sulfidity (30%) at a ratio of 1:10 (solid to liquid, S:L). Meanwhile, in the soda pulping process, 30% of active alkali was used in the same ratio as described above. The maximum temperature, time, and pressure of the two pulping processes were set to 170 °C (12 to 15 bar) for 3 h. The final products (pulp and black liquor) were collected by separation using vacuum filtration. After that, the concentrated black liquor was acidified to pH 2 by dropwise addition of 20% (v/v) of sulfuric acid (H₂SO₄) to precipitate the kraft and soda lignin. Both precipitated lignins (KL and SL) were centrifuged at 3000 rpm for 15 min and oven-dried (50 °C) for 24 h. Subsequently, dry kraft and soda lignin were washed with excess water of pH 2 to remove any impurities or the residual of the hemicellulose compound before being dried in the oven (50 °C) for another 24 h.

In organosolv pulping, the addition of ethanol: water (65:35, v/v) and 0.5% (w/w) of sulphuric acid as a catalyst was mixed with the same S:L ratio as mentioned in alkaline pulping. As outlined in the method by El Hage *et al.* (2010), the cooking temperature used was at 190 °C for 1 h with continuous stirring. After the process, the fibrous residue was washed three times (3×50 mL) with 80% warm ethanol (v/v), and the concentrated black liquor was secured using vacuum filtration. Then, the concentrated black liquor underwent precipitation with three volumes of water and the lignin was collected using the same centrifuge conditions as described above for alkaline pulping. Finally, the wet solid organosolv lignin (OL) was oven-dried at 50 for 24 h before being stored in a vial for further analysis (Hussin *et al.* 2013; Latif *et al.* 2019). The percentage yields of all collected lignin (KL, SL, and OL) were calculated using Eq. 1 below (Aziz *et al.* 2019):

$$\text{Yield (\%)} = \frac{\text{Weight of recovered lignin (g)}}{\text{Weight of CH (g)}} \times 100 \quad (1)$$

Klason Lignin (HPAEC-PAD Analyses)

All the CH pulp of kraft, soda, and organosolv pulping of extractive-free samples were powder-ground, and the moisture content was measured using a Kern MRS 120-3-Mettler Toledo moisture analyzer (Greifensee, Switzerland). Based on the method illustrated by Tahri *et al.* (2016), approximately 0.175 g of dried extractive-free pulp samples were measured and put inside the 50-mL centrifuge tubes. Then, 72% of sulfuric acid (H₂SO₄) was added to approximately 1.5 mL in the tubes and the mixture was stirred continuously around 5 min using a glass rod. After stirring, the tubes were incubated in a 30 °C water bath for 1 h before the addition of distilled water (42 mL) and kept in an autoclave for 1 h. After the tubes were cooled down, the samples were filtered using glass microfibers in a Buchner funnel. The sample residue was washed with distilled water and left to dry in a 105 °C oven for Klason lignin content. As for the filtrate (soluble fraction), it was transferred into a round bottom flask and diluted to 100 mL with distilled water for HPAEC-PAD analysis (Dionex ICS-3000, Dionex Corp; Thermo Scientific, Sunnyvale, CA, USA).

From the Klason lignin hydrolysis, the monosaccharide contents of soluble fractions can be separated and analyzed using HPAEC-PAD (ICS-3000 Dionex) equipped with a Dionex CarboPac PA-20 (3 x 150 mm) analytical column, as illustrated in Tahri *et al.* (2016) and Obame *et al.* (2019). Approximately 20 µL of filtered samples were eluted at 35 °C with a flow rate (0.4 mL/min) under the following conditions: 99.2% pure water and 0.8% 250 mM NaOH (0 to 20 min); 75% pure water, 20% 250 mM NaOH, and 5% 1 M CH₃COONa/20 mM NaOH (20 to 37 min); and 40% pure water, 20% 250 mM NaOH, and 40% 1 M CH₃COONa/20 mM NaOH (37 to 41 min). After each elution, there was a wash and a period of equilibration. The external sugar and uronic acid standards were used for calibration with seven points per curve. All the sugar-containing monosaccharides (such as fucose, glucose, xylose, galactose, mannose, rhamnose, arabinose, galacturonic acid, and glucuronic acid) standards were supplied by Sigma-Aldrich (St. Louis, MO, USA).

Lignin Characterization

Fourier transform infrared analysis

A Thermo-Nicolet IR 2000 (Thermo Scientific; Waltham, MA, USA) FTIR spectrometer coupled with attenuated total reflectance (ATR) head of Perkin Elmer model

(Waltham, MA, USA) was used to analyze the isolated alkaline and organosolv lignin (KL, SL, and OL) samples under the indirect transmittance mode. The spectra of the vibration bands were recorded across the 600 to 4000 cm^{-1} region with a resolution of 4 cm^{-1} and 20 scans per sample.

Acetylation of lignin

According to Faris *et al.* (2015), all lignin samples need to undergo an acetylation process to enhance their solubility in organic solvents, especially for NMR spectroscopy and GPC techniques. During the acetylation process, all the hydroxyl (-OH) groups are replaced with acetyl (-COCH₃) groups. This process was done by dissolving approximately 1.2 g of lignin in an acetic anhydride/pyridine mixture (1:1), stirring continuously for 24 h at ambient temperature. After cooling the mixture to 0 °C with ice cubes, approximately 150 mL of 1% (w/v) hydrochloric acid (HCl) was added dropwise until precipitation developed. Finally, the solid precipitation was collected by vacuum filtration and dried in a 60 °C oven for 24 h after being washed with an excess of water until the pH of filtrate became neutral (pH 6 to 7).

For liquid-state ¹³C and 2D-HSQC NMR spectroscopy, a volume of 0.5 mL d₆-DMSO solvent was added to the acetylated lignin samples (KL, SL, and OL), and the samples were lightly heated with continuous stirring until completely dissolved. All NMR spectra were acquired using a Bruker Avance 500 MHz NMR spectrometer (Fallanden, Switzerland) with 22000 scans at 50 °C and interpreted with Bruker TopSpin software (v.3.5, Oxford, MS, USA). Next, the measurement of the average molecular weight (M_w) and average molecular number (M_n) were measured using an Agilent GPC model 1200 series analyzer connected to a UV detector (Agilent Technologies, Santa Clara, CA, US). An HPLC grade of mobile phase (tetrahydrofuran, THF) was used with a flow rate of 1.0 mL min⁻¹ in a Waters 1525 Binary HPLC pump. In brief, approximately 15 mg of acetylated lignin was mixed in 1.5 mL of THF solvent, and elimination of impurities was done using a 0.40 μm filter. Following that, the polydispersity index (PDI) was calculated using the ratio (M_w/M_n) after about 20 μL of solution (filtered) was injected into the GPC instrument.

Thermal analysis

The thermogravimetric analyses (TGA) of lignin samples (KL, SL, and OL) were studied using a Perkin Elmer TGA 7 analyzer (Waltham, MA, USA). The TGA analysis was performed at a continuous heating rate of 20 °C min⁻¹ in a nitrogen atmosphere (N₂) across a temperature range of 30 to 900 °C. Differential scanning calorimetry (DSC) analysis was carried out with a Perkin Elmer Pyris 1 (Waltham, MA, USA) to determine the glass transition temperature (T_g) of each lignin sample. Each lignin sample was heated at a constant heating rate of 20 °C min⁻¹ over a temperature range between 30 and 200 °C.

Phenolic acids and aldehydes composition

A simple nitrobenzene oxidation method was used to determine the S/G ratio of coconut husk lignin by analyzing using high-performance liquid chromatography (HPLC) with minor modifications, as implied by the previous works (Hussin *et al.* 2013; Latif *et al.* 2019). Ground-dried lignin (approximately 50 mg) was placed in a steel autoclave with a mixture of 2 M NaOH and 5 mL of nitrobenzene before undergoing oil bath heating at 165 °C for 3 h. Then, the solution was cooled down in ice water before being transferred to a glass-liquid separation funnel. The extraction was done three times with the addition

of chloroform in the separation funnel to get rid of the nitrobenzene reduction product. The oxidation mixture (upper layer) was acidified by adding concentrated HCl solution until reaching a pH of 2 and 3 before the second extraction was done using chloroform. After the second extraction, the excess chloroform solvent inside the oxidation mixture was removed by an evaporation process using a rotary evaporator heated between 48 and 50 °C. The final volume of an organic mixture (as a stock solution) obtained was dissolved with dichloromethane solvent up to 10 mL. A volume of 0.4 mL of stock solution was pipetted into a 50-mL volumetric flask with subsequent dilution up to the mark with acetonitrile/ultrapure water mixture (1:2, v/v). To avoid any impurities and insoluble particles before being injected into the HPLC system, the mixture of stock solution was filtered using a 0.40- μm Millipore membrane filter. Without any doubt, the classification of phenolic acids and aldehydes contents in CH lignin was detected using the Shidmazu HPLC system (Kyoto, Japan) coupled with Thermo Hypersil Gold C18 column (particle size 5 μm , length 250 mm, inner diameter 64.6 mm) by injecting approximately 20 μL of the solution. For the mobile phase, a mixture of acetonitrile/ultrapure water (1:8, v/v) together with 1% (v/v) of acetic acid was used in the HPLC system and operated at a flow rate of 1 mL min^{-1} . Each chromatogram of the samples was supervised at 280 nm using an ultraviolet (UV) detector (Shidmazu HPLC system; Kyoto, Japan).

RESULTS AND DISCUSSION

Proximate Analysis

Table 2 summarizes the percentage yields of lignin samples (KL, SL, and OL), as well as the sugar analysis of the resulting hydrolysates with HPAEC-PAD. Kraft and soda lignins produced higher percentage yields (%) from the alkaline pulping than organosolv lignin. This revealed that alkaline pulping had more severe reaction conditions, which stimulates the delignification process of lignin from CH biomass (Hussin *et al.* 2013; Domínguez-Robles *et al.* 2017). At the same time, organosolv pulping produced a low percentage yield of lignin due to the use of ethanol solvent and low severity reaction conditions, which dissolves the small molecular weight fraction of lignin (Hussin *et al.* 2013; Latif *et al.* 2019). According to the literature on the monosaccharide composition of coconut husk (CH), all the pulp in this study after alkaline and organosolv pulping contained a higher proportion of glucose and xylose.

Table 2. Sugar Analysis of Kraft, Soda, and Organosolv Lignin from Coconut Husk (% w/w on Dry Matter)

	KL	SL	OL
Yield (%)	21.74 \pm 2.04	23.67 \pm 1.82	2.79 \pm 0.30
Glucose (%)	21.83 \pm 1.17	22.19 \pm 5.63	33.26 \pm 3.28
Xylose (%)	64.65 \pm 3.36	65.25 \pm 8.26	58.67 \pm 4.50
Galactose (%)	2.96 \pm 1.68	2.81 \pm 0.41	3.22 \pm 0.15
Arabinose (%)	6.89 \pm 3.92	6.61 \pm 0.90	0.58 \pm 0.05
Mannose (%)	1.32 \pm 0.72	0.96 \pm 0.33	3.34 \pm 0.38
* All samples were run in triplicate and the average values as well as the standard deviation values were reported (n = 3)			
* KL = kraft lignin; SL = soda lignin; OL= organosolv lignin			

All the glucose comes from cellulose, a linear macromolecular polysaccharide that is made up of β -1,4-glycosidic bonds that connect a long chain of glucose units. Meanwhile, hemicellulose monosaccharide units primarily consist of xylose, arabinose, galactose, mannose, and some low-content saccharides (rhamnose and fructose) (Wang *et al.* 2017). The cellulosic composition in alkaline substrates pulp (KL and SL) was lower in glucose composition than that of organosolv substrate pulp (OL), resulting in higher removal of lignin because alkaline lignin produced a higher percentage yield of lignin (Sangian and Widjaja 2018).

Structural Characterization of Lignin

FTIR analysis

Fourier-transform infrared spectroscopy was used to observe any structural differences in the peak positions of vibrational bands between the three types of lignin compounds (KL, SL, and OL). Figure 2 depicts absorption band functionalities within the expanded region (600 to 1800 cm^{-1}) of FTIR spectra in all CH lignin samples, and the assignment bands are compiled in Table 3. The KL and SL spectra revealed moderately intense G bands (around 1271 and 1032 cm^{-1}) and weakly intense S bands (around 1331, 1112, and 826 cm^{-1}) while the OL spectra revealed quite strong G and S bands. The absorbance signal of S unit bands (1327, 1114, and 826 cm^{-1}) in OL spectra was noticeably higher than that of G bands in KL and SL, suggesting that the OL sample contained more syringyl content, as reported in the previous literature (Hussin *et al.* 2013).

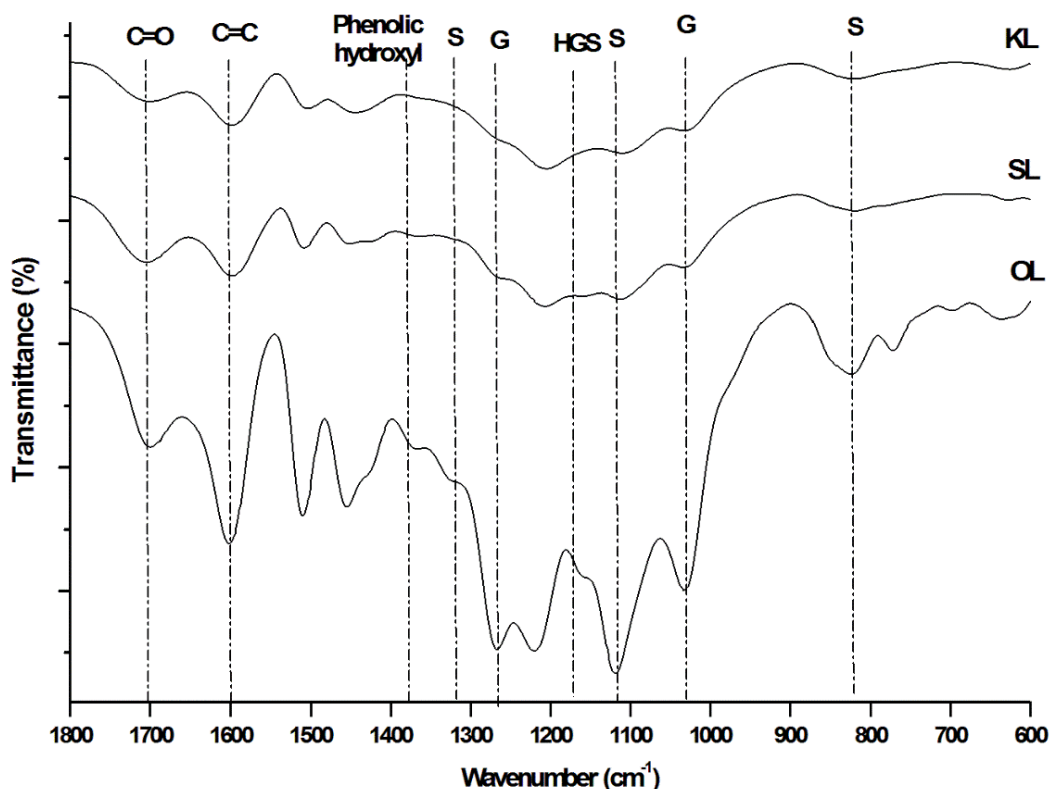


Fig. 2. FTIR spectra of KL, SL, and OL isolated from CH

Table 3. FTIR Assignment Bands of Coconut Husk Lignin

Assignment	KL	SL	OL
	Band Wavenumber (cm ⁻¹)		
C=O stretching of aldehyde/ketone groups	1701	1702	1700
C=C stretching vibration in benzene ring	1600	1603	1600
C=C stretching vibration in benzene ring	1502	1501	1503
C-H deformation (asymmetric in -CH ₃ and -CH ₂)	1444	1454	1452
C-C stretching (aromatic skeleton) with C-H in plane deformation	1422	1425	1424
O-H stretching vibration (phenolic OH)	1361	1364	1367
C-O stretching (syringyl)	1331	1330	1327
C-O (H) + C-O (Ar) (phenolic OH) and ether in syringyl and guaiacyl)	1271	1269	1269
Ar-CH in plane deformation (syringyl)	1111	1112	1114
C-O (H) + C-O (C) (first-order aliphatic OH and ether)	1032	1030	1034
C-H out of plane (aromatic)	915	915	916
C-H out of plane	826	824	826

The use of mixture solvent (ethanol-water) in organosolv pulping leads to the presence of free acetic acid from hemicelluloses, which contributes to lignin dissolution. As a result, it stimulates acid hydrolysis of the lignin structure, resulting in more intense peaks of the phenolic groups (1367 cm⁻¹) and carbonyl groups (1700 cm⁻¹) than alkaline pulping (Latif *et al.* 2019). In comparison, KL and SL samples had remarkably higher G bands intensities than OL, which was primarily because of the demethoxylation during the alkaline pulping process. It has been stated that demethoxylation reactions occur under severe reaction conditions during the alkaline pulping process, resulting in more stable G-type structures from S-type groups within the lignin molecules (Hussin *et al.* 2013; Aziz *et al.* 2019). According to Aziz *et al.* (2019), the presence of more G-type units in the lignin structure demonstrated that lignin had a better potential as an active site for polymerization. Furthermore, the typical absorption signal of HGS units for all lignin isolated from CH is comparable to wood lignin rather than occurring plant lignin due to a lack of hydroxyl phenyl propane (H) band in the spectra around 1160 cm⁻¹ (Hussin *et al.* 2018). Hereby, it was confirmed that all the CH lignin structures produced are primarily made up of S-type and G-type units.

NMR spectral analysis

The structural properties of all acetylated CH lignin were further characterized via NMR techniques. All the spectra were interpreted and compared with the other previous research as a reference on acetylated lignin (Sa'don *et al.* 2017a,b; Aziz *et al.* 2019; Latif *et al.* 2019). As shown in Fig. 3 and summarized in Table 4, the ¹³C NMR spectra of all acetylated lignin were qualitatively and quantitatively measured and placed into different categories of hydroxyl groups (-OH) and their derivatives. As the reference, the integral of the 162 to 102 ppm region was used, with six aromatic carbons and 0.12 vinylic carbons assumed. The integral value was divided by 6.12, which corresponded to one aromatic ring (Hussin *et al.* 2013). Within the integral region between 172 and 165 ppm chemical shift, the number of hydroxyl (-OH) groups in acetylated lignin samples were calculated. In all acetylated lignin spectra, the presence of guaiacyl, syringyl, and p-hydroxyphenyl units was associated with the aromatic region at 100 to 160 ppm (Fig. 3). According to the previous studies, the signals of S units were revealed around 104, 139, and 154 ppm (carbon

positions C2/C6, C4, and C3/C5), while G units signals were found at 113, 119, 135, and 146 ppm (carbon positions C2, C6, etherified C1, and non-etherified C3). The signal of 127 ppm was attributed to the H units in both alkaline and organosolv lignin samples. Furthermore, the carbon in carbonyl and carboxyl groups produced by the aliphatic esters and aliphatic carboxyls in lignin structures resulted in 172 ppm and 170 ppm showing a strong and moderate signal, respectively (El Hage *et al.* 2009; Hussin *et al.* 2013; Aziz *et al.* 2019). Moreover, the higher G unit signals in all acetylated lignin samples implied that CH lignin consists of a higher content of guaiacyl (G) units compared to the syringyl (S) units, as indicated by FTIR analysis and reported by Aziz *et al.* (2019).

Based on the integration of acetylated lignin samples, Fig. 3 and Table 4 show the amount of primary aliphatic (172.0 to 169.6 ppm), secondary aliphatic (169.6 to 168.6 ppm), phenol hydroxyl groups (168.6 to 165.0 ppm), and methoxy groups (58 to 54 ppm). The methoxy groups (-OCH₃) in G and S units in the lignin structures were assigned to the sharp peak observed at signal 57 ppm. It was detected that OL had a higher quantity of primary aliphatic -OH (0.49/Ar) than KL (0.13/Ar) and SL (0.16/Ar). For the amount of secondary aliphatic -OH, the KL (0.28/Ar) and SL (0.27/Ar) were nearly unchanged, but the OL (0.18/Ar) was remarkably lower. According to the previous study, the value of primary aliphatic-OH groups driven from integrals at 172 to 165 ppm and 65 to 58 ppm (0.97/Ar) had a good correlation for OL, indicating a higher amount of primary aliphatic-OH (0.49/Ar) functional groups in lignin structures (Hussin *et al.* 2013).

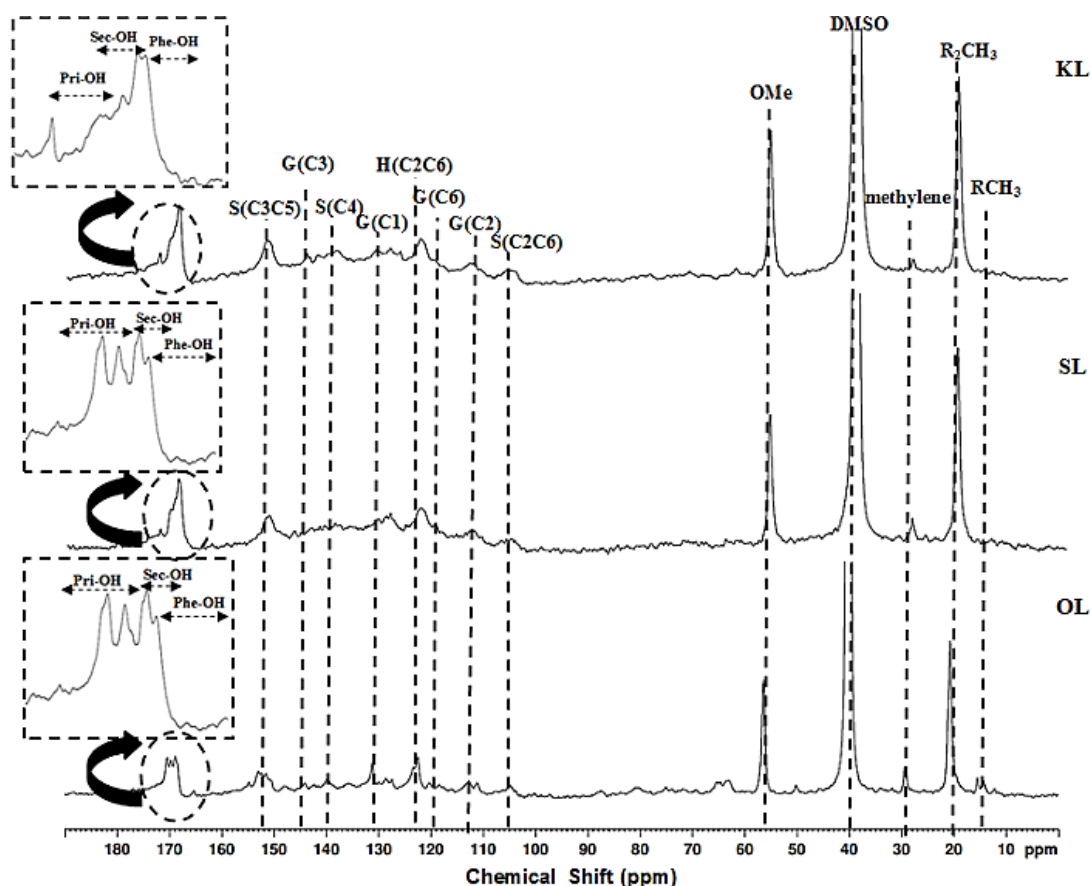


Fig. 3. ¹³C NMR spectrum of acetylated coconut husk KL, SL, and OL

Moreover, the amount of phenolic-OH in SL (0.25/Ar) and KL (0.19/Ar) were higher than in OL (0.15/Ar) because during alkaline pulping, the CH biomass undergoes a significant cleavage of aryl-ether bonds, resulting in a greater amount of phenolic hydroxyl groups. According to Aziz *et al.* (2019), in a study on the utilization of alkaline lignin from coconut husk biomass, the phenolic-OH content of SL (0.82/Ar) was also higher than that of KL (0.76/Ar), as shown in Table 4. As a result, it has been demonstrated that isolation of lignin from CH biomass using soda pulping produced lignin structures with a high amount of phenolic content. Furthermore, an increased amount of phenolic-OH content in the lignin structure allows for the activation of free ring position during the polymerization reaction, making it more reactive especially during the resinification process of phenolic resins (Hussin *et al.* 2018; Aziz *et al.* 2019).

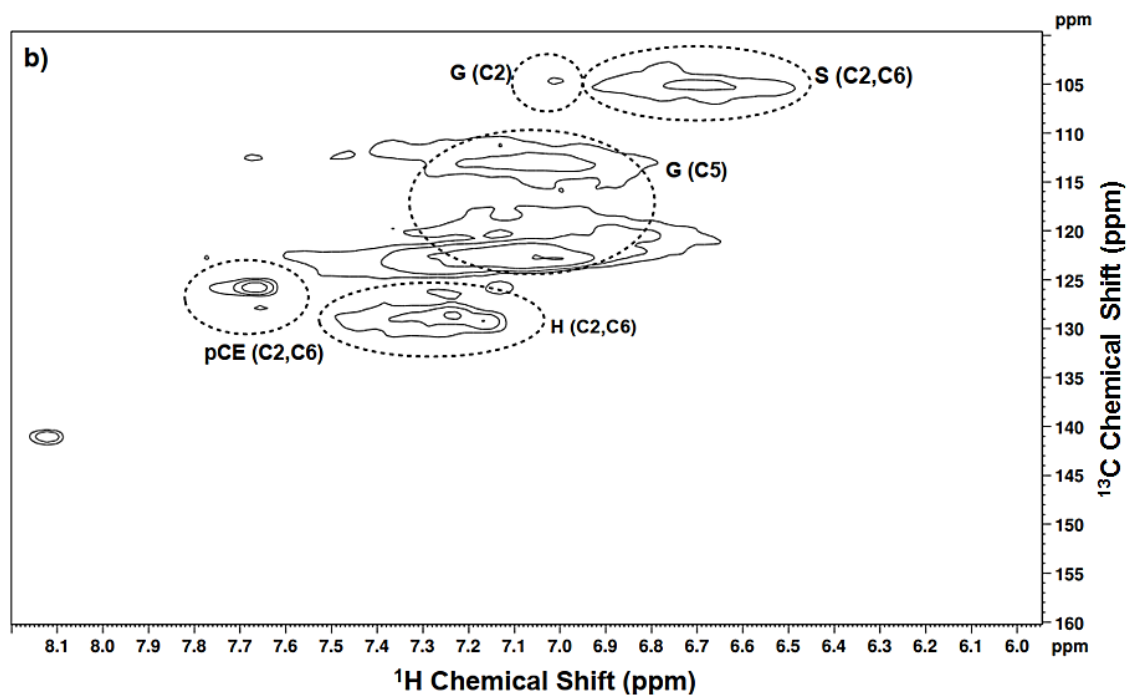
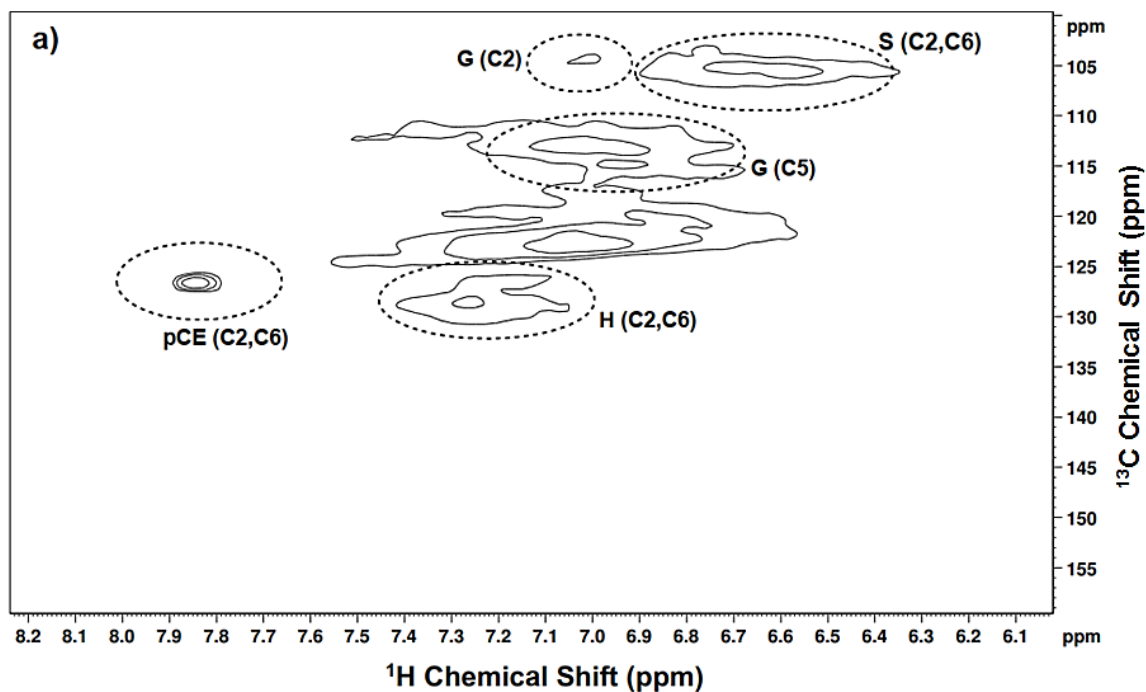
Table 4. Comparison of Signal Assignment for ^{13}C NMR Spectroscopy of Acetylated Coconut Husk of KL, SL, and OL with the Previous Study (Aziz *et al.* 2019)

Range (ppm)	Assignment	Amount (per Ar)				
		KL	SL	OL	KL*	SL*
172 to 169.6	Primary aliphatic – OH	0.13	0.16	0.49	0.16	0.22
169.6 to 168.6	Secondary aliphatic – OH	0.28	0.27	0.18	0.39	0.35
168.6 to 165	Phenolic-OH, conjugated – COOR	0.19	0.25	0.15	0.76	0.82
155 to 142	Aromatic C-O	1.78	1.94	1.96		
142 to 124	Aromatic C-C	2.21	2.68	2.02		
124 to 102	Aromatic C-H	2.12	2.66	2.13		
89 to 78	C β in β -O-4, C α in β 5, and $\beta\beta$			1.63		
73 to 71	C α in β -O-4			0.19		
64 to 61	C γ			0.44		
58 to 54	OMe, C1, and C β	0.39	0.51	0.38		
90 to 58	Alk-O			2.33		
65 to 58	OH _{prim}			0.97	0.16	1.30
31 to 29	α , β Methylene groups			0.48		
21	CH ₃ in acetyl group			0.43		
19	OCH ₂ CH ₃			0.32		
15	γ Methyl in n-propyl side chain			0.26		

*This is in reference to results obtained from Aziz *et al.* (2019)

The 2D-HSQC NMR spectrum was used to obtain details on the structural composition of all lignin samples. In the aromatic region of the HSQC spectrum, the main signals ($\delta_{\text{C}}/\delta_{\text{H}}$ 160 to 100/6.0 to 8.0 ppm) are depicted in Fig. 4 (a, b, and c). According to the previous studies (Hussin *et al.* 2014; Sa'don *et al.* 2017a; Latif *et al.* 2019), the correlation at $\delta_{\text{C}}/\delta_{\text{H}}$ (104.8/6.73 ppm) was assigned to C₂/C₆ position of syringyl (S) units, while the correlation for guaiacyl (G) units was assigned at $\delta_{\text{C}}/\delta_{\text{H}}$ 107.7/6.9 to 7.2 ppm (C₂/H₂) and $\delta_{\text{C}}/\delta_{\text{H}}$ 115.4/6.8 to 6.9 ppm (C₅/H₅). Furthermore, the signal for p-

hydroxyphenyl (H) units was assigned at a correlation of δ_C/δ_H (130.5/7.25 ppm). This information was in strong agreement with the FTIR and ^{13}C NMR results, which confirmed kraft, soda, and organosolv lignin as natural HGS units lignin. The p-coumarate (pCE 2/6) correlations were encountered at the δ_C/δ_H (126.8/7.4 to 7.6 ppm) region.



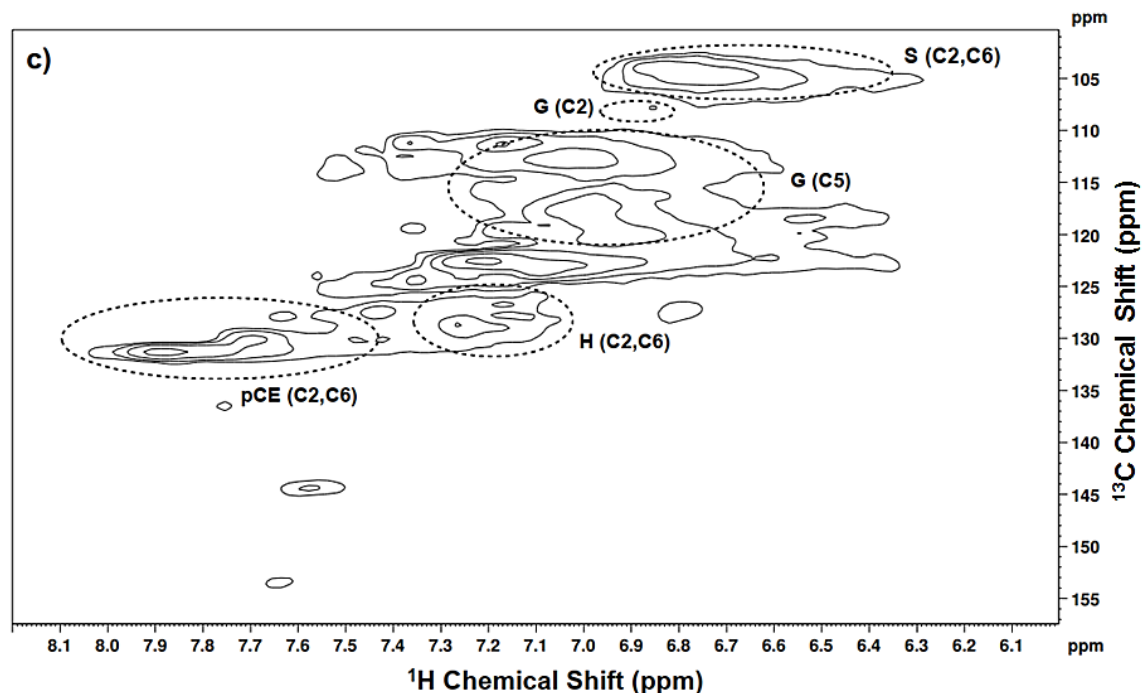


Fig. 4. 2D-HSQC NMR spectrum of acetylated coconut husk a) KL, b) SL, and c) OL

GPC analysis

Gel permeation chromatography was used to investigate the changes in molecules size and the distribution of alkaline and organosolv acetylated lignin samples. Table 5 shows the polydispersity value (PD) of the lignin fraction, which was calculated by dividing the value of average molecular weight (M_w) by the average molecular number (M_n). Based on Table 5, the SL sample had the highest M_w (M_w SL: $959 \text{ g mol}^{-1} > M_w$ KL: $769 \text{ g mol}^{-1} > M_w$ OL: 606 g mol^{-1}) and polydispersity values (PD) (PD SL: $2.44 > PD$ KL: $2.32 > PD$ OL: 1.37) when compared to KL and OL samples. This observation could be attributed to the different lignin compositions produced by the various pulping processes. According to the previous studies, the molecular weight of lignin may be related to some amounts of C-C bonds between phenylpropanoic units (involved in the C5 aromatic ring), even though β -O-4 linkages are the most common linkages in all types of lignin. It was reported that when guaiacyl-type (G) units were compared to syringyl-type (S) units, the G-type units were more adequate to form these C-C bonds because S-type units contained both substitution of C3 and C5 positions of methoxy groups. This explains why during the pulping process, this kind of C-C bond does not cleave due to its high stability. Thus, lignin with more guaiacyl-type (G) units was anticipated to result in an increased amount of molecular weight than lignin with syringyl-type (S) units (Hussin *et al.* 2013; Aziz *et al.* 2019). The difference in M_w values between KL and SL could be associated with the repolymerization reaction that occurs during the alkaline pulping method. During the kraft pulping method, the hydroxide and hydrosulfide anions react with lignin molecules, resulting in smaller water/alkali soluble-fragments of the polymer. Because of the production of a smaller molecular weight of KL, this fragmentation was extended through the cleavage of α -aryl ether and β -aryl ether bonds, which is the primary process in alkaline medium (Ibrahim *et al.* 2011). The results obtained for alkaline lignin (KL and SL) were further reviewed with the previous research (in terms of the same source of raw material

and pulping process). The M_w value of SL ($M_{w\text{ SL}}$: 2076 g mol⁻¹), and KL ($M_{w\text{ KL}}$: 1639 g mol⁻¹) (Aziz *et al.* 2019), and an ethanol soluble fraction of coconut shell ($M_{w\text{ EtOH-F}}$: 1598 g mol⁻¹) (Avelino *et al.* 2019) were higher than M_w values obtained in this study. As previously reported, the OL would result in low M_w value with smaller fragments due to the breaking of carbohydrate-lignin α -O-4 bonds and β -O-4 bonds to a limited extent during the organosolv pulping process. Because OL has a lower polydispersity (PD) value than KL and SL, it is predicted that OL will have a higher solubility in some organic solvents than other lignin derivatives (Hussin *et al.* 2013; Sa'don *et al.* 2017a, 2017b; Latif *et al.* 2019). Many studies focused on the preparation of phenolic resins with a higher molecular weight of lignin samples (KL and SL) due to the activation of more free ring positions in G-type units (Hussin *et al.* 2018; Aziz *et al.* 2019; Hussin *et al.* 2019).

Table 5. Comparison of GPC Analysis of All Acetylated Alkaline and Organosolv Lignin with the Previous Studies (Avelino *et al.* 2019; Aziz *et al.* 2019)

Lignin	M_w (g mol ⁻¹)	M_n (g mol ⁻¹)	PD (M_w/M_n)
KL	769	331	2.32
SL	959	393	2.44
OL	606	441	1.37
SL*	2076	513	4.05
KL *	1639	488	3.36
EtOH-F*	1598	807	1.98

*This is in reference to results obtained from Aziz *et al.* (2019) and Avelino *et al.* (2019)

Thermal behaviour analysis

Lignin is now extensively used in a variety of applications, especially in high temperature environments; thus, lignin's thermal decomposition has been studied (Watkins *et al.* 2015; Aziz *et al.* 2019; Latif *et al.* 2019). The thermal behaviour of alkaline lignin (KL and SL) and OL were studied using TGA and DSC. The thermogravimetric (TG) and derivative thermogravimetric (DTG) thermograms of all lignin samples are shown in Fig. 5a, 5b, and summarized in Table 6. The TG thermogram presents the weight loss of lignin as a function of thermal degradation temperature, while the DTG thermogram shows the corresponding rate of weight loss relative to temperature. All lignin thermograms were referred to and compared to the previous studies published in peer-reviewed journals (Ibrahim *et al.* 2011; Hussin *et al.* 2013; Watkins *et al.* 2015; Latif *et al.* 2019). The initial degradation step at around 30 to 180 °C was mainly caused by the evaporation of water absorbed, loss of carbon monoxide (CO), carbon dioxide (CO₂), and other volatile components caused by the lignin propanoid side-chain cleavage (Watkins *et al.* 2015; Latif *et al.* 2019). Simultaneously, the second degradation step (around 250 to 300 °C) in the lignin structures was explained based on the weight loss of hemicelluloses. Previous studies have shown that hemicellulose exists as a by-product during lignin precipitation, and degradation occurred at a temperature range from 200 to 300 °C (Ibrahim *et al.* 2011; Hussin *et al.* 2013; Aziz *et al.* 2019). In Fig. 5b, the maximum rates of weight loss (DTG_{max}) of all lignin samples were noticed at 383 °C (KL), 391 °C (SL), and 352 °C (OL). For comparison, in some previous studies on the isolation of lignin from coconut husk/fibre, a similar aspect was detected where KL and SL respectively recorded a DTG_{max} of 391 and 388 °C (Aziz *et al.* 2019) while organosolv lignins had a DTG_{max} of approximately 350 °C (Watkins *et al.* 2015).

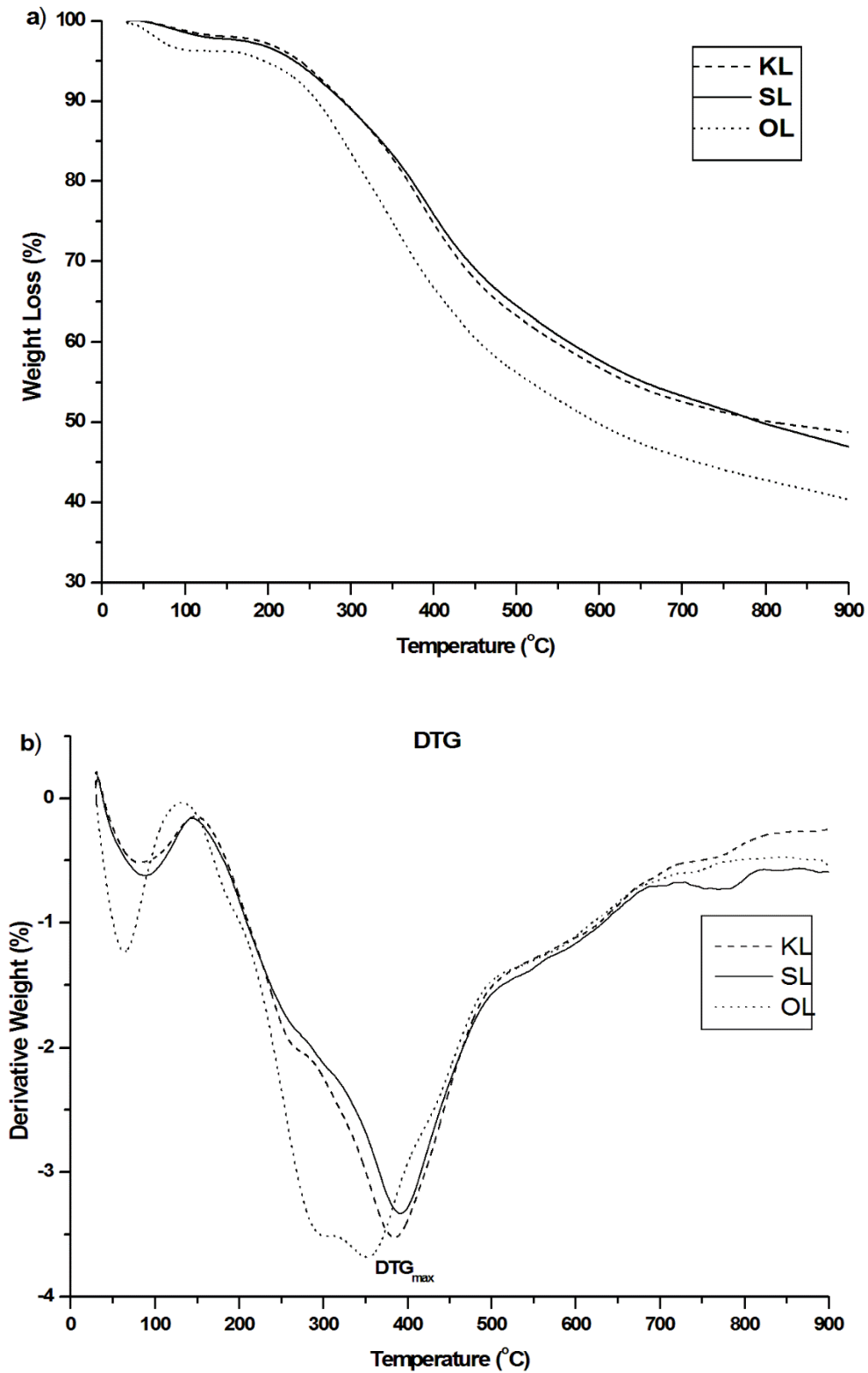


Fig. 5. (a) TGA and (b) DTG thermograms of KL, SL, and OL isolated from CH

Table 6. TGA-DTG Results of KL, SL and OL isolated from CH

CH lignin	KL	SL	OL
Initial weight loss temperature, $T_{-5\%}$ (°C)	263	262	229
Peak temperature of weight loss, DTG _{max} (°C)	383	391	352
Residual weight at 900 °C (%)	48.58	46.49	40.02

Therefore, it was concluded that the major degradation step for a complex lignin structure occurred around 300 to 400 °C. The thermal degradation of volatile products derived from the complex lignin structures, such as the inter-unit linkages fragmentation between phenolics hydroxyl, carbonyl groups (aldehyde acids), and benzylic hydroxyl will release the monomeric phenols in the form of vapor phase (Ibrahim *et al.* 2011; Watkins *et al.* 2015). A broad DTG_{max} curve was observed in the OL thermogram, which could be related to the fact that under low-temperature lignin could undergo degradation, implying that the structure of lignin consisted of smaller fragments, indicating an additional amount of β -O-4 linkages present with fewer C-C interlinked bonds (Hussin *et al.* 2013). At temperatures above 400 °C, all lignin samples showed a continuous weight loss until up to 900 °C, which could be relevant to the degradation or condensation reactions of aromatic moieties (Hussin *et al.* 2013; Latif *et al.* 2019). At the same time, around 900 °C in Fig. 5a, approximately 48.58% (KL), 46.49% (SL), and 40.02% (OL) of non-volatile residues remained intact and were not completely burned. Hence, it was suggested that a higher percentage of residues revealed that the char products in lignin samples are thermally stable at high temperatures, which can be attributed to the formation of a highly condensed aromatic structure with a high degree of branching (Hussin *et al.* 2013; Obame *et al.* 2019).

Differential scanning calorimetry analysis is an effective thermal method for determining the glass transition temperature (T_g) of polymers concerning their industrial applications and processing. The T_g value is an important temperature for determining the significant properties of amorphous polymers (lignin), which undergo a transition from a glassy-to-rubbery state at a specific temperature (Ibrahim *et al.* 2011; Hussin *et al.* 2018). It was discovered that SL had the highest T_g value (103.69 °C) when compared to KL (T_g : 91.31 °C) and OL (T_g : 70.89 °C), which could be attributed to the influence of the molecular weight and free volume present in the lignin samples (Hussin *et al.* 2013). In theory, free volume refers to the space of a solid or liquid sample that is not occupied by polymer molecules, whereas activation energies can be used to describe T_g (Ibrahim *et al.* 2011). The T_g value will appear only after the main chains have accumulated enough energy to rotate their segments. The lignin chains will vibrate as usual with the addition of energy, but as extra energy is added, rotation of the chain segment occurred and resulted in the presence of free space. With the increasing T_g value, lignin with higher molecular weight produced fewer chains ends and resulted in lower mobility, thus demanding more energy for rotation to occur (Ibrahim *et al.* 2011; Hussin *et al.* 2013; Latif *et al.* 2019). Therefore, the higher molecular weight of alkaline lignin (SL and KL) observed in GPC analysis indicated that alkaline samples had a few chain ends, which increased the energy required to rotate the chains due to the more free space created compared to OL. These findings also correlated with previous studies: kraft lignin, KL (T_g : 66.73 °C), soda lignin, SL (T_g : 75.78 °C) (Aziz *et al.* 2019), and organosolv lignin, EtOH-F (T_g : 133.00 °C) (Avelino *et al.* 2019), and it demonstrated that the alkaline lignin (SL and KL) in this study had higher T_g value but lower T_g value of OL in terms of using coconut husk wastes.

According to the thermal analysis data obtained in this study following the GPC analysis, lignin produced by the alkaline pulping process had a higher molecular weight with larger fragments than the organosolv pulping process, resulting in an increase of the T_g value.

Aldehydes and phenolic acids chemical composition

Alkaline nitrobenzene oxidation is a simple and widely used method to elucidate the constitutive structural compositions of the lignin samples. During the oxidation process in Fig. 6, the main components of lignin units (H, G, and S) were oxidized into aromatic aldehydes and ketones (syringic acid (SA), syringaldehyde (S), vanillin (V), vanillic acid (VA), *p*-hydroxybenzaldehyde (PHB), and *p*-hydroxybenzoic acid (PHBA) by forming a methane semiquinone intermediate (Huang *et al.* 2019; Jiang *et al.* 2019). In contrast, the presence of common HGS units in each sample showed that CH lignin is the HGS type. Based on our previous studies on oil palm fronds (OPF) lignin (Hussin *et al.* 2013; Latif *et al.* 2019), Table 7 summarizes the analysis of phenolic acids and aldehydes produced by the alkaline nitrobenzene oxidation of CH lignin by a different pulping process. The molar ratio values of G, S, and H were calculated at a molar ratio of G:S:H (49:38:10) in SL, 39:36:22 in KL, and 33:34:20 in OL, respectively. The molar ratio between G:S units of alkaline samples (KL and SL) revealed that guaiacyl (G) units had a higher value than S units but in the OL sample, the higher value of S units was obtained. Similar studies have found that non-condensed S units can easily be degraded using the organosolv pulping method when compared to lignin isolated from alkaline pulping (Hussin *et al.* 2013; Latif *et al.* 2019). The higher value of G units present in alkaline lignin samples (KL and SL) was supported in FTIR and GPC analysis, suggesting that CH lignin could be a valuable active site for cross-linking, especially during the resinification process in the wood adhesives industry (Hussin *et al.* 2018; Aziz *et al.* 2019). Therefore, it was discovered that lignin isolated from coconut husk *via* different pulping methods yielded a high amount of non-condensed G and S units, which will be useful in a variety of applications.

Table 7. Yield (% Dry Sample, w/w) of Phenolic Acids and Aldehydes from Alkaline Nitrobenzene Oxidation of CH Lignin

Component	Rt (min)	% Weight (Dry Basis)		
		KL	SL	OL
<i>p</i> -Hydroxybenzoic acid (PHBA)	~8.01	1.07	0.58	0.69
Vanillic acid (VA)	~9.52	0.59	1.34	0.19
Syringic acid (SA)	~10.47	0.70	1.23	0.31
<i>p</i> -Hydroxybenzaldehyde (PHB)	~10.96	0.11	0.25	0.16
Vanillin (V)	~13.96	1.49	2.79	1.19
Syringaldehyde (S)	~16.80	1.21	1.89	1.12
Ferulic acid (FA)	~18.00	0.18	0.23	0.56
Molar ratio (G:S:H)		39 : 36 : 22	49 : 38 : 10	33 : 34 : 20
S/G Ratio		0.92	0.78	1.03

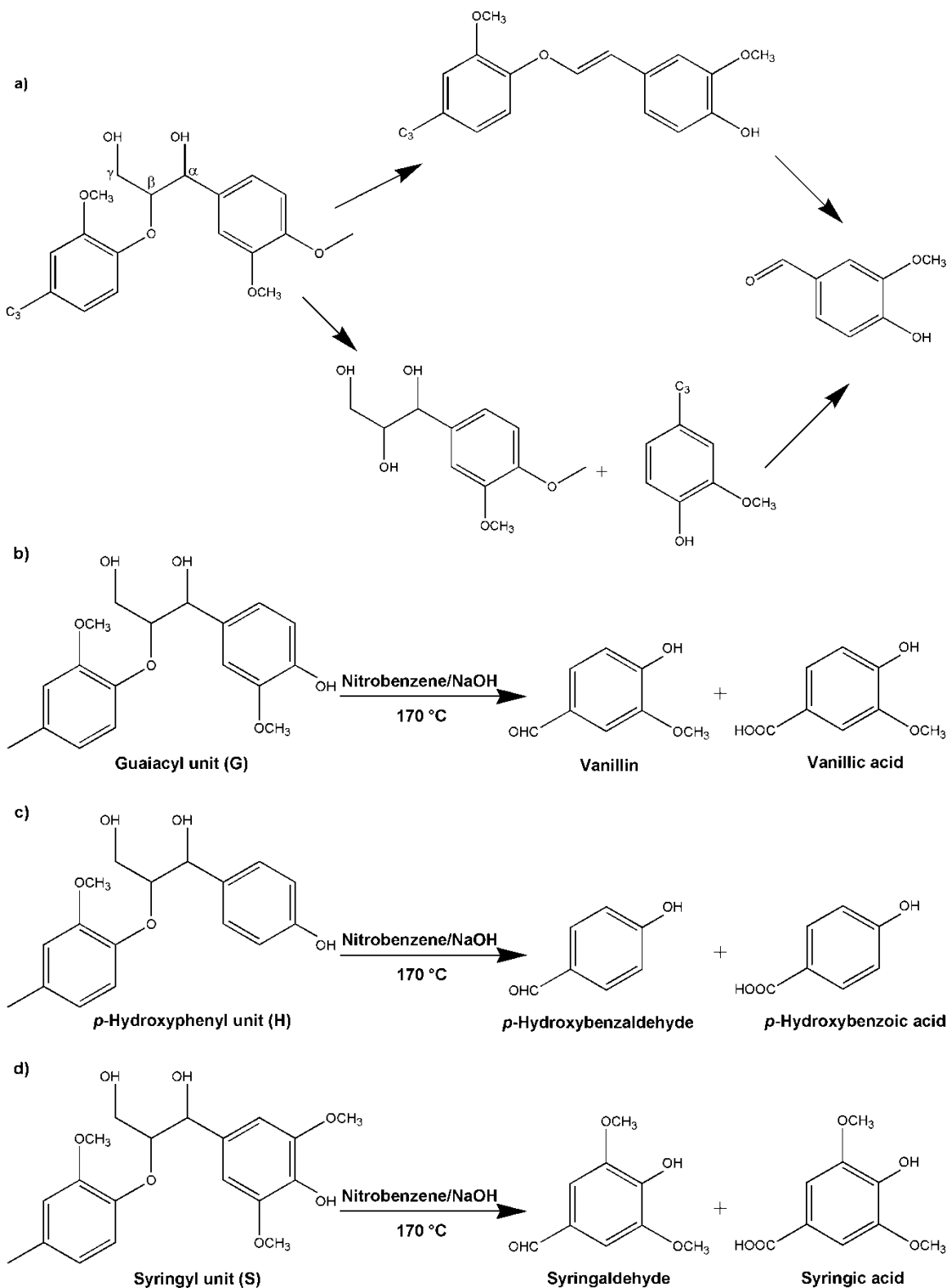


Fig. 6. a) The alkaline nitrobenzene oxidation reaction at β -O-4 lignin (Huang *et al.* 2019) and its oxidized products of b) guaiacyl (G), c) *p*-hydroxyphenyl (H), and d) syringyl (S) units (Jiang *et al.* 2019)

The Capability of CH Lignin in Lignin-based Applications

Based on the findings of this research, the structural characteristics of lignin isolated from CH waste demonstrated a strong link to various alternative approaches in its potential applications. The presence of more G-type units, as well as higher phenolic content in CH lignin structure, confirmed that CH lignin can activate the free ring position, resulting in a more reactive potential active site in the polymerization process of phenolic resins, as determined by FTIR, NMR, and GPC analyses (Ibrahim *et al.* 2007; Aziz *et al.* 2019). At the same time, the presence of higher S-type units (C3 and C5 positions linked to methoxy group) in CH lignin may provide better reducing power ability and result in increased antioxidant activity (Latif *et al.* 2019). From the authors' previous research, higher phenolic contents can increase the hydrogen-donating ability and phenoxyl radical formation, which could be beneficial in metal corrosion inhibition (Sa'don *et al.* 2017b; Latif *et al.* 2019). Because lignin has been used in a broad area of applications (including corrosion inhibition and adhesives), high T_g values of CH lignin can be advantageous in lignin-based applications. That is why the current focus of this work is on the isolation of lignin from CH as a phenol substitute in lignin-phenol-glyoxal adhesives.

CONCLUSIONS

1. The isolation of coconut husk (CH) lignin from different pulping processes (kraft, soda, and organosolv) produced different spectral characteristics and analytical results.
2. The molecular weight of alkaline lignin (kraft lignin (KL) and soda lignin (SL)) was found to be higher compared to organosolv lignin (OL), which is influenced by the repolymerization reaction.
3. During the alkaline oxidation process, the chemical composition analysis in CH lignin revealed that non-condensed S units were degraded more preferentially in the organosolv pulping process than in the alkaline pulping process, which contains more non-condensed G units.
4. Alkaline CH lignin exhibited a better potential in a diversity of lignin-based applications, especially as phenol substitution in lignin-phenol-glyoxal wood adhesives due to the increased amount of phenolic-OH content, molecular weight, and thermal decomposition.

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