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High added-value products from the hydrothermal carbonisation of olive stones

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Abstract

Olive stones (OS) were submitted to hydrothermal carbonisation (HTC) in order to evaluate the possibility of producing high added-value products, mainly furfural (FU) and 5-hydroxymethylfurfural (5-HMF) on one hand, and hydrochars and carbons on the other hand. Temperature (160 – 240°C), residence time (1 – 8h), initial pH (1 – 5.5) and liquid/solid ratio (4 – 48 w/w) were systematically varied in order to study the main products and to optimise FU production. FU production yield up to 19.9%, based on the hemicellulose content, was obtained. Other minor, but valuable, compounds such as 5-MF and some phenolic compounds were also produced. The hydrochar was carbonised at 900°C and the resultant carbon material was highly ultramicroporous with a peak of pore size distribution centred on 0.5 nm and a surface area as high as 1065 m² g⁻¹, typical of most carbon molecular sieves.

Keywords: Hydrothermal carbonisation; olive stones; furfural production; hydrochar.

1. Introduction

Olive trees are extensively grown in Mediterranean countries such as Spain, Greece, Italy and Tunisia, and olive stones (OS) are among the multiple by-products that are obtained from olive oil production. Olive production in the Mediterranean basin is estimated to be around 19 million tons according to the Food and Agriculture Organisation Corporate Statistical Database (FAOSTAT, 2016). Nowadays, OS industrial uses are nearly exclusively limited to energy cogeneration but other uses are also possible. In the present study, hydrothermal carbonisation (HTC) of OS, i.e., treatment in hot pressurised water was suggested for producing high added-value products (Knežević, 2009).

After submitting biomass to HTC, the resultant liquid fraction contains small molecules such as formic or acetic acids but also compounds with molecular weights higher than several thousand Da. Out of them, furfural (FU) and 5-hydroxymethylfurfural (5-HMF) are among the most interesting ones. FU, is industrially produced by acid hydrolysis of pentosans or directly from biomass (Dashtban et al., 2015), from 5-HMF (not obtained at high-scale), from glucoses and from fructoses. Possible formation mechanisms are shown in Fig. 1(a), where R is 'H' in the case of FU production from pentosan or 'CH₂OH' in the case of 5-HMF production from fructose (Zeistch, 2000). Glucose can be converted into fructose in aqueous media over homogeneous and heterogeneous catalysts, (Marianou et al., 2016) and through isomerisation in a wide temperature range (Kooyman et al., 1977), as shown in Fig. 1(b).

FU can be used as fungicide or nematicide, in special adhesives or flavours, for refinery lubricants recovery or as precursor of 5-methyl furfural, furfuryl alcohol, tetrahydrofurfuryl alcohol and tetrahydrofuran (Raman and Gnansounou, 2015). FU

production is around 430 000 ton per year (Raman and Gnansounou, 2015), and its price is close to \$1 000 per ton (Yemis and Mazza, 2012). The uses of 5-HMF are focused on substitutes of petroleum-based monomers for the production of fine chemicals and plastics, but industrial methods have not been developed yet (Chedda et al., 2007).

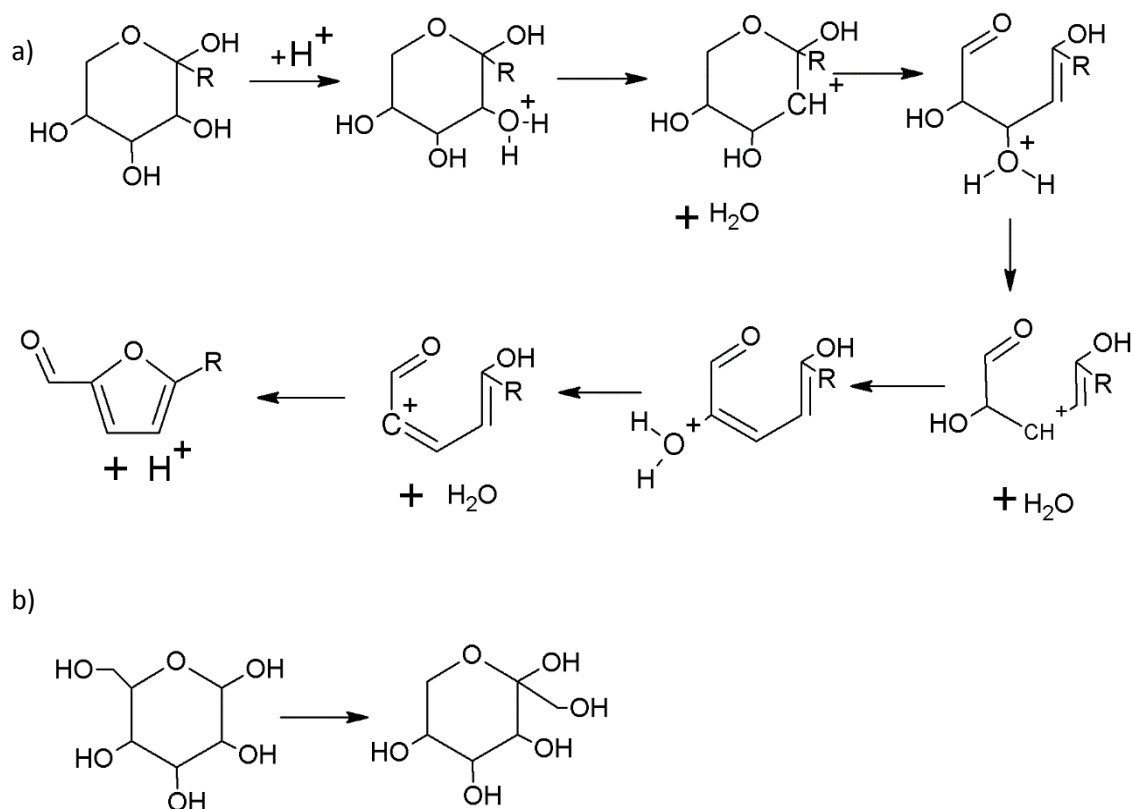


Figure 1: (a) FU and 5-HMF reaction mechanism from pentosan (R = H) and fructose (R = CH₂OH), respectively; (b) Isomerisation of glucose to fructose.

FU production from biomass has been extensively studied and different processes have been carried out to achieve good yields. These processes are generally one-step and batch processes, although two-steps processes (Raman and Gnansounou, 2015, Riansa-Ngawong and Prasertsan, 2011) and continuous processes (You et al., 2015) have been also proposed. FU yield was 3.3 wt. % on dry basis after HTC of rice hull using sulphuric acid as catalyst for 30 min at 125°C and 1.5 atm, with a liquid / solid

(L/S) ratio of 25 mL g⁻¹ (Mansilla et al., 1998). Using a microwave-assisted process at 180°C, L/S ratio of 100:1, pH 1.12 and 20 min of residence time, FU yields based on initial xylose were 48.4%, 45.7%, and 72.1% for wheat straw, triticale straw and flax shives, respectively (Yemis and Mazza, 2012). Faujasites and mordenites were used as catalysts for the production of FU (Moreau et al., 1998) and 5-HMF (Moreau et al., 1996). Xylose was submitted to HTC in water / toluene mixtures (vol./vol. 1:3) at 170°C, leading to a maximum yield of 34% and 42% after 30 and 50 min of reaction, respectively (Moreau et al., 1998). 5-HMF was obtained from fructose when submitted to HTC using water / methyl isobutyl ketone mixtures (vol./vol. 1:5) at 165°C. In this case, a maximum yield of 50% and 70% were achieved after 30 and 60 min, respectively (Moreau et al., 1996). Lactic acid was also used a catalyst for FU production from fructose (Agirrezabal-Telleria et al., 2011). Fructose was submitted to HTC in a 50 wt. % lactic acid water solution at 150°C for 2 hours, and a 5-HMF yield up to 64% was obtained (Lopes de Souza et al., 2012). FU production from OS in HTC conditions was also studied by Montané et al., 2001. These authors got up to 50 – 65% of FU yield with respect to the initial pentosan content in the presence of sulphuric acid (0.05 to 0.250 mol L⁻¹) at a temperature between 220 and 240°C and with 60 – 780 s of residence time.

The aforementioned studies only took liquid products into account. However, solid products from HTC processes are gaining increasing importance such as potential precursors of carbon-rich materials (Schneider et al., 2011 Braghiroli et al., 2012, Braghiroli et al., 2014, Braghiroli et al., 2015, Braghiroli et al., 2015 b), Braghiroli et al., 2015 c)). After hydrochar carbonisation, the carbon materials have applications in the fields of catalysis, electrochemistry, energy storage, selective gas sequestration, water purification or soil amendment (Steinbeiss et al., 2009, Titirici et al., 2012, Kang

et al., 2012, Elmouwahidi et al., 2012 and Li et al., 2014; Schaefer et al., 2016). In the present work, HTC of olive stones was carefully considered to obtain a better understanding of the effect of the process variables (i.e., temperature, time, initial pH, and L/S ratio) on FU and 5-HMF production. The yield to hydrochar was also measured; one hydrochar also was pyrolysed and the textural properties of the resultant carbon material was analysed.

2. Materials and methods

2.1. Raw materials

Olive stones (OS) were obtained from a local factory in Gabès, Tunisia. Lignin, cellulose and hemicellulose contents were determined according to the following Tappi standards: T 264, T 203 and T 222 respectively. OS were milled to a particle size smaller than 2 mm. Distilled water was used as reaction medium. Hydrochloric acid (0.1 mol L^{-1} , Sigma-Aldrich) was used to decrease the initial pH.

2.2. Hydrothermal carbonisation (HTC)

HTC experiments were performed in 125 mL autoclaves (Anton Parr). The liquid (water) to solid weight ratio (L/S) was 8, i.e. 16 g of water and 2 g of OS, unless otherwise specified. Water and OS were both placed into a glass vessel, which was then introduced in the autoclave. The latter was next installed in a ventilated oven preheated at the desired temperature: 160, 170, 180, 190, 200, 220 or 240°C, for different reaction times: 1, 2, 4, 6 or 8 h. Once HTC was finished, the autoclaves were removed from the oven and left for cooling at room temperature for several hours.

Liquid and solid fractions were separated by vacuum filtration. The liquid fraction was weighed immediately after filtration and placed in a fridge until analysis. The solid

fraction was first placed in a vacuum oven for 6 hours at 60°C for complete drying before being weighed accurately.

Taking as a reference the experimental conditions leading to the highest FU concentration, the pH was changed from 5.5 (natural pH) to 1 by adding 0.1 mol L⁻¹ HCl solution. As the liquid solid (L/S) ratio has a significant effect on the reaction products, it was set to 4/1, 6/1, 8/1, 12/1, 16/1, 24/1 and 48/1 (w/w). For all the experiments, 16g of water were used.

Temperature and time are two critical components of liquid hot water (LHW) treatments. The reaction ordinate, $\log R_0$, based on the combination of temperature, T (°C), and residence time, t (min), was defined by Overend and Chornet (1987) to describe the impact of LHW treatments on lignocellulosic components:

$$\log R_0 = \log \left(t \cdot \exp \left(\frac{T-100}{14.75} \right) \right) \quad (1)$$

14.75 is the activation energy based on the assumption that the reaction is hydrolytic and the overall conversion is first order. The logarithm of the reaction ordinate, ($\log R_0$), is defined at the severity of the treatment or severity factor.

2.3. Liquid fraction analysis

The main compounds in the liquid fraction were identified by gas chromatography coupled with mass spectrometry (GC/MS) analysis using a Clarus 500 GC/MS (PerkinElmer Inc.). Gas chromatography was carried out on a fused-silica capillary column (DB-5ms Ultra Inert; 30m x 0.25mm, 0.25µm film thickness; Agilent J&W). The gas chromatograph was equipped with an electronically controlled split/splitless injection port. The injection (injection volume of 0.5µL) was performed at 275°C in the splitless mode with a 1min splitless time. Helium (Alphagaz 2, Air Liquide) was used as carrier gas, with a constant flow of 1mL.min⁻¹. The oven temperature program was as

follows: 40°C constant for 2min, 40°C to 325°C at a rate of 7.5°C.min⁻¹ and then 325°C constant for 5min. Ionization was achieved under the electron impact mode (ionization energy of 70eV). The ion source and transfer line temperatures were 250°C and 330°C, respectively. Detection was carried out in scan mode: m/z 20 to m/z 500. The detector was switched off in the initial 5.5min (solvent delay). Compounds were identified by comparison with spectra from the NIST (US National Institute of Standards and Technology) mass spectral library.

Furans and phenolic compounds were quantified by a Dionex Ultimate 3000 high-performance liquid chromatograph, equipped with auto sampler, diode array and fluorescence detectors trough (Thermo Scientific). A five-point calibration curve was carried out for each compound using standard solutions and values were accepted only when results get inside the values of the calibration curve, concentrating or diluting for that FU, 5-HMF and 5-MF were separated using a Hypersyl Green PAH column (ThermoFisher Scientific). Four UV absorption wavelengths were used: 220, 276, 284 and 291 nm. Otherwise, fluorescence excitation and emission wavelengths were 360 nm and 443 nm, respectively. Mixtures of water and acetonitrile were used as mobile phases with water / acetonitrile volume ratios changing from 95:5 to 0:100 during the 35 min of the total duration of the experiments program. In the first 1.5 min, a ratio of 95:5 was fixed. Then, from 1.5 min to 15 min, the acetonitrile fraction increased linearly up to a ratio of 50:50 at 15 min. From 15 to 20 min, a linear increase was carried out until the ratio 0:100 was achieved, and this ratio was maintained for 5 min more. From 25 to 30 min, the acetonitrile amount decreased down to the initial ratio of 95:5, and was maintained 5 min until the end of the analysis.

Phenolic compounds were separated using a Pinnacle DB Biphenyl (Restek) column. 195, 201, 231 and 300 nm were used as UV absorption wavelengths. The

mobile phases used were water and acetonitrile too. The program started at 10% of acetonitrile during the first 3 minutes. Later, the acetonitrile ratio increased up to 15% at minute 5, and remained the same until minute 13. From 13 to 15 minutes, the acetonitrile increased up to 20% and remained constant for 13 minutes more. From 28 to 33 minutes, the ratio increased to 100%. Until 38 minutes, 100% of acetonitrile was kept. From 38 to 43 minutes, the acetonitrile decreased down to 10%, and remained constant until the end of the experiment at minute 50.

2.4. FU and 5-HMF yield assessment

Pentosans, such as xylose, and hexoses, such as glucose and fructose, are precursors of FU and 5-HMF, respectively, as already shown in Fig. 1(a). As cellulose and lignin do not produce FU, the FU yield is only related to hemicellulose content of OS and was therefore calculated from Eq. (2). 5-HMF, only related to cellulose content, was calculated from Eq. (3).

$$\text{FU Yield} = \frac{\text{FU massic concentration} \times \text{liquid volume}}{\text{mass of hemicellulose}} \quad (2)$$

$$\text{5 – HMF Yield} = \frac{\text{5-HMF massic concentration} \times \text{liquid volume}}{\text{mass of cellulose}} \quad (3)$$

2.5. Hydrochar yield

Hydrochar yield was calculated as the ratio of hydrochar mass after vacuum drying to OS mass on dry basis (index “db”), according to Eq. (4):

$$\text{Hydrochar Yield} = \left(\frac{\text{Hydrochar mass}}{\text{Initial Olive Stones mass}} \right)_{db} \quad (4)$$

2.6. Hydrochar carbonisation and carbon textural analysis

One hydrochar, prepared at 180°C and 6h, was carbonised under nitrogen flow (80 mL min⁻¹) in a tubular furnace. The heating rate was set at 1°C min⁻¹ up to 900°C, dwell time was 3 h, and cooling was carried out under nitrogen flow.

Pore texture parameters were derived from nitrogen and carbon dioxide adsorption isotherms at -196 and 0°C, respectively, using a Micromeritics ASAP 2020 apparatus. Samples were degassed for 48 h under secondary vacuum at 270°C. Surface area (S_{NLDFT}), micropore volume ($V_{\mu,\text{NLDFT}}$), as well as pore size distribution (PSD) were determined by application of the 2D-NLDFT heterogeneous surface model non-local ([Jagiello and Olivier, 2013](#)) to both CO₂ and N₂ adsorption data using the SAIEUS® routine provided by Micromeritics.

3. Results and discussion

Lignin, cellulose and hemicellulose contents in OS were found to be 29.88, 40.53 and 21.68 wt. %, respectively.

3.1. Liquid fraction analysis

The optimisation of FU production was focused over a range of severity factors ($\log R_0$) ranging from 4.32 to 6.2 and centred on an average value of 4.91, which corresponds to standard time and temperature conditions of 6 h and 180°C, respectively. All experimental conditions used in this work are presented in [Table 1](#).

Table 1: Experimental conditions used for optimising FU and 5-HMF production, as well as severity factors ($\log R_0$) and resultant FU and HMF concentrations and yields.

Sample	$\log R_0$	[FU] (g L ⁻¹)	[FU] yield (%)	[5-HMF] (g L ⁻¹)	[5-HMF] yield (%)
160°C, 6h	4.32	2.62	6.64	0.08	0.15
170°C, 2h	4.14	0.05	0.12	0.002	0.005
170°C, 4h	4.44	2.65	6.72	0.07	0.13
170°C, 6h	4.62	5.99	15.20	0.20	0.37
180°C, 2h	4.43	0.26	0.67	0.01	0.018
180°C, 4h	4.74	5.73	14.53	0.19	0.36
180°C, 6h	4.91	7.50	19.02	0.34	0.64
190°C, 4h	5.03	7.61	19.30	0.32	0.59
190°C, 6h	5.21	6.55	16.62	0.62	1.17
190°C, 8h	5.33	6.18	15.69	0.50	0.94
200°C, 2h	5.02	4.64	11.77	0.08	0.14
200°C, 4h	5.32	7.01	17.78	0.55	1.04
200°C, 6h	5.50	4.39	11.14	1.06	1.98
220°C, 2h	5.61	4.10	10.39	0.78	1.47
240°C, 1h	5.90	1.90	4.83	0.27	0.50
240°C, 2h	6.20	2.44	6.18	1.78	3.35

Figure 2 shows the GC/MS analysis of the liquid sample obtained after submitting OS to HTC at 190°C for 4h ($\log R_0 = 5.03$), where the maximum FU yield was obtained. Other compounds were, in order of appearance, 5-methylfurfural (5-MF), 2,6-dimethoxyphenol, vanillin, 1-(4-hydroxy-3-methoxyphenyl)-2-propanone (or 4-hydroxy-3-methoxyphenylacetone), 1-(4-hydroxy-3-methoxyphenyl)-ethanone, 4-

hydroxy-3,5-dimethoxy-benzaldehyde (or syringaldehyde) and 1-(2,4,6-trihydroxyphenyl)-2-pentanone. Especially interesting is 2,6-dimethoxyphenol, which is used as flavouring agent in food preparation and as substitute of phenol in phenol-formaldehyde resin, a commonly used adhesive for plywood (Bridgwater et al., 2008). Vanillin is used as flavour too, in perfumes, as a chemical intermediate in the production of pharmaceuticals and other fine chemicals, and for developing thin layer chromatography plates (Esposito et al., 1997). Syringaldehyde is used in pharmaceuticals, food, cosmetics, textiles, pulp and paper industries, and even in biological control applications (Nasir et al., 2012). Moreover, most of the aforementioned compounds are sold separately for fine chemical applications such as chromatography detection.

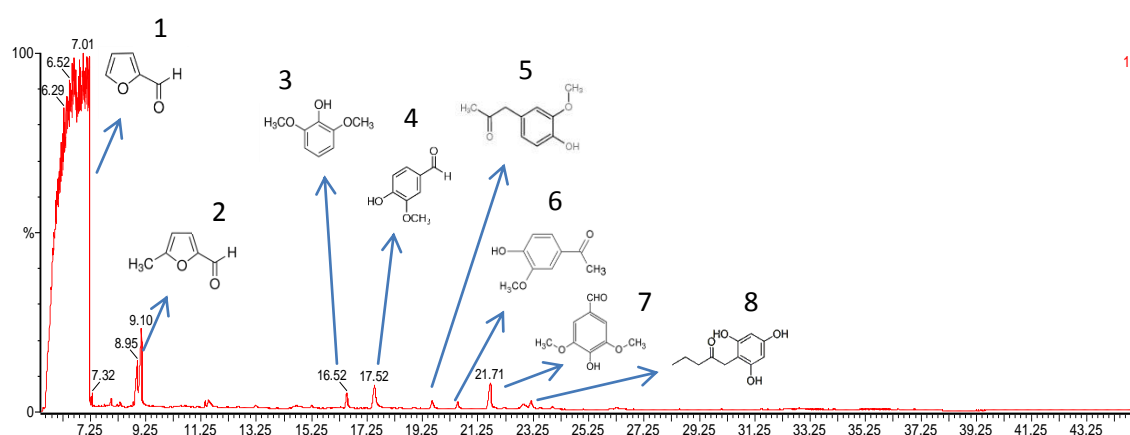


Figure 2: Main products identified in the liquid fraction produced at $\log R_0 = 5.03$ (190°C; 4h). [1] FU, 2) 5-HMF, 3) 2,6-dimethoxyphenol, 4) Vanillin, 5) 4-hydroxy-3-methoxyphenylacetone, 6) 1-(4-hydroxy-3-methoxyphenyl)ethanone, 7) syringaldehyde, 8) 1-(2,4,6-trihydroxyphenyl)-2-pentanone].

3.1.1 Effect of severity ($\log R_0$)

Figure 3(a) and 3(b) show the concentration of the two main products, FU and 5-HMF, as a function of $\log R_0$. FU concentration increased until arriving to a maximum at

7.608 g L⁻¹ at logR₀ = 5.03 (190°C and 4h), and decreased at higher logR₀. As for 5-HMF, no optimum was found but a strong increase of yield with logR₀. Thus, at the experimental conditions corresponding to the highest logR₀ = 6.20 (240°C, 2h), a maximum in the 5-HMF production was reached, 1.784 g L⁻¹. These results are in agreement with the literature. Hemicellulose degradation is indeed fast and starts at low temperatures: 60 min at 100°C and 15 min at 180°C produced a loss of 10 and 77.7% of the initial hemicellulose content, respectively (Sun et al., 2014). Moreover, hemicellulose does not require high severity conditions to produce FU, which degrades when increasing values of logR₀ (Zeistch, 2000). However, cellulose degradation is known to start above 200°C (Gao et al., 2012), thereby explaining the low 5-HMF production. All results including concentration and production yield of both compounds are shown in Table 1. The maxima in the FU and 5-HMF yields were 19.30 and 3.35%, respectively, based on hemicellulose and cellulose contents in OS, respectively. Based on OS, the maxima were 4.18 and 0.98%, respectively.

The maximum yield of 5-MF was found at logR₀ = 5.32, i.e., close to that of FU, see Fig. 3(d). This is due to the fact that FU and 5-MF are produced from hemicellulose and follow similar degradation paths. Vanillin and syringaldehyde showed an initial increase in their production with logR₀, but a plateau was reached at logR₀ higher than 4.9. In contrast, see Fig. 3(c), 2,6-dimethoxyphenol and 4-hydroxy-3-methoxyphenylacetone concentrations increased linearly with logR₀, and the maximum concentration they could reach here were 0.0656 and 0.052 g L⁻¹, respectively, at the highest logR₀ equal to 6.20. Phenolic compounds (vanillin, syringaldehyde, 2,6-dimethoxyphenol and 4-hydroxy-3-methoxyphenylacetone) are produced by lignin degradation, which occurs at more severe conditions than hemicellulose degradation.

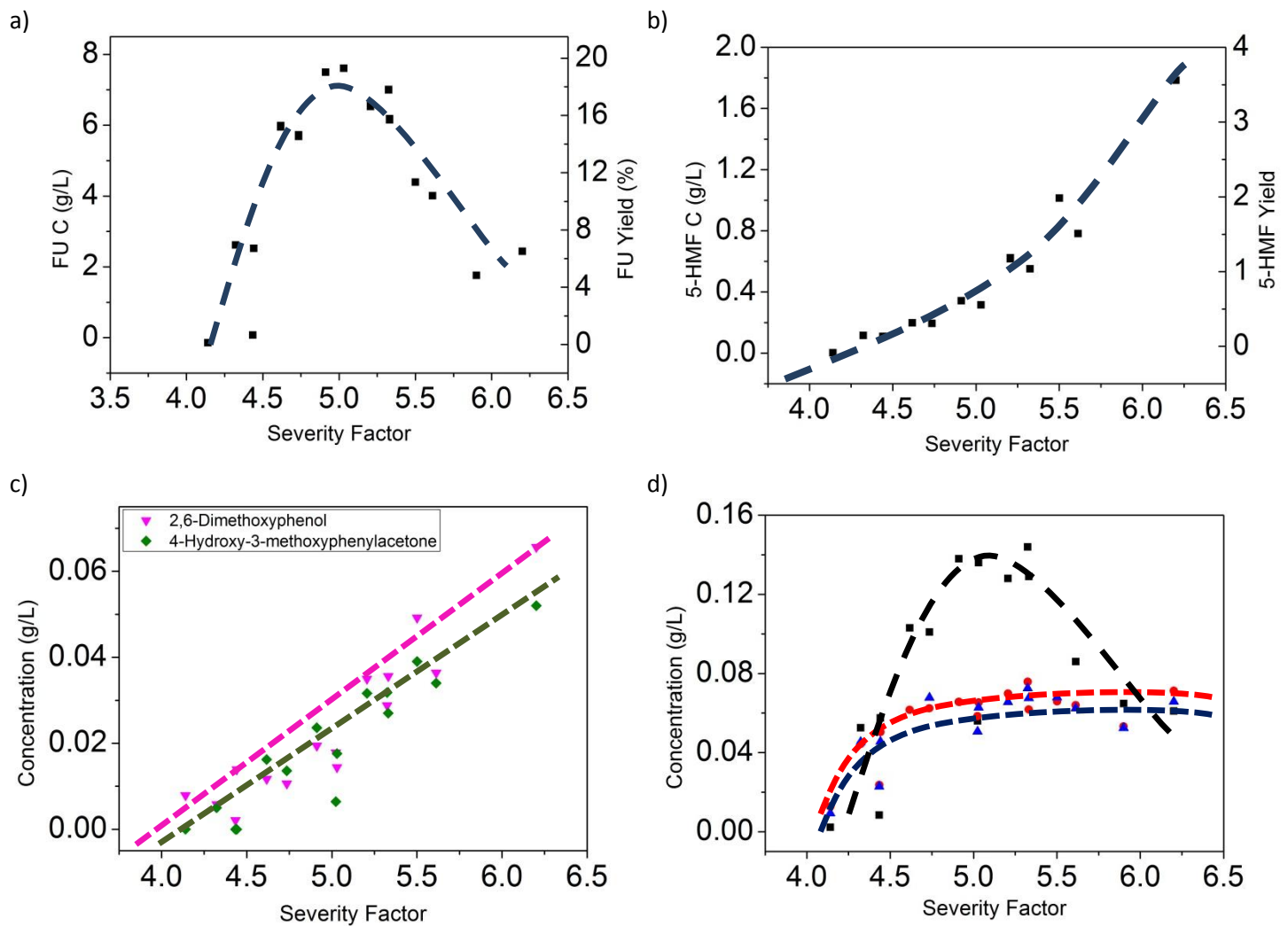


Figure 3: Production of major compounds: (a) FU and (b) 5-HMF, and of minor compounds: (c) 2,6 dimethoxyphenol and 4-hydroxy-3-methoxyphenylacetone, and (d), 5-MF (black), vanillin (red) and syringaldehyde (blue) as a function of $\log R_0$. For FU and 5-HMF, the yield is also indicated. Dashed lines are guides for the eyes.

3.1.2 Effect of pH

Acids have been widely used as catalysts in HTC processes, and it has been demonstrated that pH strongly affects FU and 5-HMF production (Yemis and Mazza, 2012). Taking as a reference the experimental conditions leading to the highest FU concentration, 190°C and 4h, the pH was studied from 5.5 to 1. Fig. 4(a) shows that higher fractions of FU and 5-HMF were obtained at lower pH, with optima at pH 2 and 1.5, respectively. Furfural yield reached 19.89% at pH 2. Despite 5-HMF concentration

increased by around 3 times from the initial natural pH, it was still lower than that obtained at 240°C and 2 hours. Once solid and liquid fractions were separated at the end of the experiment, the pH of the liquids was measured for observing whether pH changes might be related to FU and 5-HMF production. The corresponding data are shown in Fig. 4(b): the highest FU concentrations were clearly obtained at the lowest final pH, whereas a maximum of 5-HMF was obtained at pH = 2.8, as shown in Fig. 4(c).

As for minor compounds, i.e., 5-MF and phenolics, different behaviours were observed related to the initial pH and they are showed in Fig. 4(d). Just like for FU and 5-HMF, the production of 5-MF, vanillin and syringaldehyde were strongly affected by the initial pH. Thus, no 5-MF was detected when the initial pH was equal to 1. However, 2,6-dimethoxyphenol appeared to be not much affected by initial pH, whereas the production of 4-hydroxy-3-methoxyphenylacetone was the highest at low initial pH. Two opposite reactions may explain the different evolutions of products concentration. On one hand, hydronium ions facilitate dissolution and degradation of hemicellulose and lignin. On the other hand, high hydronium ion concentrations promote degradation. Therefore, 5-MF, vanillin and syringaldehyde concentrations sharply decreased when the initial pH decreased because degradation was more important. 4-hydroxy-3-methoxyphenylacetone concentrations rose due to higher lignin degradation while this product was affected by degradation in a lower extent when decreasing pH, and hence 4-hydroxy-3-methoxyphenylacetone increased at lower pHs.

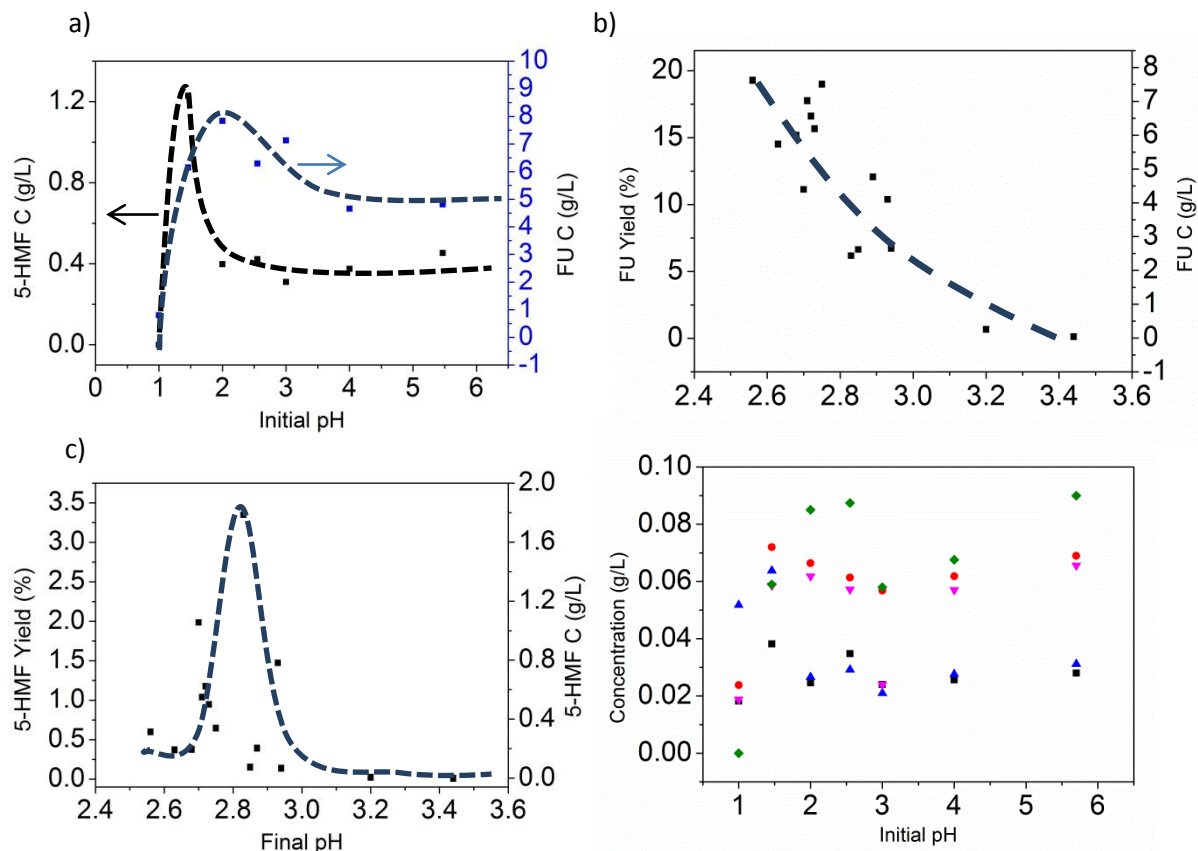


Figure 4: (a) FU and 5-HMF concentrations as a function of initial pH; (b) FU concentration and yield as a function of final pH; (c) 5-HMF concentration and yield as a function of final pH; (d) 5-MF and phenolic compounds concentrations (2,6 – dimethoxyphenol (black), vanillin (red) 4-hydroxy-3-methoxyphenylacetone (blue), syringaldehyde (pink), 5-MF (green)), vs. initial pH. Lines are guides for the eyes.

3.1.3 Effect of liquid/solid (L/S) ratio

As seen in Fig. 5(a)-5(c), FU and 5-HMF concentrations and yields depend on the L/S ratio used, which plays a fundamental role in the reaction mechanisms. Fig. 5(a) and 5(c) show FU and 5-HMF productions, respectively, as a function of the L/S ratio set to 4/1, 6/1, 8/1, 12/1, 16/1, 24/1 and 48/1 (w/w). FU and 5-HMF yields presented a steep maximum which was not expected from literature results. FU and 5-HMF highest yields were obtained at L/S ratios 24/1 and 12/1, respectively. This suggests that FU and 5-HMF production can be affected by interaction with other compounds and with themselves; therefore, too diluted solutions do not favour FU and 5-HMF production.

However, too low L/S ratio decrease selectivity to FU, since it is known that FU can react with itself. The same behaviour is expected for 5-HMF. Hence both FU and 5-HMF present production maxima as a function of L/S ratio.

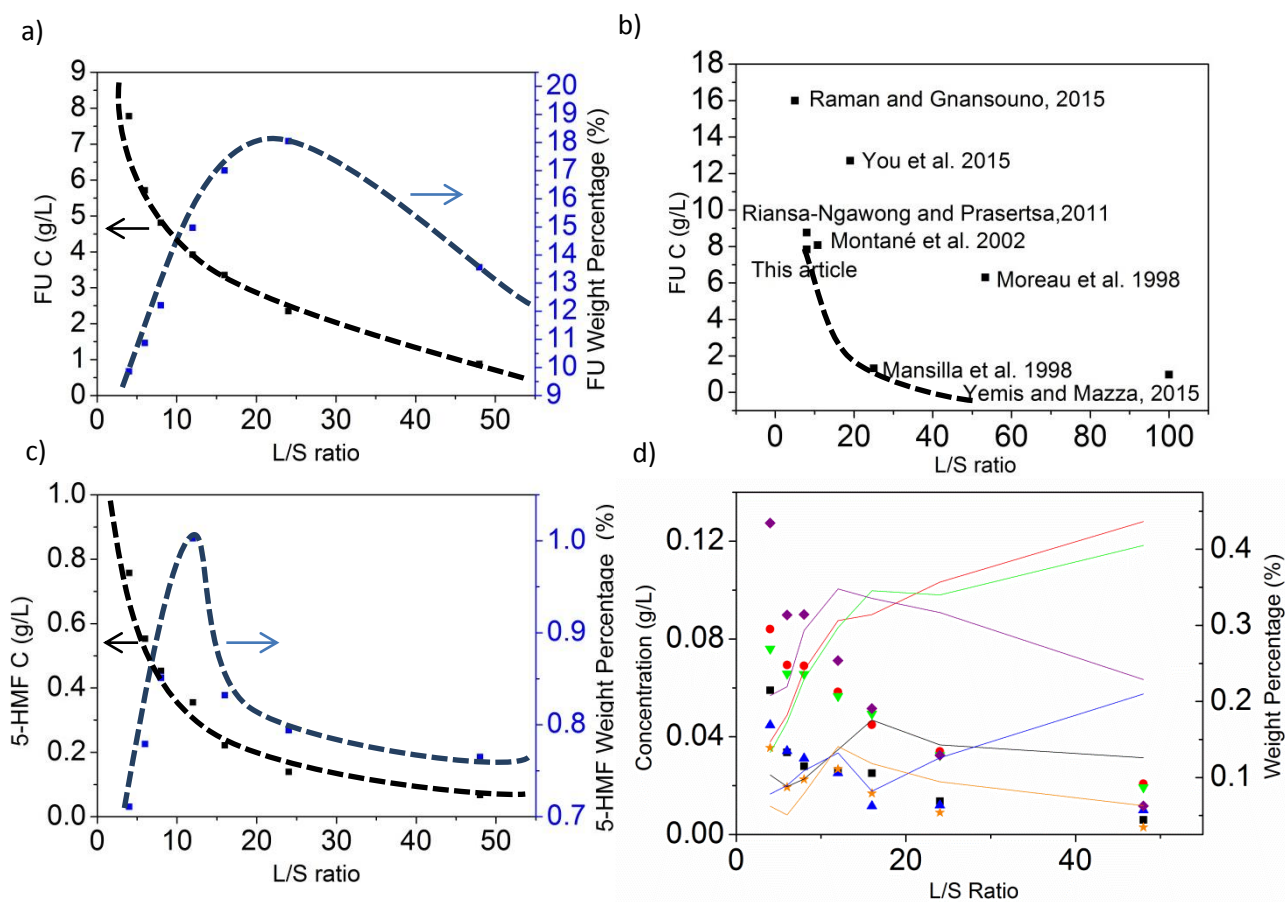


Figure 5: Evolution with the L/S ratio of: (a) FU concentrations and yield; (b) FU concentrations reported in literature compared to results obtained in this study (dashed line) (c) 5-HMF concentration and yield; (d) Concentrations (points) and yields (lines) of minor compounds (2,6 – dimethoxyphenol (black), vanillin (red), 4-hydroxy-3-methoxyphenylacetone (blue), syringaldehyde (green), 5-MF (purple) and 2-methoxyphenol (golden)).

Fig. 5(b) shows FU concentrations as a function of L/S ratio for data reported in the open literature, and compared with the maximum amount of FU obtained in the present work. The highest FU concentration obtained here, 7.608 g L^{-1} , was not far from that reported in other studies, although the L/S ratio was among the lowest ever used.

Therefore, higher FU yields might certainly be obtained by increasing the L/S ratio. In this figure, the data presented at lower L/S ratios than ours were obtained with two-stage processes, which suggests that a pre-treatment would be required to obtain an even higher FU production.

Fig. 5(d) shows the production yield of 5-MF and of minor phenolic compounds. 5-MF, 2-methoxyphenol (2MP) and 2,6-dimethoxyphenol presented a maximum, i.e., the same trend as for FU and 5-HMF. The other three phenolic compounds exhibited an increase of yield when submitting more diluted samples to HTC. For vanillin and syringaldehyde, this increase was even higher than that of 4-hydroxy-3-methoxyphenylacetone. The observed behaviour for 5-MF, 2MP and 2,6-dimethoxyphenol can be explained in the same way as for FU and 5-HMF. On the other hand, the behaviour of vanillin, syringaldehyde and 4-hydroxy-3-methoxyphenylacetone can be explained by taking into account that these compounds are not favoured by interactions between the different molecules and themselves, therefore an increase in concentration does not favour a rising yield; on the contrary, as water acts as catalyser, the higher is the water concentration, the higher is the production.

3.2 Solid fraction analysis

Fig. 6(a) shows the hydrochar yield as a function of $\log R_0$. By increasing $\log R_0$, the hydrochar yield decreased and the hydrochar was correspondingly enriched in carbon due to the higher conversion of cellulose, hemicellulose and lignin at higher treatment time and temperature. The L/S ratio also had an important effect on the hydrochar yield obtained. Fig. 6(b) shows that the hydrochar yield decreased exponentially with the L/S ratio. Increasing the water fraction indeed favoured dissolution and kinetics reaction,

due to the increase in the concentration gradient between water and OS, thus a lower yield was obtained for higher L/S ratios (You et al., 2015).

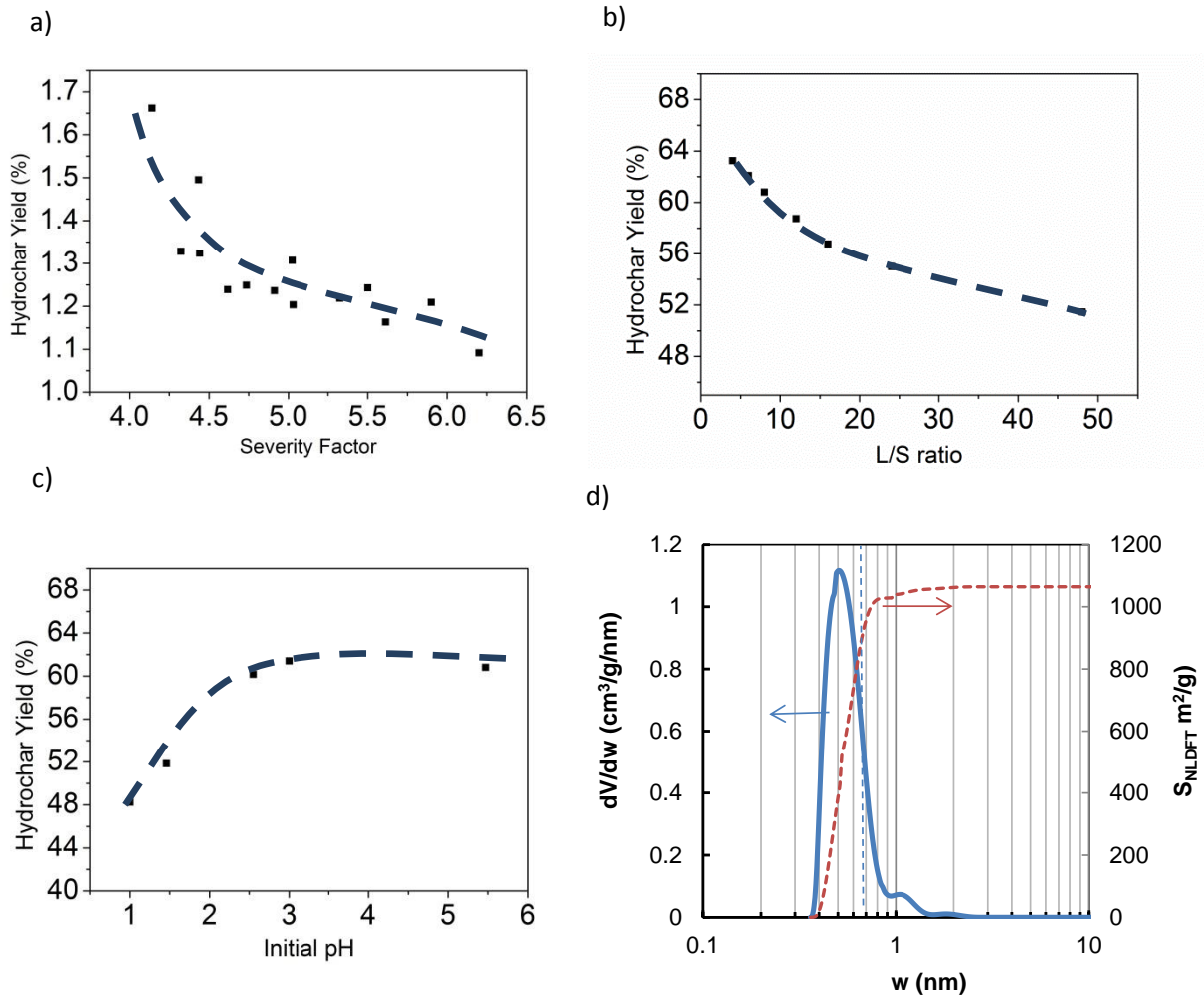


Figure 6: Hydrochar yield as a function of: a) $\log R_0$; b) L/S ratio (the solid line is an exponential fit); and c) initial pH .d) Pore size distribution (PSD) and cumulated surface area of the carbon produced from hydrochar at $\log R_0 = 4.91$. Lines are guides for the eyes in a), b) and c).

Fig. 6(c) shows that the hydrochar yield decreased when decreasing the pH below 3, due to hydrolysis and dehydration reactions (Lu et al., 2014). Finally, Fig. 6(d) shows the pore size distribution of the carbon produced after hydrothermal treatment at $\log R_0 = 4.91$ and further pyrolysis under nitrogen atmosphere at 900°C. Nitrogen adsorption

analysis was extremely slow, lasting up to seven days due to the diffusional resistance at -196°C, and the pore volume was not fully accessible to nitrogen. Textural surface properties were therefore assessed by nitrogen and carbon dioxide adsorption at -196°C and 0°C, respectively. The PSD of the resultant carbon materials was unimodal and narrow, centred on 0.5 nm, i.e., in the range of ultramicroporosity (pore diameter narrower than 0.7 nm). Total pore volume was equal to micropore volume, no mesoporosity (pore diameter between 2 and 50 nm) was detected. The surface area and the micropore volume (pore diameter narrower than 2 nm), calculated from the NLDFT model, were as high as 1065 m² g⁻¹ and 0.26 cm³ g⁻¹, respectively. Most part of the microporosity was in the range of ultramicropores and was equal to 0.22 cm³ g⁻¹. Therefore, the pyrolysis of OS-derived hydrochars produced ultramicroporous carbon materials with pore texture characteristics comparable to molecular sieves (CMSs) presently commercialised for gas separation. For instance, Carboxen 1003 ([Sigma-Aldrich, 2016](#)) has a surface area of 1000 m² g⁻¹, a micropore volume equal to 0.35 cm³ g⁻¹ and a pore diameter between 0.5 and 0.8 nm. The carbon materials produced here were not activated carbons (ACs) and they cannot be directly compared with them. ACs are produced by chemical or physical activation that develops the pore texture producing materials with higher surface area and wider PSD than those of CMSs. The carbon materials produced in this study, if they are not used as CMSs, could be further activated to obtain ACs similar to those previously reported in the abundant open literature (Ouederni et al., 2006, Spahis et al., 2008, Kula et al., 2008, Ghouma et al., 2015, Bohli et al., 2015).

3.3 Economic balance

There are about 850 million olive trees all over the world, whose space occupied is more than 10 million hectares (ASEMESA, 2016). Considering that around 15-25 wt. % of the olive corresponds to the stone and taking only the Spain and Tunisia olive production, the production of OS would range from 1.5 to 2.5 million tons. In Spain, the cost of OS are around 60-140 € ton⁻¹, depending on variety and packaging (Olihueso, 2016). The actual approximate prices for the main products that can be obtained from OS are: 50 \$ g⁻¹ for CMS (Sigma-Aldrich, 2016), 1200 \$ ton⁻¹ for large-scale activated carbon production (Alibaba, 2016), up to 3000 \$ ton⁻¹ for biochar fertiliser at small-scale production (Ecowarehouse, 2016) and 1000 \$ ton⁻¹ for FU (Win, 2005), which production is expected to double from 2013 to 2020 (Grand View Research, 2016).

Keeping in mind that 60 – 65% in weight could be obtained as hydrochar and 4% as furfural, scaling up the present HTC process would therefore lead to 650 kg of a remarkable hydrochar and 40 kg of FU per ton of OS. Taking as a basis only the cost of fertiliser, FU and OS, great benefits could be obtained by HTC of OS. An accurate calculation of the investment and benefits should take into account operational and investment costs, as well as manpower, which are extremely variable from one country to another.

Fig. 7 shows the scheme of a possible industrial process for producing both biochar and furfural. The reaction would take place in a batch reactor, the solid product would be washed, and the washing water with all the rest of the liquid would be treated for recovering FU and 5-HMF. The solid fraction might be used directly as fertiliser or for preparing activated carbons (Ubago-Pérez et al., 2006). FU and 5-HMF might be separated from HTC liquids in: (i) a steam column, regarding their differences of boiling points, 162°C and 116°C for FU and 5-HMF, respectively; or (ii) in a stripping

column using nitrogen, which is considered as a cleaner alternative (Agirrezabal-Telleria et al., 2011). A further analysis of operational costs, waste treatment, as well as recovery of other interesting compounds and even valorisation of gas produced in HTC should be carried out in order to improve the energy and economic balance of the suggested process.

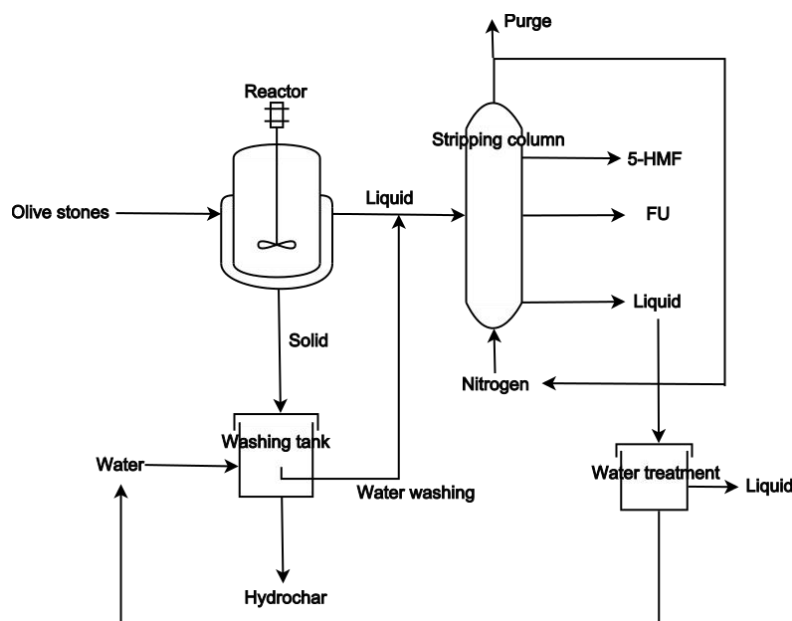


Figure 7: Possible process diagram for simultaneous hydrochar, FU and 5-HMF production from olive stones.

4 Conclusion

The treatment of olives stones (OS) in pressurised hot water, lately called hydrothermal carbonisation (HTC), allowed producing furfural (FU) and 5-hydroxymethyl furfural (5-HMF) as main products in the liquid phase, as well as a valuable solid product.

The highest FU yield was obtained at short times (4 and 6h) and moderate temperatures (180 and 190°C) while 5-HMF best yield was obtained in the hardest

severity conditions (240°C, 2h) considered in this work. Moreover, pH and L/S ratio were shown to be important in obtaining higher yields; pH 2 and 1.5 and L/S ratio of 24/1 and 12/1 were indeed found to be the best conditions for FU and 5-HMF production, respectively. The maximum FU production was close to 20%, based on hemicellulose content in OS.

The solid phase consisted in a hydrochar likely to be exploitable for soil amendment or as precursors of highly porous carbon materials. Indeed, such hydrochar was shown to be easily converted into carbon molecular sieve (CMS) by simple pyrolysis at 900°C. Producing such kind of materials might even improve the economical balance of the process, as far as the cost of commercial CMS having very similar pore texture characteristics, i.e., extremely narrow pore size distribution and high surface area, is considered.

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12

Figure captions

Figure 2: (a) FU and 5-HMF reaction mechanism from pentosan ($R = H$) and fructose ($R = CH_2OH$), respectively; (b) Isomerisation of glucose to fructose.

Figure 2: Main products identified in the liquid fraction produced at $\log R_0 = 5.03$ ($190^\circ C$; 4h). [1) FU, 2) 5-HMF, 3) 2,6-dimethoxyphenol, 4) Vanillin, 5) 4-hydroxy-3-methoxyphenylacetone, 6) 1-(4-hydroxy-3-methoxyphenyl)-ethanone, 7) syringaldehyde, 8) 1-(2,4,6-trihydroxyphenyl)-2-pentanone].

Figure 3: Production of major compounds: (a) FU and (b) 5-HMF, and of minor compounds: (c) 2,6 dimethoxyphenol and 4-hydroxy-3-methoxyphenylacetone, and (d), 5-MF (black), vanillin (red) and syringaldehyde (blue) as a function of $\log R_0$. For FU and 5-HMF, the yield is also indicated. Lines are guides for the eyes.

Figure 4: (a) FU and 5-HMF concentrations as a function of initial pH; (b) FU concentration and yield as a function of final pH; (c) 5-HMF concentration and yield as a function of final pH; (d) 5-MF and phenolic compounds concentrations (2,6 – dimethoxyphenol (black), vanillin (red) 4-hydroxy-3-methoxyphenylacetone (blue), syringaldehyde (pink), 5-MF (green)), vs. initial pH. Lines are guides for the eyes.

Figure 5: Evolution with the L/S ratio of: (a) FU concentrations and yield; (b) FU concentrations reported in literature compared to results obtained in this study (dashed line) (c) 5-HMF concentration and yield; (d) Concentrations (points) and yields (lines) of minor compounds (2,6 – dimethoxyphenol (black), vanillin (red), 4-hydroxy-3-methoxyphenylacetone (blue), syringaldehyde (green), 5-MF (purple) and 2-methoxyphenol (golden)).

Figure 6: Hydrochar yield as a function of: a) $\log R_0$; b) L/S ratio (the solid line is an exponential fit); and c) initial pH .d) Pore size distribution and cumulated surface area

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2 a), b) and c).

3 **Figure 7:** Possible process diagram for simultaneous hydrochar, FU and 5-HMF
4 production from olive stones.

5

6 **Table captions**

7 **Table 1:** Experimental conditions used for optimising FU and 5-HMF production, as
8 well as severity factors ($\log R_0$) and resultant FU and HMF concentrations and yield