

# Progress in aqueous dispersion RAFT PISA

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1	<b>Progress in Aqueous Dispersion RAFT PISA</b>
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#### 29 Abstract.

30 Polymerization induce self-assembly (PISA) is an emerging platform technology offering many advantages compared to the solvent switch methods. Aqueous dispersion reversible addition fragmentation chain transfer PISA 31 32 (aqueous dispersion RAFT PISA) has progressively won over many research groups due to its green chemical 33 process and amazing efficiency to produce wide range of morphologies. This review summarizes the recent works 34 reported in the literature on the aqueous dispersion RAFT PISA. After a general introduction on the PISA process, 35 exhaustive lists of the different RAFT activation techniques, the hydrophilic steric stabilizers, and the monomers 36 used are shown and discussed. Our objective is to provide an overview for experts and a toolbox for nonexperts that 37 aim to explore this robust and efficient route to produce block copolymer nano-objects for a specific application.

## 38 Keywords.

39 RAFT polymerization; PISA; Polymerization induced self-assembly, Aqueous; nano-object; dispersion

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## 41 **1. Introduction.**

Over the past decades, considerable efforts have been made to design functional polymeric nano-objects of 42 43 advanced physico-chemical properties, higher mechanical stability and modulable shape.[1-4] Such 44 nanostructures have found applications in various area especially in nanomedicine to improve the 45 bioavailability of drugs or to mimic natural cells, [5-7] and in cosmetic/food industries to stabilize 46 emulsions for instance.[8, 9] Polymeric nano-objects resulting from self-assembly of amphiphilic copolymers (ACPs) are probably the most studied systems in the literature.[10-20] Thanks to the 47 48 development of reversible deactivation radical polymerization (RDRP) techniques, [21, 22] including nitroxide-mediated polymerization (NMP),[23, 24] atom transfer radical polymerization (ATRP),[25, 26] 49 50 and reversible addition fragmentation chain transfer (RAFT),[27] that facilitates and expands the synthesis 51 of well-defined ACPs of various architectures (block, star and graft) compared to ionic polymerizations. Moreover, RDRP allowed introduction of new functional monomers hitherto difficult or impossible to 52 53 achieve using ionic polymerization. [28] ACPs-based nano-objects have been traditionally prepared via 54 direct dissolution in water [29, 30] or solvent switch methods [3, 4, 31, 32] such as nanoprecipitation, [33, 55 34] film-rehydration[35-37] and emulsion-solvent evaporation.[38] In these self-assembly methods, presynthesized ACPs are dissolved in a common solvent for both blocks, followed by a gradual addition of a 56 57 selective solvent of one specific block to form nano-objects. However, the low solid content (typically <158 % wt) of suspensions produced and the multiple processing/purification steps required by these methods 59 limit their potential upscaling.[39]

60 Polymerization induce self-assembly (PISA) is an attractive platform technology offering many advantages

61 compared to the solvent switch methods.[40-43] PISA is an one-pot methodology leading to formulation of

62 high solid content (up to 50 wt%) without performing additional purification steps. As shown in Scheme 1,

63 a solvophilic macromolecular stabilizer is chain-extended by polymerizing a desired monomer that forms a

#### 64 solvophobic block when reaching a critical chain-length. This results in a spontaneous assembly of the



65 ACPs into nano-objects of various morphologies, depending on the relative volume ratio of each block.

66 Scheme 1. Schematic Polymerization-Induced Self-Assembly conducted either in emulsion or in dispersion

67 *media. Cases of diblock and grafted amphiphilic copolymers.* 

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In academia, PISA was first carried out in heterogeneous media, previously called "ab Initio Emulsion 69 70 Polymerization", today known as "emulsion PISA". [44] In this case, the monomer is solvophobic such as benzyl methacrylate, [45-47] butyl acrylate, [48-51] glycidyl methacrylate, [52-56] methyl methacrylate, [57-71 72 59] styrene[60-65] and vinyl acetate.[66-68] However, this approach leads mainly to kinetically trapped nanospheres.[69] On the other hand, PISA in dispersion media,[70] either in aqueous[71, 72] or non-73 aqueous solvents, [17, 72, 73] offers an interesting alternative to produce nano-objects with advanced 74 morphologies.[69, 72, 74, 75] In practice, this is achieved by polymerizing a solvophilic monomer which 75 76 forms solvophobic polymer from a critical degree of polymerization. Several solvents have been used to 77 conduct non-aqueous PISA including: i) polar solvents (methanol, [40, 76] ethanol, [77] isopropanol, [78] 78 mixture of water and alcohol, [79-81] and acetic acid[82]) ii) non-polar solvents (dodecane,[83] 79 heptane,[84] chloroform,[85] and toluene[86]), and iii) some specific organic media (supercritical 80 CO<sub>2</sub>,[87] ionic liquids[88] and PEG of low molar masses[89]). Several monomers meeting the PISA prerequisites in non-aqueous dispersion have been reported in the literature such as benzyl 81 82 methacrylate, [77, 81, 83, 84] methyl methacrylate, [89] and styrene. [40, 78, 80, 88]

83 Aqueous dispersion PISA[71, 90] has progressively won over many research groups regarding its green chemical process (use of water) and its efficiency to produce wide range of morphologies, as judged by the 84 85 growth number of publications over the last fifteen years (Fig. 1). It has been successfully carried out using 86 almost all living/controlled polymerization techniques.[91-96] However, RAFT polymerization remains 87 the most used mechanism in PISA, [97, 98] thanks to its simplicity, its tolerance to water and its ability to 88 control the polymerization of a wide variety of monomers. [22, 99, 100] We termed here this process "aqueous dispersion RAFT PISA". The aim of this review is to provide an overview on the state of the 89 90 current literature for experts and a tutorial for nonexperts on the aqueous dispersion RAFT PISA. Readers

can refer to some other recent reviews dealing with specific topics related to PISA process, including
mechanisms for initiating and controlling the PISA process, [90, 101-103] approaches used for
crosslinking[104, 105] and formulation of functional nano-objects[32, 106-108] for biomedical
applications.[109-113]



95

96 Fig. 1 Published articles per year dealing with "polymerization induced self-assembly" (source Web of Science,
97 November 2022).

## 98 2. Aqueous dispersion **RAFT** polymerization.

99 RAFT polymerization is a conventional radical polymerization with a controlled and living behavior 100 reached by adding a chain transfer agent (CTA) to the reaction medium. It involves rapid reversible chain 101 transfer between the CTA and a growing macroradical *via* a degenerative chain transfer mechanism.[22, 102 27] **Table 1** shows an overview of the systems and the experimental conditions reported up today to 103 conduct aqueous dispersion RAFT PISA.

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Table 1. Aqueous dispersion RAFT PISA of different water-soluble monomers from various steric stabilizer blocks.
 TI: thermal initiated, PI: photo-initiated, EI: enzymatic initiated, DD: direct dissociation, PET: photo electron
 transfer. See Fig. 2-6 for the abbreviations of CTA, initiation systems, steric stabilizer blocks, monomers and nano objects morphology, respectively.

Monomer	Steric stabilizer block	RAFT	experimental conditions	Morphology	ref
	PGMA	TI	CTA-1, CTA-3, CTA-4, CTA-6, V-501 (70°C), V-50 (56°C)	S, WLM, ULV, BW, J, OM, FV	[43, 114- 123]
НРМА		PI	CTA-4, CTA-7, CTA-10 SPTP (405nm, 0.095-17.1 mW/cm <sup>2</sup> , 0°C, 25°C, 70°C). SBAPO (465nm, 3.0 mW/cm <sup>2</sup> , RT). SPTP (405nm 0.45 mW/cm <sup>2</sup> , 25°C, 60°C)	S, WLM, ULV, TV, DV	[124- 128]

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PIO         CTA-7, CTA-10, PP-OH (405nn, 160 – 800 mW, 37°C) SYPT (405nn, 0.4-0.5 mW/cm <sup>2</sup> , 25°C, 55°C)         S, WLM, ULV, BW, L, 135- GULV, MLV         1132, 139]           PEO         DD         CTA-7, CTA-10 (365nn, RT), (405nn, 37°C), (460nn, 0.8-1.6 mW/cm <sup>2</sup> , R1)         S, WLM, ULV, BW, L, GULV, MLV         1132, 140]           PEO         DD         CTA-7, CTA-10 (365nn, RT), (405nn, 30°C), ZpTMPyP (595nn, 10.2 mW/cm <sup>2</sup> , RT).         S, WLM, ULV, MLV         1143, 140]           PET         Rubpy) (450nn, 30°C), ZpTMPyP (595nn, 10.2 mW/cm <sup>2</sup> , RT).         S, WLM, ULV, MLV         1144, 146]           DD         CTA-7, CTA-10, CTA-16         S, WLM, ULV         1144, 146]           PFEGMA         PF1         SPTP (405mn, 0.6 mW/cm <sup>2</sup> , 42.5°C), (530 nn, 10 mW/cm <sup>2</sup> , RT)         S, WLM, ULV         1149, mW/cm <sup>2</sup> , RT)           PDEGMA         PF1         CTA-10, CTA-16         1127, CTA-10, CTA-10, CTA-16         1127, S, WLM, ULV         1141, 1460           PD         CTA-9, (365nn, 4-27 mW/cm <sup>2</sup> , RT)         S, WLM, ULV         1143, NOM, WIM, ULV, ULV         1151, 155, CTA-10, (365nn, 37°C)         S, WLM, ULV         1157, 156, SNPs           DD         CTA-40, (405nn, 10-60 mW/cm <sup>2</sup> , 20-60°C)         M, WLM, ULV, ULV         1157, 157, 158, CTA-10, (365ann, 37°C)         S, WLM, ULV, ULV         1151, 159, CTA-10, (365ann, 37°C)         SNPs         1161, 157, 158, SNPs         1161, 157, 158, SNPs         116	$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$		1341	
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$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	I	135-	
PEO         DD         CTA-7, CTA-10 (365nm, RT), (405nm, 37°C), (460nm, 0.8-1.6 mW/cm <sup>2</sup> , RT).         S, WLM, ULV, BW, L, GULV, MLV         [132, 140- GULV, MLV           PET         CTA-7, CTA-10 PET         FR (hopp), (450nm, 37°C), 2nTMPyP (595nm, 10.2 mW/cm <sup>2</sup> , RT).         S, WLM, ULV, MLV         [143- rate]           PET         CTA-7, CTA-10         S, WLM, ULV, MLV         [143- rate]           PET         CTA-7, CTA-10, CYA-16         S, WLM, ULV         [147]           PFEGMA         PI         SPIP (405m, 0.5 mW/cm <sup>2</sup> , 4.25°C), (530 nm, 10 mW/cm <sup>2</sup> , RT)         S, WLM, ULV         [148]           PFEGMA         PI         SPIP (405m, 0.5 mW/cm <sup>2</sup> , 4.25°C), ULV         [151]           PPEGMA         D         CTA-9, (655m, 4.27 mW/cm <sup>2</sup> , RT)         M, OM, WLM, ULV         [157]           Dextran         DD         CTA-10, (465m, 10-660 mW/cm <sup>2</sup> , 20-66°C)         M, WLM, ULV, WCV         [157]           PSar         CTA-10, (365nm, 87°C)         S, WLM, ULV, ULV         [160]         [161]           PMMEE         CTA-10, (465m, 37°C)         S, WLM, ULV, ULV         [162]           PNMEP         CTA-4, V-404 (44°C)         SNPs         [161]           PNMEP         CTA-4, V-404 (44°C)         SNPs         [162]           PNMEP         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV,	EO DD $\frac{CTA-7, CTA-10}{(365nm, RT), (405nm, 37°C), (460nm, 0.8-1.6 mW/cm2, S, WLM, ULV, BW, L, RT), CTA-7, CTA-10 PET Rubps),(450nm, 30°C), ZaTMPyP (595nm, 10.2 mW/cm2, RT), ZaPCS, C730nm, 60mW/cm2, RT) EI CTA-7, CTA-10, CTA-10 PET Rubps),(450nm, 30°C), ZaTMPyP (595nm, 10.2 mW/cm2, RT), ZaPCS, C730nm, 60mW/cm2, RT) S, WLM, ULV, MLV [1] EI CTA-7, CTA-10, CTA-16 PET Rubps, 23°C) S, WLM, ULV [1] Nono CTA-10 (090 Hz, 23°C) S, WLM, ULV [1] RT CTA-110 (090 Hz, 23°C) S, WLM, ULV [1] PEGMA PI CTA-16, KPS/NAAs (30°C) ULV [1] HPMAc TI CTA-3, V-501 (70°C) S, WLM, ULV [1] CTA-12, TPO-L (405nm, 10-60 mW/cm2, 20-60°C) M, WLM, ULV, MCV [1] CTA-10, G56nm, RT) S, WLM, ULV, ULV [1] SA PET CTA-13, Rubpsyl, CT, 460 nm, 37°C) S, WLM, EUV, ULV [1] SA PET CTA-13, Rubpsyl, CT, 460 nm, 37°C) S, WLM, EUV, ULV [1] SA PET CTA-13, Rubpsyl, CT, 460 nm, 37°C) S, WLM, EUV, ULV [1] CTA-2, SPTP, 405 nm, 0.5 mW/cm2, 27T) S, NPs [1] VMPC CTA-1, CTA-4, V-501 (70°C) S, WLM, ULV, II] CTA-2, SPTP, 405 nm, 0.5 mW/cm, (GON), RT S, W [1] CTA-4, V-501 (70°C) S, WLM, ULV [1] SBMA CTA-1, V-501 (70°C) S, WLM, ULV [1] SBMA CTA-4, V-501 (70°C) S, WLM, ULV [1] SBMA PDMAA CTA-4, V-501 (70°C) S, WLM, ULV [1] SBMA PDMAA CTA-4, V-501 (70°C) S, WLM, ULV [1] SBMA PDMAA CTA-4, V-501 (70°C) S, WLM, ULV [1] SBMA PDMAA CTA-4, V-501 (70°C) S, WLM, ULV [1] SBMA PDMAA CTA-4, V-501 (70°C) S, WLM, ULV [1] SBMA PDMAA CTA-4, V-501 (70°C) S, WLM, ULV [1] SBMA PDMAA CTA-4, V-501 (70°C) S, WLM, ULV $	-	1391	
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$ \begin{array}{c} \mbox{PEO} \\ \mbod{PEO} \\ \mbod{PEO}$	$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		[120	
PEO         DD         GSDIM, R17, (400MM, 05-1.6 MW/cm <sup>2</sup> , GULV, MLV         1421           TA-7, CTA-10         TTA-7, CTA-10         S, WLM, ULV, MLV         [143]           EI         CTA-7, CTA-10         S, WLM, ULV, MLV         [144]           EI         CTA-7, TRP, ZaPC S, (250mm, 10.2)         S, WLM, ULV         [147]           Sono         CTA-10 (990 KHz, 23°C)         S, WLM, ULV         [147]           PFEGMA         PI         SPTP (405mn, 0.5 mW/cm <sup>2</sup> , 425°C), (530 nm, 10         S, WLM, ULV         [147]           PMW(cm <sup>2</sup> , RT)         S, WLM, ULV         [147]         [153]         [127]         [127]           PHPAc         TI         CTA-3, V.501 (70°C)         ULV         [153]           PAT         PTO-1, (405nm, 10-60 mW/cm <sup>2</sup> , 20-60°C)         M, WLM, ULV, MLV         [145]           PAT         CTA-12, TPO-1, (405nm, 10-60 mW/cm <sup>2</sup> , 20-60°C)         M, WLM, ULV, WCV         [157]           CTA-12, TPO-1, (405nm, 37°C)         S, WLM, ULV, ULV         [140]           Sar         CTA-4, VA-044 (44°C)         SNPs         [161]           PNMEP         CTA-4, VA-044 (44°C)         SNPs         [162]           PNMPC         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV, ULV         [164]           PSBMA			[132,	
$\begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	K1).         T<         T         T         T         T         T<         T< <th< td=""><td>.0 1</td><td>140-</td><td></td></th<>	.0 1	140-	
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P11         Ku(pp)(430m, 30°), 20°, 20°, MMPy (350m, 10.2         S, WLM, ULV         [146]           WiCm <sup>2</sup> , RT, JHKPACACHAO, (RT)         S, WLM, ULV         [147]           Sono         CTA-10, Q00 MHz, 23°C)         S, WLM, ULV         [147]           PD         CTA-7, CTA-10, CTA-16         [127, PEGMA         [127, PTA-10, Q00 MHz, 23°C)         S, WLM, ULV         [149]           PHPMAc         TI         CTA-3, CTA-16, GV SNMA, G3°C)         ULV         [151]           PHPMAc         TI         CTA-3, C30 (70°C)         S, WLM, ULV         [151]           PATA         CTA-3, G56m, 4-27 mW/cm <sup>2</sup> , RT)         M, OM, WLM, ULV, MCV         [151]           PDMAEMA         PI         CTA-42, (365m, 4-27 mW/cm <sup>2</sup> , RT)         SNPs         [161]           SN PS         CTA-10, (365m, RT)         S, WLM, ULV, MCV         [159]           PDMAEMA         PI         CTA-13, Ru(pp) <sub>2</sub> Cl <sub>2</sub> , (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         [161]           SN K         CTA-4, V-501 (70°C), VA-044 (40°C)         SNPs         [162]         [130,           PMPC         CTA-4, V-501 (70°C)         S, WLM, ULV, LR         [164]           PGSMA         CTA-4, V-501 (70°C)         S, WLM, ULV, LR         [164]           PGMA         CTA-4, V-501 (70°C)         <	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Ŧ	[143-	
$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	Image: Big Strain Str	1	146]	
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$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Solution         CTA-10 (300 kTI2, 25 C)         S, WLM, ULV         II           CTA-7, CTA-10, CTA-16         FTP (405nm, 0.5 mW(cm <sup>2</sup> , 4-25°C), (530 nm, 10 mW(cm <sup>2</sup> , RT)         S, WLM, ULV         II           RI         CTA-16, KPS/NaAs (30°C)         ULV         II           PEGMA         RI         CTA-16, KPS/NaAs (30°C)         ULV         II           RI         CTA-16, KPS/NaAs (30°C)         ULV         II           PEGMA         DD         CTA-9, (365nm, 4-27 mW(cm <sup>2</sup> , RT)         S, WLM, ULV, II           SNPs         T         CTA-10, (365nm, 87°C)         S, WLM, ULV, MCV         II           Sar         CTA-10, (365nm, 37°C)         S, WLM, ULV, ULV         II         CTA-10, (365nm, 37°C)         S, WLM, ULV, ULV         II           Sar         CTA-4, V-501 (365nm, 37°C)         S, WLM, ULV, ULV         II         CTA-2, VA-044 (44°C)         SNPs         II           SMEM         CTA-4, V-501 (70°C)         S, WLM, ULV, IR         II         CTA-4, V-501 (70°C)         S, WLM, ULV, IR         II           CTA-4, V-501 (70°C)         S, WLM, ULV         II         CTA-4, V-501 (70°C)         S, WLM, ULV         II           SysMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II         CTA-3, V-501 (70°C)         S, WLM,	1	[14/]	-
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$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	т	140	
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$	RI         CTA-16, KPS/NaAs (30°C)         ULV         1           HPMAc         TI         CTA-16, KPS/NaAs (30°C)         S, WLM, ULV         [1]           HPMAc         TI         CTA-3, V-501 (70°C)         S, WLM, ULV         [1]           extran         D         CTA-9, (365nm, 4-27 mW/cm <sup>2</sup> , RT)         M, OM, WLM, ULV, MCV         [1]           SNPs         1         CTA-10, (365nm, KT)         S, WLM, ULV, MCV         [1]           Sar         CTA-12, TPO-L (405nm, 10-60 mW/cm <sup>2</sup> , 20-60°C)         M, WLM, ULV, MCV         [1]           Sar         CTA-10, (365nm, KT)         S, WLM, EULV, ULV         [1]           SA         PET         CTA-13, Rubpy),Cl <sub>2</sub> , (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         [1]           NMEP         CTA-4, V-4044 (44°C)         SNPs         [1]           NA (Nucleic acid)         CTA-4, V-404 (44°C)         SNPs         [1]           QPMA         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CysMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [1]           QDMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [1]           CA-4, V-501 (70°C)         S, WLM, ULV         [1]           QDMA         CTA-4, V-501 (70°C)	EGMA	149-	
PHPMAc         R         CTA-3, V-S01 (70°C)         S, WLM, ULV         [154]           Dextran         DD         CTA-9, (365nm, 4-27 mW/cm <sup>2</sup> , RT)         M, OM, WLM, ULV, II55, IS6]         SNPs         156]           PDMAEMA         PI         CTA-12, TPO-L (405nm, 10-60 mW/cm <sup>2</sup> , 20-60°C)         M, WLM, ULV, MCV         [157]           PSar         CTA-10, 365nm, RT)         S, WLM, ULV         [141]           PSar         CTA-13, Ru(bpy),Cl <sub>2</sub> , (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         [161]           PNMEP         CTA-10, 365nm, RT)         S, WLM, ULV         [141]           PNMEP         CTA-4, VA-044 (44°C)         SNPs         [162]           PNA (Nucleic acid)         CTA-4, VA-044 (44°C)         SNPs         [163]           PMPC         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         164,           POBMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [163]           PQDMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [166]           PGMA + PGB         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [163]           PGMA + PEG         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [170]           PGMA + PEG         CTA-4, V-501 (70°C)         S, WLM, ULV         [171]<	HPMAc         TI         CTA-3, V-501 (70°C)         S, WLM, ULV         [1]           extran         DD         CTA-9, (365nm, 4-27 mW/cm <sup>2</sup> , RT)         M, OM, WLM, ULV, MCV         [1]           DMAEMA         PI         CTA-12, TPO-L (405nm, 10-60 mW/cm <sup>2</sup> , 20-60°C)         M, WLM, ULV, MCV         [1]           Sar         PET         CTA-13, Ru(bpy), CL, (460 nm, 3 mW/cm <sup>2</sup> , RT)         S, WLM, ULV         [1]           SA         PET         CTA-13, Ru(bpy), CL, (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         [1]           YMEP         CTA-4, VA-044 (44°C)         S, WLM, ULV, LR         [1]           YA (Nucleic acid)         CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [1]           YPC         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CA-1, CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CA-4, V-501 (70°C)         S, WLM, ULV         [1]           SEMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]	Ī	[151]	-
$ \begin{array}{c} \mbox{III MAC} & \mbox{III MC} & \mbox$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $		[154]	-
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	bD         CTA-9, (365nm, 4-27 mW/cm <sup>2</sup> , RT)         M, WLM, ULV, CLV, MLM, ULV, MCV         II           MAEMA         PI         CTA-12, TPO-L (405nm, 10-60 mW/cm <sup>2</sup> , 20-60°C)         M, WLM, ULV, MCV         II           Sar         CTA-10 (365nm, RT)         S, WLM, ULV         II         II           Sar         CTA-13, Ru(bpy), CL, (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         II           NMEP         CTA-4, VA-044 (44°C)         SNPs         II           VA (Nucleic acid)         CTA-4, VA-044 (44°C)         SNPs         II           VPC         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         II           CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV         II         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II           CTA-4, V-501 (70°C)         S, WLM, ULV         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II </td <td></td> <td>[155</td> <td>-</td>		[155	-
Dextran         CTA-12, TPO-L (405nm, 10-60 mW/cm <sup>2</sup> , 20-60°C)         M, WLM, ULV, MCV         [157]           PSar         CTA-10 (365nm, RT)         S, WLM, ULV         [141]           PSar         CTA-13, Ru(bpy);Cl <sub>2</sub> , (460 nm, 3 mW/cm <sup>2</sup> , RT)         S, WLM, EULV, ULV         [161]           PNMEP         CTA-4, VA-044 (44°C)         SNPs         [161]           PMRC         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT         S, WLM, ULV, LR         [64, 163]           PMPC         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [64, 165]           PCysMA         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV         [166]           PSBMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [167]           PQDMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [43, 168]           PCoAEMACI         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [44]           PAA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [170]           PKSPMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [171]           PGAA         CTA-4, V-501 (70°C)         S, WLM, ULV         [171]           PGMA+ PGA         CTA-4, V-501 (70°C)         S, WLM, ULV         [171]           PGMA + PGA         CTA-4,	extran         DNAEMA         PI         CTA-12, TPO-L (405nm, 10-60 mW/cm <sup>2</sup> , 20-60°C)         M, WLM, ULV, MCV         II           Sar         CTA-12, TPO-L (405nm, 8T)         S, WLM, ULV         II         CTA-10 (365nm, RT)         S, WLM, ULV, UV         II           SA         PET         CTA-13, Ru(bpy) <sub>3</sub> Cl <sub>2</sub> , (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         [I]           NMEP         CTA-4, VA-044 (44°C)         SNPs         [I]           VA (Nucleic acid)         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT         S, WLM, ULV, LR         II           MPC         CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         II           CysMA         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV         II           CysMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II           CoAEMACI         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II         II           CTA-1, V-501 (70°C)         S, WLM, ULV         II         II           CTA-1, V-501 (70°C)         S, WLM, ULV         II         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II         II           CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV <td< td=""><td>I</td><td>156]</td><td></td></td<>	I	156]	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $		[157-	-
PDMAEMA         PI         CTA-10 ( $365nm, RT$ )         S, WLM, ULV         [141]           PSar         CTA-7 ( $405nm, 37^{\circ}C$ )         S, WLM, EULV, ULV         [160]           BSA         PET         CTA-13, Ru(bpy) <sub>3</sub> CL <sub>2</sub> ( $460 nm, 3 mW/cm^2, RT$ )         SNPs         [161]           PNMEP         CTA-4, VA-044 ( $44^{\circ}C$ )         SNPs         [162]         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOX), RT         S, WLM, ULV, LR         [163]           PMPC         CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [164]           PSBMA         CTA-4, V-501 (70°C)         S, WLM, ULV, LR         [164]           PQDMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [166]           PSBMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [164]           PCoAEMACI         CTA-4, V-501 (70°C)         S, WLM, ULV         [166]           PSBMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [171]           PMAA         CTA-4, V-501 (70°C)         S, SNPs         [162]           POJlysine         CTA-4, V-501 (70°C)         S, WLM, ULV         [166]           PGMA PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [172]           PGMA PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1	DMAEMA         PI         CTA-10 (365nm, RT)         S, WLM, ULV         II           Sar         CTA-7 (405nm, 37°C)         S, WLM, ULV         II           SA         PET         CTA-13, Ru(bpy) <sub>Cl2</sub> , (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         II           VMEP         CTA-4, VA-044 (44°C)         SNPs         II           VA (Nucleic acid)         CTA-2, SPTP, 405 nm, 0.5 mW/cm, (GOx), RT         S, W         II           VPC         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         II           CysMA         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV, LR         II           CysMA         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV         II           Cachemacl         CTA-1, V-501 (70°C)         S, WLM, ULV         II           Cachemacl         CTA-4, V-501 (70°C)         S, SNPs         II           Cachemacl         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         II           Cachemacl         CTA-1, OTA-9, V-501 (70°C)         S, WLM, ULV         II           Cachemacl         CTA-1, OTA-9, V-501 (70°C)         S, WLM         II           Cachemacl         CTA-1, OTA-9, V-501 (70°C)         S, WLM         II           Cachemacl         CTA-4, V-501 (70°C)         S, WLM <td></td> <td>1591</td> <td></td>		1591	
PSar         CTA-7 (405nm, 37°C)         S, WLM, EULV, ULV         [160]           BSA         PET         CTA-13, Ru(bpy) <sub>3</sub> Cl <sub>2</sub> (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         [161]           PNMEP         CTA-4, VA-044 (44°C)         SNPs         [162]           FNA (Nucleic acid)         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOX), RT         S, W         [130,           PMPC         CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         164,           PSBMA         CTA-4, V-501 (70°C)         S, WLM, ULV, IR         164,           PQDMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [167]           PQDMA         CTA-4, V-501 (70°C)         S, SNPs         166]           PCoAEMACI         CTA-3, VA-044 (70°C)         S, WLM, ULV         [170]           Polylysine         CTA-4, V-501 (70°C)         S, SNPs         163]           PCA-4, V-501 (70°C)         S, WLM, ULV         [170]         [171]           PMAA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [171]           PGMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [171]           PGMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [172]           PGMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM, ULV	Sar         CTA-7 ( $405nm, 37^{\circ}C$ )         S, WLM, EULV, ULV         I           SA         PET         CTA-7 ( $405nm, 37^{\circ}C$ )         S, WLM, EULV, ULV         I           SA         PET         CTA-13, Ru(bpy) <sub>3</sub> Cl <sub>2</sub> ( $460 nm, 3 mW/cm^2, RT$ )         SNPs         [1]           NMEP         CTA-4, VA-044 ( $44^{\circ}C$ )         SNPs         [1]           VA (Nucleic acid)         CTA-2, SPTP, $405 nm, 0, 5 mW/cm, (GOx), RT$ S, W         [1]           MPC         CTA-4, VA-044 ( $44^{\circ}C$ )         S, WLM, ULV, LR         [1]           CysMA         CTA-1, CTA-4, V-501 (70°C), VA-044 ( $50^{\circ}C$ )         S, WLM, ULV, LR         [1]           CysMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [1]           CoAEMACI         CTA-1, V-501 (70°C)         S, SNPs         [1]           CoAEMACI         CTA-4, V-501 (70°C)         S, SNPs         [1]           CTA-3, VA-044 (70°C)         S, WLM, ULV         [1]           SEMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [1]           GTA-4, V-501 (70°C)         S, SNPs         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           SEMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV	MAEMA	[141]	-
BSA         PET         CTA-13, Ru(bpy) <sub>3</sub> Cl <sub>2</sub> , (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         [161]           PNMEP         CTA-13, Ru(bpy) <sub>3</sub> Cl <sub>2</sub> , (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         [161]           PNMEP         CTA-4, VA-044 (44°C)         SNPs         [162]           FNA (Nucleic acid)         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT         S, WL         [163]           PMPC         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [64, 165]           PCysMA         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV         [166]           PBBMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [166]           PCosEMACI         CTA-4, V-501 (70°C)         S, WLM, ULV         [167]           Polyisine         CTA-4, V-501 (70°C)         S, SNPs         [168]           CTA-3, VA-044 (70°C)         S, WLM, ULV         [170]           PKSPMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [171]           PMAA         CTA-4, V-501 (70°C)         S, WLM, ULV         [172]           PGMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM, ULV         [174]           PGMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [174]           PGMA + PAMA         CTA-4, V-501 (7	SA         PET         CTA-13, Ru(bpy) <sub>3</sub> Cl <sub>2</sub> , (460 nm, 3 mW/cm <sup>2</sup> , RT)         SNPs         [1]           NMEP         CTA-4, VA-044 (44°C)         SNPs         [1]           NA (Nucleic acid)         CTA-4, VA-044 (44°C)         SNPs         [1]           MPC         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [1]           CysMA         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [1]           CysMA         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV, LR         [1]           CysMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [1]           CysMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [1]           CoAEMACI         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           SemA         CTA-1, V-501 (70°C)         S, WLM         [1]         CTA-4, V-501 (70°C)         S, WLM         [1]           GMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PDAA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]	ar	[160]	-
BSA         PE1         CTA-15, Ku(opy)_3cl <sub>2</sub> (400 min, 3 m/vcm, K1)         SNPs         [161]           PNMEP         CTA-4, VA-044 (4°C)         SNPs         [162]           FNA (Nucleic acid)         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT         S, W         [163]           PMPC         CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [66]           PSBMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [167]           PQDMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [167]           PQDMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [167]           PQDMA         CTA-1, CTA-4, V-501 (70°C)         S, WLM, ULV         [167]           PQDMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [167]           PQDMA         CTA-4, V-501 (70°C)         S, NPs         [43, 168]           POA         CTA-4, V-501 (70°C)         S, WLM, ULV         [170]           PKSPMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [171]           PGMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM, ULV         [172]           PGMA + PAMA         PGMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [171]           PGMA + PQAIMA         CTA-4, V-501 (70°	SA         PET         CTA-15, R(0)p7);C12, (400 lml, 5 lmV;Cln (, K1)         SNPs         [1]           NMEP         CTA-4, VA-044 (44°C)         SNPs         [1]           VA (Nucleic acid)         CTA-4, VA-044 (44°C)         SNPs         [1]           WPC         CTA-4, VA-044 (44°C)         SNPs         [1]           CysMA         CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [1]           CysMA         CTA-4, V-501 (70°C)         S, WLM, ULV, LR         [1]           CysMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [1]           CoAEMACI         CTA-4, V-501 (70°C)         S, SNPs         [4]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         [1]           CTA-4, V-501 (70°C)         S, SNPs         [4]         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         [1]           GTA-4, V-501 (70°C)         S, WLM, U		[160]	-
PNMEP         CTA-4, VA-044 ( $4^{4C}$ )         SNPs         [162]           FNA (Nucleic acid)         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOX), RT         S, W         [130,           PMPC         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [164,           PCysMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [166]           PSBMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [167]           PQDMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [43,           Polylysine         CTA-4, V-501 (70°C)         S, WLM, ULV         [179]           PMAA         CTA-4, V-501 (70°C)         S, WLM, ULV         [170]           PMAA         CTA-4, V-501 (70°C)         S, WLM, ULV         [170]           Polylysine         CTA-4, V-501 (70°C)         S, WLM, ULV         [170]           PMAA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [171]           PMAA         CTA-4, V-501 (70°C)         S, WLM, ULV         [172]           PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [173]           PGMA + PGA         TTI         CTA-4, V-501 (70°C)         S, WLM, ULV         [174]           PGMA + PQDMA         CTA-4, V-501 (70°C)         S, WLM, ULV <t< td=""><td>NMEP         C1A-4, VA-044 (4*C)         SNPs         [1]           VA (Nucleic acid)         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT         S, W         [1]           WPC         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT         S, W         [1]           CysMA         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [1]           CysMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           QDMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [1]           CoAEMACI         CTA-4, V-501 (70°C)         S, SNPs         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-3, V-501 (70°C)         S, WLM         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]</td><td></td><td>[101]</td><td>_</td></t<>	NMEP         C1A-4, VA-044 (4*C)         SNPs         [1]           VA (Nucleic acid)         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT         S, W         [1]           WPC         CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT         S, W         [1]           CysMA         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [1]           CysMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           QDMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [1]           CoAEMACI         CTA-4, V-501 (70°C)         S, SNPs         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-3, V-501 (70°C)         S, WLM         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]		[101]	_
FNA (Nucleic acid)       CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT       S, W       [163]         PMPC       [130,       [130,         PCysMA       CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)       S, WLM, ULV, LR       164;         PSBMA       CTA-1, V-501 (70°C)       S, WLM, ULV       [166]         PQDMA       CTA-1, V-501 (70°C)       S, WLM, ULV       [167]         PQDMA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [43,         PCoAEMACI       CTA-4, V-501 (60°C)       S, WLM, ULV       [170]         PKSPMA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [169]         Polylysine       CTA-4, V-501 (70°C)       S, WLM, ULV       [171]         PMAA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [143,         PGMA       CTA-4, V-501 (70°C)       S, WLM       [172]         PGMA + PEG       TI       CTA-4, V-501 (70°C)       S, WLM       [171]         PGMA + PEG       TI       CTA-4, V-501 (70°C)       S, WLM, ULV       [173]         PGMA + PEG       TI       CTA-4, V-501 (70°C)       S, WLM, ULV       [174]         PGMA + PEG       TI       CTA-4, V-501 (70°C)       S, WLM, ULV       [171]         PGMA + PAMA       CTA-4, V-501 (70°C)       <	NA (Nucleic acid)       CTA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT       S, W       [1]         MPC       (TA-2, SPTP, 405 nm, 0,5 mW/cm, (GOx), RT       S, W       N       [1]         CysMA       (TA-1, CTA-4, V-501 (70°C), VA-044 (50°C)       S, WLM, ULV, LR       [1]         SBMA       (TA-4, V-501 (70°C)       S, WLM, ULV       [1]         QDMA       (TA-1, CTA-9, V-501 (70°C)       S, WLM, ULV       [1]         CoAEMACI       (TA-3, VA-044 (70°C)       S, NPs       [4]         CoAEMACI       (TA-4, V-501 (70°C)       S, NPs       [1]         CTA-1, CTA-9, V-501 (70°C)       S, WLM, ULV       [1]       [1]         SMA       (TA-4, V-501 (70°C)       S, WLM, ULV       [1]         CTA-1, CTA-9, V-501 (70°C)       S, WLM, ULV       [1]         SMA       (TA-4, V-501 (70°C)       S, WLM, ULV       [1]         SMA       (TA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PEG       (TI       (TA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PAMA       (TA-4, V-501 (70°C)       S, WLM, ULV       [1]       (TA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PAMA       (TA-4, V-501 (70°C)       S, WLM, ULV       [1]       (TA-4, V-501 (70°C) <td< td=""><td>IMEP</td><td>[162]</td><td>_</td></td<>	IMEP	[162]	_
PMPC $(130, 164, 164, 164)$ PCysMACTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)S, WLM, ULV, LRPSBMACTA-1, V-501 (70°C)S, WLM, ULVPQDMACTA-1, CTA-9, V-501 (70°C)S, WLM, ULVPQDMACTA-1, CTA-9, V-501 (70°C)S, SNPsPOJylysineCTA-4, V-501 (70°C)S, NPsPolylysineCTA-4, V-501 (70°C)S, NPsPKSPMACTA-1, CTA-9, V-501 (70°C)S, NPsPGSHMACTA-1, CTA-9, V-501 (70°C)S, NPsPGSHMACTA-1, CTA-9, V-501 (70°C)S, NLM, ULVPGMA + PEGTICTA-3, V-501 (70°C)S, WLM, ULVPGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULVPGMA + PAGSHMACTA-4, V-501 (70°C)S, WLM, ULVPGMA-co-MAm-CTA-4, V-501 (70°C)S, WLM, ULVP(GMA-co-MAm-CTA-4, V-501 (70°C)S, WLM, ULVP(GMA-co-GlyMA)CTA-4, V-501 (70°C)F, SpP(GMA-co-CTA-4, V-501 (70°C)F, SpP(GMA-co-CTA-4, V-501 (70°C)WLMP(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULVP(GMA-co-CTA-4, V-501 (70°C)S, WLMP(GMA-co-CTA-4, V-501 (70°C)S, WLMP(GMA-co-CTA-4, V-501 (70°C)S, WLMP(GMA-co-CTA-4, V-501 (70°C)S, WLMP(GMA-co- </td <td>MPC         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [1] (1] (1] (1] (1] (1] (1] (1] (1] (1] (</td> <td>IA (Nucleic acid)</td> <td>[163]</td> <td>_</td>	MPC         CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)         S, WLM, ULV, LR         [1] (1] (1] (1] (1] (1] (1] (1] (1] (1] (	IA (Nucleic acid)	[163]	_
PMPCCTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)S, WLM, ULV, LR164, 165]PCysMACTA-4, V-501 (70°C)S, WLM, ULV1167]PgBMACTA-1, CTA-9, V-501 (70°C)S, SNPs168]PCoAEMACICTA-1, CTA-9, V-501 (70°C)S, SNPs168]PolylysineCTA-4, V-501 (70°C)S, WLM, ULV1170]PKSPMACTA-1, CTA-9, V-501 (70°C)S, SNPs168]CTA-4, V-501 (60°C)S, WLM, ULV1170]PKSPMACTA-1, CTA-9, V-501 (70°C)S, SNPs143,PGSHMACTA-1, CTA-9, V-501 (70°C)S, WLM1172]PGMA + PEGTICTA-4, V-501 (70°C)S, WLM1172]PGMA + PEGTICTA-4, V-501 (70°C)S, WLM, ULV1173]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV1173]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV1174]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV1176]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV1176]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV1176]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV1176]PGMA-roa-GilyMA)CTA-4, V-501 (70°C)S, WLM, ULV1178]P(GMA-co-GilyMA)CTA-3, VA-044 (50°C), pH=6.8S, ULV1130]P(GMA-co-GlyMA)CTA-3, VA-044 (50°C), pH=6.8S, ULV1130]P(GMA-roa-GlyMA)CTA-3, V-501 (70°C)G, WLM1179]P(GMA-roa-GlyMA)CTA-3, V-501 (70°C)S, WLM, ULV1168] </td <td>MPC       CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)       S, WLM, ULV, LR       10         CysMA       CTA-4, V-501 (70°C), VA-044 (50°C)       S, WLM, ULV, LR       10         SBMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         QDMA       CTA-1, CTA-9, V-501 (70°C)       S, WLM, ULV       [1]         CoAEMACI       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [4]         CTA-3, VA-044 (70°C)       SNPs       [1]         CTA-4, V-501 (60°C)       S, WLM, ULV       [1]         KSPMA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [4]         CTA-4, V-501 (60°C)       S, WLM, ULV       [1]         GTA-3, V-501 (70°C)       S, WLM, ULV       [1]         GTA-4, V-501 (70°C)       S, WLM       [1]         CTA-4, V-501 (70°C)       S, WLM       [1]         GMA PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGaISMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGSHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA-co-MAm- FF)       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA-co-GlyM</td> <td></td> <td>[130,</td> <td></td>	MPC       CTA-1, CTA-4, V-501 (70°C), VA-044 (50°C)       S, WLM, ULV, LR       10         CysMA       CTA-4, V-501 (70°C), VA-044 (50°C)       S, WLM, ULV, LR       10         SBMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         QDMA       CTA-1, CTA-9, V-501 (70°C)       S, WLM, ULV       [1]         CoAEMACI       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [4]         CTA-3, VA-044 (70°C)       SNPs       [1]         CTA-4, V-501 (60°C)       S, WLM, ULV       [1]         KSPMA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [4]         CTA-4, V-501 (60°C)       S, WLM, ULV       [1]         GTA-3, V-501 (70°C)       S, WLM, ULV       [1]         GTA-4, V-501 (70°C)       S, WLM       [1]         CTA-4, V-501 (70°C)       S, WLM       [1]         GMA PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGaISMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGSHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA-co-MAm- FF)       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA-co-GlyM		[130,	
PCysMA $[165]$ PCysMACTA-4, V-501 (70°C)S, WLM, ULV $[166]$ PSBMACTA-1, V-501 (70°C)S, WLM, ULV $[167]$ PQDMACTA-1, CTA-9, V-501 (70°C)S, SNPs $[43, 168]$ PCoAEMACICTA-4, V-501 (60°C)SNPs $[169]$ PolylysineCTA-4, V-501 (60°C)S, WLM, ULV $[170]$ PKSPMACTA-1, CTA-9, V-501 (70°C)S, SNPs $[43, 168]$ PGMACTA-1, CTA-9, V-501 (70°C)S, SNPs $[143]$ PMAACTA-1, CTA-9, V-501 (70°C)S, WLM, ULV $[172]$ PSEMACTA-4, V-501 (70°C)S, WLM $[172]$ PGMA + PEGTICTA-4, V-501 (70°C)S, WLM, ULV $[173]$ PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV $[174]$ PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV $[174]$ PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV $[174]$ PGMA + PGAISMACTA-4, V-501 (70°C)S, WLM, ULV $[174]$ PGMA + PGAISMACTA-4, V-501 (70°C)S, WLM, ULV $[176]$ PGMA + PGAISMACTA-4, V-501 (70°C)S, WLM, ULV $[176]$ PGMA-co-MAm-CTA-4, V-501 (70°C)S, WLM, ULV $[178]$ P(GMA-co-GiyMA)CTA-3, V-501 (70°C)F, Sp $[178]$ P(GMA-co-CTA-4, V-501 (70°C)VLM $[179]$ P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV $[168]$ P(GMA)PCTA-3, V-501 (70°C)VLM $[168]$ P(KSPMA-c0-CTA-4, V-501 (70°C)S, WLM, ULV <t< td=""><td>CysMAInterpret (CTA-4, V-501 (70°C))S, WLM, ULVInterpret (TA-4, V-501 (70°C))BBMACTA-1, V-501 (70°C)S, WLM, ULVIQDMACTA-1, CTA-9, V-501 (70°C)S, SNPsICoAEMACICTA-3, VA-044 (70°C)SNPsIDylysineCTA-4, V-501 (60°C)S, WLM, ULVIKSPMACTA-1, CTA-9, V-501 (70°C)S, SNPsIMAACTA-1, CTA-9, V-501 (70°C)S, WLMIGMA + PEGCTA-4, V-501 (70°C)S, WLMIGMA + PAAACTA-4, V-501 (70°C)S, WLM, ULVIGMA + PAAACTA-4, V-501 (70°C)S, WLM, ULVIGMA + PQBMACTA-4, V-501 (70°C)S, WLM, ULVIGMA + PGAISMACTA-4, V-501 (70°C)S, WLM, ULVIGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULVIGMA-co-Ghyma)CTA-4, V-501 (70°C)S, WLM, ULVIGMA-co-Giyma)CTA-4, V-501 (70°C)S, WLM, ULVIGMA-co-Giyma)CTA-3, V-501 (70°C)S, WLM, ULVIGMA-co-Ghyma)CTA-4, V-501 (70°C)S, WLM, ULVIGMA-co-BmA)CTA-4, V-501 (70°C)S, WLMIGMA-co-BmA)CTA-4, V-501 (70°C)S, WL</td><td>1PC</td><td>164,</td><td></td></t<>	CysMAInterpret (CTA-4, V-501 (70°C))S, WLM, ULVInterpret (TA-4, V-501 (70°C))BBMACTA-1, V-501 (70°C)S, WLM, ULVIQDMACTA-1, CTA-9, V-501 (70°C)S, SNPsICoAEMACICTA-3, VA-044 (70°C)SNPsIDylysineCTA-4, V-501 (60°C)S, WLM, ULVIKSPMACTA-1, CTA-9, V-501 (70°C)S, SNPsIMAACTA-1, CTA-9, V-501 (70°C)S, WLMIGMA + PEGCTA-4, V-501 (70°C)S, WLMIGMA + PAAACTA-4, V-501 (70°C)S, WLM, ULVIGMA + PAAACTA-4, V-501 (70°C)S, WLM, ULVIGMA + PQBMACTA-4, V-501 (70°C)S, WLM, ULVIGMA + PGAISMACTA-4, V-501 (70°C)S, WLM, ULVIGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULVIGMA-co-Ghyma)CTA-4, V-501 (70°C)S, WLM, ULVIGMA-co-Giyma)CTA-4, V-501 (70°C)S, WLM, ULVIGMA-co-Giyma)CTA-3, V-501 (70°C)S, WLM, ULVIGMA-co-Ghyma)CTA-4, V-501 (70°C)S, WLM, ULVIGMA-co-BmA)CTA-4, V-501 (70°C)S, WLMIGMA-co-BmA)CTA-4, V-501 (70°C)S, WL	1PC	164,	
PCysMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [166]         PSBMA       CTA-1, V-501 (70°C)       S, WLM, ULV       [167]         PQDMA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [43, 168]         PCoAEMACI       CTA-3, VA-044 (70°C)       SNPs       [169]         Polylysine       CTA-4, V-501 (60°C)       S, WLM, ULV       [170]         PKSPMA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [43, 171]         PMAA       CTA-1, V-501 (70°C)       S, WLM       [172]         PSEMA       CTA-1, V-501 (70°C)       S, WLM       [173]         PGMA + PEG       TI       CTA-4, V-501 (70°C)       S, WLM, ULV       [173]         PGMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [174]         PGMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [174]         PGMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [174]         PGMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [177]         PGMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [177]         PGMA + PGBISMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [176]         PGMA + PGSHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [178] <td>CysMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           SBMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [1]           QDMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [1]           CoAEMACI         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [4]           bylysine         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (60°C)         S, WLM, ULV         [1]           KSPMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [4]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           MAA         CTA-1, V-501 (70°C)         S, WLM         [1]           SBMA         CTA-1, V-501 (70°C)         S, WLM         [1]           GMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGaISMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV         [1</td> <td></td> <td>165]</td> <td>_</td>	CysMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           SBMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [1]           QDMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [1]           CoAEMACI         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [4]           bylysine         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (60°C)         S, WLM, ULV         [1]           KSPMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [4]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           MAA         CTA-1, V-501 (70°C)         S, WLM         [1]           SBMA         CTA-1, V-501 (70°C)         S, WLM         [1]           GMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGaISMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]         CTA-4, V-501 (70°C)         S, WLM, ULV         [1		165]	_
PSBMACTA-1, V-501 (70°C)S, WLM, ULV[167]PQDMACTA-1, CTA-9, V-501 (70°C)S, SNPs[43, 168]PCoAEMACICTA-3, VA-044 (70°C)SNPs[169]PolylysineCTA-4, V-501 (60°C)S, WLM, ULV[170]PKSPMACTA-1, CTA-9, V-501 (70°C)S, SNPs[43, 171]PMAACTA-1, CTA-9, V-501 (70°C)S, SNPs[43, 171]PMAACTA-1, V-501 (70°C)S, WLM[172]PSEMACTA-4, V-501 (70°C)SS[43]CTA-4, V-501 (70°C)SS[43]CTA-4, V-501 (70°C)SS[43]CTA-4, V-501 (70°C)SS[166]PGMA + PEGTICTA-4, V-501 (70°C)S, WLM, ULV[173]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV[174]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV[175]PGMA + PGaISMACTA-4, V-501 (70°C)S, WLM, ULV[176]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[176]PGMA-co-GlyMA)CTA-4, V-501 (70°C)S, WLM, ULV[176]PGMA-co-GlyMA)CTA-4, V-501 (70°C)F, Sp[178]P(GMA-co-GlyMA)CTA-3, V-501 (70°C)WLM[179]P(GMA-co-GlyMA)CTA-3, V-501 (70°C)ULV[180]P(GMA-co-GlyMA)CTA-4, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-GlyMA)CTA-3, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-GlyMA)CTA-3, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-G	SBMA         CTA-1, V-501 (70°C)         S, WLM, ULV         [4]           QDMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [4]           CoAEMACI         CTA-3, VA-044 (70°C)         SNPs         [1]           Dylysine         CTA-4, V-501 (60°C)         S, WLM, ULV         [1]           KSPMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [1]           MAA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [4]           MAA         CTA-1, CTA-9, V-501 (70°C)         S, WLM, ULV         [1]           MAA         CTA-1, V-501 (70°C)         S, WLM         [1]           GMA         CTA-4, V-501 (70°C)         S, WLM         [1]           GMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAGISMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGBISMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGBHMA         CT	CysMA	[166]	_
PQDMA $CTA-1, CTA-9, V-501 (70^{\circ}C)$ S, SNPs[43, 168]PCoAEMAC1 $CTA-3, VA-044 (70^{\circ}C)$ $SNPs$ [169]Polylysine $CTA-4, V-501 (60^{\circ}C)$ $S, WLM, ULV$ [170]PKSPMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [171]PMAA $CTA-3, V-501 (70^{\circ}C)$ $S, WLM$ [172]PSEMA $CTA-3, V-501 (70^{\circ}C)$ $S, WLM$ [172]PSEMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM$ [172]PGMA + PEG $TI$ $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [173]PGMA + PAMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [173]PGMA + PAMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [175]PGMA + PQDMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [176]PGMA + PGalSMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [176]PGMA + PGSHMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [176]PGMA + PGSHMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [176]PGMA + PGSHMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [176]PGMA + PGSHMA $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [176]PGMA-co-GlyMA) $CTA-3, V-501 (70^{\circ}C)$ $F, Sp$ [178]P(GMA-co-GlyMA) $CTA-3, V-501 (70^{\circ}C)$ $VLM$ [179]P(GMA-co-GlyMA) $CTA-4, V-501 (70^{\circ}C)$ $VLM$ [179]P(GMA-co-GlyMA) $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [168]P(KSPMA-core $V-501 (70^{\circ}C)$ $S, WLM, ULV$ [168]P(GMA-co-	QDMACTA-1, CTA-9, V-501 (70°C)S, SNPs[4]CoAEMACICTA-3, VA-044 (70°C)SNPs[1]chylysineCTA-4, V-501 (60°C)S, WLM, ULV[1]KSPMACTA-1, CTA-9, V-501 (70°C)S, SNPs[4]MAACTA-1, CTA-9, V-501 (70°C)S, SNPs[4]CTA-3, V-501 (70°C)S, SNPs[4]MAACTA-1, CTA-9, V-501 (70°C)S, WLMSEMACTA-1, V-501 (70°C)S, WLMGMA + PEGTICTA-4, V-501 (70°C)S, WLM, ULVGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULVGMA + PQDMACTA-4, V-501 (70°C)S, WLM, ULVGMA + PGaISMACTA-4, V-501 (70°C)S, WLM, ULVGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULVGMA-co-MAm-FF)CTA-4, V-501 (70°C)S, WLM, ULVGMA-co-GiyMA)CTA-3, V-501 (70°C)F, SpGMA-co-GiyMA)CTA-3, V-501 (70°C)WLMCTA-4, V-501 (70°C)F, SpCTA-4, V-501 (70°C)S, WLM, ULVCTA-4, V-501 (70°C)F, SpCTA-4, V-501 (70°C)F, SpCTA-4, V-501 (70°C)WLMCTA-4, V-501 (70°C)VLMCTA-4, V-501 (70°C)S, WLM, ULVCTA-4, V-501 (70°C)S, WLM, ULVCTA-4, V-501 (70°C)F, SpCTA-4, V-501 (70°C)VLMCTA-4, V-501 (70°C)S, WLM, ULVCTA-4, V-501 (70°C)S, WLM, ULVCTA-4, V-501 (70°C)S, WLM, ULVCTA-4, V-501 (70°C)S, WLM, ULVCTA-4, V-501 (70°C)S, WLM,	BMA	[167]	
PGDMACTA-1, CTA-9, V-301 (70 °C)S, SINPS168]PCoAEMACICTA-3, VA-044 (70°C)SNPs[169]PolylysineCTA-4, V-501 (60°C)S, WLM, ULV[170]PKSPMACTA-1, CTA-9, V-501 (70°C)S, SNPs[43]PMAACTA-1, CTA-9, V-501 (70°C)S, WLM[172]PSEMACTA-1, V-501 (70°C)S, WLM[172]PGMA + PEGTICTA-4, V-501 (70°C)S[43]PGMA + PEGTICTA-4, V-501 (70°C)S, WLM, ULV[173]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV[174]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV[175]PGMA + PGalSMACTA-4, V-501 (70°C)S, WLM, ULV[176]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[177]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[176]PGGMA-co-MAm-CTA-4, V-501 (70°C)S, WLM, ULV[176]P(GMA-co-GlyMA)CTA-4, V-501 (70°C)S, WLM, ULV[178]P(GMA-co-GlyMA)CTA-4, V-501 (70°C)F, Sp[178]P(GMA-co-CTA-4, V-501 (70°C)WLM[179]P(GMA-co-CTA-3, V-501 (70°C)WLM[179]P(GMA-co-CTA-3, V-501 (70°C)WLM[180]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[166]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[166]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168] </td <td>QDMA       C1A-1, C1A-3, V-301 (10 C)       S, SNFS       10         CoAEMACI       CTA-3, VA-044 (70°C)       SNPs       [1         Dylysine       CTA-4, V-501 (60°C)       S, WLM, ULV       [1         KSPMA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [4         MAA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [4         MAA       CTA-1, V-501 (70°C)       S, WLM, ULV       [1         GMA + PEG       TI       CTA-4, V-501 (70°C)       S, WLM       [1         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PGalSMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PGBHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PGBIMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PGBIMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + Co-BIMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA-co-GlyMA)<td></td><td>[43,</td><td></td></td>	QDMA       C1A-1, C1A-3, V-301 (10 C)       S, SNFS       10         CoAEMACI       CTA-3, VA-044 (70°C)       SNPs       [1         Dylysine       CTA-4, V-501 (60°C)       S, WLM, ULV       [1         KSPMA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [4         MAA       CTA-1, CTA-9, V-501 (70°C)       S, SNPs       [4         MAA       CTA-1, V-501 (70°C)       S, WLM, ULV       [1         GMA + PEG       TI       CTA-4, V-501 (70°C)       S, WLM       [1         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PGalSMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PGBHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PGBIMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + PGBIMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA + Co-BIMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1         GMA-co-GlyMA) <td></td> <td>[43,</td> <td></td>		[43,	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	CoAEMACI         CTA-3, VA-044 (70°C)         SNPs         [1]           Dylysine         CTA-4, V-501 (60°C)         S, WLM, ULV         [1]           KSPMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [4]           MAA         CTA-3, V-501 (70°C)         S, SNPs         [4]           SEMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM         [1]           GSHMA         CTA-4, V-501 (70°C)         S, WLM         [1]           GMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGBIMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGBIMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA-co-GlyMA)         CTA-4, V-501 (70°C)         F, Sp         [1]           GMA-co-GlyMA)         C	<b>ZD</b> MA	168]	
PolylysineCTA-4, V-501 (60°C)S, WLM, ULV[170]PKSPMACTA-1, CTA-9, V-501 (70°C)S, SNPs $[43, 171]$ PMAACTA-3, V-501 (70°C)S, WLM[172]PSEMACTA-4, V-501 (70°C)S, WLM[172]PGMA PEGTICTA-4, V-501 (70°C)S[43]PGMA + PEGTICTA-4, V-501 (70°C)SNPs[166]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV[173]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV[174]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV[175]PGMA + PGalSMACTA-4, V-501 (70°C)S, WLM, ULV[177]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[177]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[178]PGMA-co-MAm-CTA-4, V-501 (70°C)F, Sp[178]P(GMA-co-GlyMA)CTA-4, V-501 (70°C)F, Sp[178]P(GMA-co-GlyMA)CTA-3, V-501 (70°C)ULV[180]P(GMA-co-CTA-4, V-501 (70°C)VLM[179]P(GMA-co-CTA-4, V-501 (70°C)VLM[179]P(GMA-co-CTA-4, V-501 (70°C)VLM[168]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-CTA-4, V-501 (70°C)VLM[168]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]P(GMA-co	blylsine         CTA-4, V-501 (60°C)         S, WLM, ULV         [4]           KSPMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [4]           MAA         CTA-3, V-501 (70°C)         S, WLM         [1]           SEMA         CTA-1, CTA-9, V-501 (70°C)         S, WLM         [1]           GSHMA         CTA-1, V-501 (70°C)         S, WLM         [1]           GMA + PEG         TI         CTA-4, V-501 (70°C)         S         [4]           GMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGalSMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + Co-MAm- FF)         CTA-4, V-501 (70°C)         F, Sp         [1]           GMA-	CoAEMACl	[169]	
PKSPMACTA-1, CTA-9, V-501 (70°C)S, SNPs $\begin{bmatrix} 43, \\ 171 \end{bmatrix}$ PMAACTA-3, V-501 (70°C)S, WLM $\begin{bmatrix} 172 \end{bmatrix}$ PSEMACTA-1, V-501 (70°C)SS $\begin{bmatrix} 43 \end{bmatrix}$ PGSHMACTA-4, V-501 (70°C)SS $\begin{bmatrix} 43 \end{bmatrix}$ PGMA + PEGTICTA-4, V-501 (70°C)SS $\begin{bmatrix} 43 \end{bmatrix}$ PGMA + PAMACTA-4, V-501 (70°C)SSWLM $\begin{bmatrix} 172 \end{bmatrix}$ PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV $\begin{bmatrix} 173 \end{bmatrix}$ PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV $\begin{bmatrix} 177 \end{bmatrix}$ PGMA + PQDMACTA-4, V-501 (70°C)S, WLM, ULV $\begin{bmatrix} 177 \end{bmatrix}$ PGMA + PGalSMACTA-4, V-501 (70°C)S, WLM, ULV $\begin{bmatrix} 177 \end{bmatrix}$ PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV $\begin{bmatrix} 177 \end{bmatrix}$ PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV $\begin{bmatrix} 178 \end{bmatrix}$ P(GMA-co-MAm-CTA-4, V-501 (70°C)WLM $\begin{bmatrix} 179 \end{bmatrix}$ P(GMA-co-GlyMA)CTA-3, V-501 (70°C)WLM $\begin{bmatrix} 179 \end{bmatrix}$ P(GMA-co-GlyMA)CTA-3, V-501 (70°C)VLV $\begin{bmatrix} 180 \end{bmatrix}$ P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV $\begin{bmatrix} 168 \end{bmatrix}$ P(SSPMA-co-CTA-4, V-501 (70°C)S, WLM, ULV $\begin{bmatrix} 168 \end{bmatrix}$ P(KSPMA-co-S, WLM, ULV $\begin{bmatrix} 168 \end{bmatrix}$	KSPMA         CTA-1, CTA-9, V-501 (70°C)         S, SNPs         [4]           MAA         CTA-1, CTA-9, V-501 (70°C)         S, WLM         [1]           SEMA         CTA-3, V-501 (70°C)         S, WLM         [1]           GSHMA         CTA-1, V-501 (70°C)         S, WLM         [1]           GMA + PEG         TI         CTA-4, V-501 (70°C)         SNPs         [1]           GMA + PEG         TI         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PQDMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PQDMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGalSMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA - co-MAm- FF)         CTA-4, V-501 (70°C)         F, Sp         [1]           GMA-co- DMA)         CTA-4, V-501 (70°C)         KLV         [1]           GMA-	lylysine	[170]	
TKM MACTA-1, CTA-5, V-501 (70°C)S, StA13171]PMAACTA-3, V-501 (70°C)S, WLM[172]PSEMACTA-1, V-501 (70°C)S[43]PGSHMACTA-4, V-501 (70°C)S[43]PGMA + PEGTICTA-4, VA-044 (44°C)S, WLM, ULVPGMA + PAMACTA-4, VA-044 (44°C)S, WLM, ULV[173]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV[174]PGMA + PQDMACTA-4, V-501 (70°C)S, WLM, ULV[175]PGMA + PGalSMACTA-4, V-501 (70°C)S, WLM, ULV[176]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[177]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[176]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[177]PGMA + PGSHMACTA-4, V-501 (70°C)F, Sp[178]P(GMA-co-MAm-CTA-4, V-501 (70°C)F, Sp[178]P(GMA-co-GIMA)CTA-3, VA-044 (50°C)WLM[179]P(GMA-co-CTA-3, VA-044 (50°C)ULV[180]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-CTA-4, V-501 (70°C)ULV[180]P(GMA-co-CTA-4, V-501 (70°C)ULV[168]P(KSPMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]	MAA       CTA-1, CTA-2, V-501 (70°C)       S, SIAT 3       12         MAA       CTA-3, V-501 (70°C)       S, WLM       [1]         SEMA       CTA-4, V-501 (70°C)       S, WLM       [1]         GMA + PEG       TI       CTA-4, V-501 (70°C)       SNPs       [1]         GMA + PEG       TI       CTA-4, V-501 (70°C)       SNPs       [1]         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PQDMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGalSMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGSHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGSHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGSHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA - co-MAm- FF)       CTA-4, V-501 (70°C)       F, Sp       [1]         GMA-co-GlyMA)       CTA-3, V-501 (70°C)       WLM       [1]         (GMA-co- SDMA)       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         (GMA-co- DMA)       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]	SPMΔ	[43,	
PMAACTA-3, V-501 (70°C)S, WLM[172]PSEMACTA-1, V-501 (70°C)S[43]PGSHMACTA-4, V-501 (70°C)SNPs[166]PGMA + PEGTICTA-4, VA-044 (44°C)S, WLM, ULV[173]PGMA + PMAACTA-3, V-501 (70°C)S, WLM, ULV[174]PGMA + PAMACTA-4, V-501 (70°C)S, WLM, ULV[175]PGMA + PQDMACTA-4, V-501 (70°C)S, WLM, ULV[176]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[177]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[177]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[176]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[166]POE + PMPCCTA-4, V-501 (70°C)S, WLM, ULV[166]PGMA-co-MAm-CTA-4, V-501 (70°C)F, Sp[178]P(GMA-co-GlyMA)CTA-3, VA-044 (50°C)WLM[179]P(GMA-co-CTA-3, VA-044 (50°C)ULV[180]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-CTA-4, V-501 (70°C)VLV[168]P(GMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]P(SSPMA-co-CTA-4, V-501 (70°C)S, WLM, ULV[168]P(KSPMA-co-SSSSP(SSPMA-co-SSSSP(SSPMA-co-SSSSP(SSPMA-co-SSSSP(SSPMA-co-SSSSP(SSPMA-	MAA       CTA-3, V-501 (70°C)       S, WLM       [1]         SEMA       CTA-1, V-501 (70°C)       S       [4]         GSHMA       CTA-4, V-501 (70°C)       SNPs       [1]         GMA + PEG       TI       CTA-4, V-501 (70°C)       SNPs       [1]         GMA + PAA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PAMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGBISMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGSHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGSHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA + PGSHMA       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA - co-MAm- FF)       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         GMA-co-GlyMA)       CTA-3, V-501 (70°C)       WLM       [1]         GMA-co-DMA       CTA-4, V-501 (70°C)       WLM       [1]         (KSPMA-co- EMA)       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         (CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         (CTA-4, V-501 (70°C)       S, WLM, UL		171]	
PSEMA         CTA-1, V-501 (70°C)         S         [43]           PGSHMA         TI         CTA-4, V-501 (70°C)         SNPs         [166]           PGMA + PEG         TI         CTA-4, VA-044 (44°C)         S, WLM, ULV         [173]           PGMA + PMAA         CTA-3, V-501 (70°C)         S, WLM, ULV         [174]           PGMA + PAMA         CTA-4, VA-044 (44°C)         S, WLM, ULV         [173]           PGMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [174]           PGMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [175]           PGMA + PGBISMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [176]           PGMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [177]           PGMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [166]           POE + PMPC         CTA-4, V-501 (70°C)         S, WLM, ULV         [166]           P(GMA-co-MAm-GF)         CTA-4, V-501 (70°C)         F, Sp         [178]           P(GMA-co-GlyMA)         CTA-3, VA-044 (50°C)         WLM         [179]           P(GMA-co-QDMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [168]           P(KSPMA-co-QD         CTA-4, V-501 (70°C)         S, WLM, ULV	SEMA         CTA-1, V-501 (70°C)         S         [4]           GSHMA         TI         CTA-4, V-501 (70°C)         SNPs         [1]           GMA + PEG         TI         CTA-4, V-044 (44°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-044 (44°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGBISMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA-co-MAm- FF)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA-co-GiyMA)         CTA-3, V-501 (70°C)         WLM         [1]           GMA-co- DMA)         CTA-4, V-501 (70°C)         WLM         [1]           GMA-co- EMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA-co- EMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]	IAA	[172]	_
PGSHMACTA-4, V-501 (70°C)SNPs[166]PGMA + PEGTICTA-4, VA-044 (44°C)S, WLM, ULV[173]PGMA + PMAACTA-3, V-501 (70°C)S, WLM, ULV[174]PGMA + PAMACTA-4, V-50 (60°C)S, WLM, ULV[175]PGMA + PQDMACTA-4, V-501 (70°C).S, WLM, ULV[176]PGMA + PGalSMACTA-4, V-501 (70°C).S, WLM, ULV[177]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[177]PGMA + PGSHMACTA-4, V-501 (70°C)S, WLM, ULV[166]POE + PMPCCTA-4, V-501 (70°C)S, WLM, ULV[166]P(GMA-co-MAm-GFF)CTA-4, V-501 (70°C)F, Sp[178]P(GMA-co-GlyMA)CTA-3, VA-044 (50°C)WLM[179]P(GMA-co-GlyMA)CTA-3, V-501 (70°C)ULV[180]P(GMA-co-GlyMA)CTA-4, V-501 (70°C)S, WLM, ULV[168]P(GMA-co-QDMA)P(GMA-co-GOC)CTA-4, V-501 (70°C)S, WLM, ULV[168]	GSHMA         CTA-4, V-501 (70°C)         SNPs         [1]           GMA + PEG         TI         CTA-4, VA-044 (44°C)         S, WLM, ULV         [1]           GMA + PMAA         CTA-3, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-4, VA-044 (44°C)         S, WLM, ULV         [1]           GMA + PAMA         CTA-3, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PQDMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGalSMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (70°C)         F, Sp         [1]           GMA-co-MAm- FF)         CTA-4, V-501 (70°C)         F, Sp         [1]           GMA_{43}-co- SDMA)         CTA-3, VA-044 (50°C)         WLM         [1]           (GMA-co- DMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (KSPMA-co- EMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co- DMA)         CTA-4, V-501 (70°C	EMA	[43]	_
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	GMA + PEG $GMA + PMAA$ TI $CTA-4, VA-044 (44°C)$ S, WLM, ULV[1] $GMA + PMAA$ $CTA-3, V-501 (70°C)$ S, WLM, ULV[1] $GMA + PAMA$ $CTA-4, V-50 (60°C)$ S, WLM, ULV[1] $GMA + PQDMA$ $CTA-4, V-50 (60°C)$ S, WLM, ULV[1] $GMA + PGalSMA$ $CTA-4, V-501 (70°C)$ S, WLM, ULV[1] $GMA + PGSHMA$ $CTA-4, V-501 (70°C)$ S, WLM, ULV[1] $DE + PMPC$ $CTA-4, V-501 (70°C)$ S, WLM, ULV[1] $GMA-co-MAm$ - FF) $CTA-4, V-501 (70°C)$ F, Sp[1] $GMA-co-GIyMA)$ $CTA-3, VA-044 (50°C)$ WLM[1] $GMA-co-GDMA$ $CTA-3, VA-044 (50°C)$ WLM[1] $(GMA-co-GDMA)$ $CTA-3, VA-044 (50°C)$ WLM[1] $(GMA-co-BMA)$ $CTA-4, V-501 (70°C)$ S, WLM, ULV[1] $(GMA-co-BMA)$ $CTA-4, V-501 (70°C)$ S, WLM, ULV[1] $(GMA-co-BMA)$ $CTA-4, V-501 (70°C)$ $VLV$ [1] $(GMA-co-BMA)$ $CTA-4, V-501 (70°C)$ $VA 044 (50°C)$ $VA 044 (50°C)$ <tr< td=""><td>SHMA</td><td>[166]</td><td>_</td></tr<>	SHMA	[166]	_
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$GMA + PMAA$ $CTA-3, V-501 (70^{\circ}C)$ S, WLM, ULV[1] $GMA + PAMA$ $CTA-3, V-501 (70^{\circ}C)$ S, WLM, ULV[1] $GMA + PQDMA$ $CTA-4, V-50 (60^{\circ}C)$ S, WLM, ULV[1] $GMA + PGalSMA$ $CTA-4, V-501 (70^{\circ}C)$ S, WLM, ULV[1] $GMA + PGSHMA$ $CTA-4, V-501 (70^{\circ}C)$ S, WLM, ULV[1] $DE + PMPC$ $CTA-4, V-501 (70^{\circ}C)$ S, WLM, ULV[1] $GMA-co-MAm$ - FF) $CTA-4, V-501 (70^{\circ}C)$ F, Sp[1] $GMA-co-GlyMA)$ $CTA-3, VA-044 (50^{\circ}C)$ WLM[1] $GMA_43-co-$ SDMA) $CTA-3, VA-044 (50^{\circ}C)$ ULV[1] $(GMA-co-DMA)$ $CTA-4, V-501 (70^{\circ}C)$ VLM[1] $(GMA-co-DMA)$ $CTA-3, VA-044 (50^{\circ}C)$ ULV[1] $(GMA-co-BMA)$ $CTA-4, V-501 (70^{\circ}C)$ VLM[1] $(GMA-co-DMA)$ $CTA-4, V-501 (70^{\circ}C)$ S, WLM, ULV[1] $(GMA-co-EMA)$ $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [1] $(GMA-co-EMA)$ $CTA-4, V-501 (70^{\circ}C)$ $S, WLM, ULV$ [1] $(GMA-co-EMA)$	GMA + PEG	[173]	_
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	GMA + PAMA         CTA-4, V-50 (60°C)         S, WLM, ULV         [1]           GMA + PQDMA         CTA-4, V-50 (70°C).         S, WLM, ULV         [1]           GMA + PGalSMA         CTA-4, V-501 (70°C).         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           CTA-4, V-501 (70°C)         F, Sp         [1]           (GMA-co-MAm-FF)         CTA-4, V-501 (70°C)         F, Sp         [1]           (GMA-co-GlyMA)         CTA-3, VA-044 (50°C)         WLM         [1]           (GMA-co-SDMA)         CTA-3, VA-044 (50°C)         WLM         [1]           (GMA-co-BDMA)         CTA-4, V-501 (70°C)         VLV         [1]           (GMA-co-DMA)         CTA-4, V-501 (70°C)         VLV         [1]           (GMA-co-DMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co-BMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co-BMA)         CTA-4, V-501 (70°C) pH=5.5         NPs         [1]           (GDE DOECMA)         CTA-4, V-501 (70°C) pH=5.5         NPA MU	BMA + PMAA	[174]	_
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	GMA + PQDMA         CTA-4, V-501 (70°C).         S, WLM, ULV         [1]           GMA + PGalSMA         CTA-4, V-501 (70°C).         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           DE + PMPC         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co-MAm- FF)         CTA-4, V-501 (70°C)         F, Sp         [1]           (GMA-co-GlyMA)         CTA-3, VA-044 (50°C)         WLM         [1]           (GMA-co- SDMA)         CTA-3, VA-044 (50°C)         WLM         [1]           (GMA-co- DMA)         CTA-4, V-501 (70°C)         VLV         [1]           (CTA-4, V-501 (70°C)         ULV         [1]         [1]           (GMA-co- DMA)         CTA-4, V-501 (70°C)         ULV         [1]           (KSPMA-co- EMA)         CTA-4, V-501 (70°C) pH=5.5         NPs         [1]           (CTA-4, V-501 (70°C) pH=5.5         NPs         [1]	BMA + PAMA	[175]	_
$\begin{array}{c ccccc} PGMA + PGalSMA & CTA-4, V-501 (70^{\circ}C) & S, WLM, ULV & [177] \\ \hline PGMA + PGSHMA & CTA-4, V-501 (70^{\circ}C) & S, WLM, ULV & [166] \\ \hline POE + PMPC & CTA-4, V-501 (70^{\circ}C) & S, WLM, ULV & [130] \\ \hline P(GMA-co-MAm-\\ GFF) & CTA-4, V-501 (70^{\circ}C) & F, Sp & [178] \\ \hline P(GMA-co-GlyMA) & CTA-3, VA-044 (50^{\circ}C) & WLM & [179] \\ \hline P(GMA_{43}-co-\\ DSDMA) & CTA-3, V-501 (70^{\circ}C) & ULV & [180] \\ \hline P(GMA-co-\\ QDMA) & CTA-4, V-501 (70^{\circ}C) & S, WLM, ULV & [168] \\ \hline P(KSPMA-co-\\ \hline \end{array}$	GMA + PGalSMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           DE + PMPC         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co-MAm- FF)         CTA-4, V-501 (70°C)         F, Sp         [1]           (GMA-co-GlyMA)         CTA-3, VA-044 (50°C)         WLM         [1]           (GMA-co-SDMA)         CTA-3, VA-044 (50°C)         WLM         [1]           (GMA-co-BDMA)         CTA-3, VA-044 (50°C)         ULV         [1]           (GMA-co-DMA)         CTA-3, V-501 (70°C)         ULV         [1]           (GMA-co-BDMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co-DMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co-BMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co-BMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co-BMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA-co-EMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (GMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]	MA + PQDMA	[176]	_
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	GMA + PGSHMA         CTA-4, V-501 (70°C)         S, WLM, ULV         [1] $OE + PMPC$ CTA-4, VA-044 (50°C), pH=6.8         S, ULV         [1] $(GMA-co-MAm-FF)$ CTA-4, V-501 (70°C)         F, Sp         [1] $(GMA-co-GlyMA)$ CTA-3, VA-044 (50°C)         WLM         [1] $(GMA_{43}-co-SDMA)$ CTA-3, VA-044 (50°C)         WLM         [1] $(GMA-co-BDMA)$ CTA-3, V-501 (70°C)         ULV         [1] $(GMA-co-DDMA)$ CTA-4, V-501 (70°C)         S, WLM, ULV         [1] $(CTA-4, V-501 (70°C))$ CTA-4, V-501 (70°C)         S, WLM, ULV         [1] $(CTA-4, V-501 (70°C))$ CTA-4, V-501 (70°C)         S, WLM, ULV         [1] $(CTA-4, V-501 (70°C))$ CTA-4, V-501 (70°C)         S, WLM, ULV         [1] $(CTA-4, V-501 (70°C))$ CTA-4, V-501 (70°C)         S, WLM, ULV         [1] $(CTA-4, V-501 (70°C))$ CTA-4, V-501 (70°C)         S, WLM, ULV         [1] $(CTA-4, V-501 (70°C))$ CTA-4, V-501 (70°C)         S, WLM, ULV         [1]	MA + PGalSMA	[177]	_
POE + PMPC         CTA-4, VA-044 (50°C), pH=6.8         S, ULV         [130]           P(GMA-co-MAm- GFF)         CTA-4, V-501 (70°C)         F, Sp         [178]           P(GMA-co-GlyMA)         CTA-3, VA-044 (50°C)         WLM         [179]           P(GMA_43-co- DSDMA)         CTA-3, V-501 (70°C)         ULV         [180]           P (GMA-co- QDMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [168]           P(KSPMA-co- QDMA)         P(KSPMA-co- QDMA)         EXECUTE FOR	$\begin{array}{c c} \underline{OE} + \underline{PMPC} & \underline{CTA-4, VA-044 (50^{\circ}C), pH=6.8} & \underline{S, ULV} & [1] \\ \hline GMA-co-MAm-\\ FF) & \underline{CTA-4, V-501 (70^{\circ}C)} & F, Sp & [1] \\ \hline GMA-co-GlyMA) & \underline{CTA-3, VA-044 (50^{\circ}C)} & \underline{WLM} & [1] \\ \hline GMA_{43}^{-}co-\\ SDMA) & \underline{CTA-3, VA-044 (50^{\circ}C)} & \underline{ULV} & [1] \\ \hline CTA-3, V-501 (70^{\circ}C) & \underline{ULV} & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & S, WLM, ULV & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & F, Sp & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & S, WLM, ULV & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & F, Sp & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & S, WLM, ULV & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & F, Sp & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & F, F, Sp & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & F, F, Sp & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & F, F, Sp & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & F, F, Sp & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & F, F, Sp & [1] \\ \hline CTA-4, V-501 (70^{\circ}C) & F, $	MA + PGSHMA	[166]	_
P(GMA-co-MAm- GFF)         CTA-4, V-501 (70°C)         F, Sp         [178]           P(GMA-co-GlyMA)         CTA-3, VA-044 (50°C)         WLM         [179]           P(GMA_43-co- DSDMA)         CTA-3, VA-044 (50°C)         WLM         [179]           P(GMA_43-co- DSDMA)         CTA-3, V-501 (70°C)         ULV         [180]           P(GMA-co- QDMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [168]	$\begin{array}{cccc} (GMA-co-MAm-\\FF) & CTA-4, V-501 (70^{\circ}C) & F, Sp & [1] \\ (GMA-co-GlyMA) & CTA-3, VA-044 (50^{\circ}C) & WLM & [1] \\ (GMA_{43}^{-}co-\\SDMA) & CTA-3, V-501 (70^{\circ}C) & ULV & [1] \\ (GMA-co-\\DMA) & CTA-4, V-501 (70^{\circ}C) & S, WLM, ULV & [1] \\ (KSPMA-co-\\EMA) & CTA-4, V-501 (70^{\circ}C) pH=5.5 & NPs & [1] \\ (CTA-4, V-501 (70^{\circ}C) pH=5$	DE + PMPC	[130]	_
GFF)       CTA-4, V-501 (70°C)       1, 5p°       [170]         P(GMA-co-GlyMA)       CTA-3, VA-044 (50°C)       WLM       [179]         P(GMA_43-co-       CTA-3, V-501 (70°C)       ULV       [180]         P (GMA-co-       QDMA)       CTA-4, V-501 (70°C)       S, WLM, ULV       [168]         P(KSPMA-co-       CTA-4, V-501 (70°C)       S, WLM, ULV       [168]	FF)       CTA-3, VA-044 (50°C)       I, Sp       I         (GMA-co-GlyMA)       CTA-3, VA-044 (50°C)       WLM       [1]         (GMA-co- SDMA)       CTA-3, V-501 (70°C)       ULV       [1]         (GMA-co- DMA)       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         (CTA-4, V-501 (70°C)       CTA-4, V-501 (70°C)       S, WLM, ULV       [1]         (CTA-4, V-501 (70°C)       CTA-4, V-501 (70°C)       NPs       [1]         (CTA-4, V-501 (70°C)       CTA-4, V-501 (70°C)       NPs       [1]	GMA-co-MAm-	[178]	
P(GMA-co-GlyMA)         CTA-3, VA-044 (50°C)         WLM         [179]           P(GMA_43-co- DSDMA)         CTA-3, V-501 (70°C)         ULV         [180]           P (GMA-co- QDMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [168]           P(KSPMA-co- QDMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [168]	(GMA-co-GlyMA)         CTA-3, VA-044 (50°C)         WLM         [1]           (GMA_43-co- SDMA)         CTA-3, VA-044 (50°C)         ULV         [1]           (GMA-co- DMA)         CTA-3, V-501 (70°C)         ULV         [1]           (CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (CTA-4, V-501 (70°C) pH=5.5         NPs         [1]           (CTA-4, V-501 (70°C) pH=5.5         NPs         [1]	FF)	[170]	_
P(GMA <sub>43</sub> -co- DSDMA)         CTA-3, V-501 (70°C)         ULV         [180]           P (GMA-co- QDMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [168]           P(KSPMA-co- QDMA)         P(KSPMA-co- QDMA)         D         (161)	(GMA <sub>43</sub> -co- SDMA)         CTA-3, V-501 (70°C)         ULV         [1]           (GMA-co- DMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (KSPMA-co- EMA)         CTA-4, V-501 (70°C) pH=5.5         NPs         [1]	GMA-co-GlyMA)	[179]	_
DSDMA)         CTA 4, V-501 (70°C)         CTA 4, V-501 (70°C)         S, WLM, ULV         [168]           P(KSPMA-co-         P(KSPMA-co- <t< td=""><td>SDMA)         CTA-4, V-501 (70°C)         CDV         [1]           (GMA-co- DMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (KSPMA-co- EMA)         CTA-4, V-501 (70°C) pH=5.5         NPs         [1]</td><td>GMA<sub>43</sub>-<i>co</i>-</td><td>[180]</td><td></td></t<>	SDMA)         CTA-4, V-501 (70°C)         CDV         [1]           (GMA-co- DMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (KSPMA-co- EMA)         CTA-4, V-501 (70°C) pH=5.5         NPs         [1]	GMA <sub>43</sub> - <i>co</i> -	[180]	
P (GMA-co- QDMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [168]           P(KSPMA-co-	(GMA-co- DMA)         CTA-4, V-501 (70°C)         S, WLM, ULV         [1]           (KSPMA-co- EMA)         CTA-4, V-501 (70°C) pH=5.5         NPs         [1]	SDMA)	[100]	_
QDMA)         Official (100)         S, (120)         [100]           P(KSPMA-co-         Interview         Interview         Interview         Interview	DMA)         CTA 4, V-501 (70°C) pH=5.5         NPs         [1]           (KSPMA-co- EMA)         CTA-4, V-501 (70°C) pH=5.5         NPs         [1]	GMA-co-	[168]	
P(KSPMA-co-	(KSPMA-co- EMA)         CTA-4, V-501 (70°C) pH=5.5         NPs         [1]           (TA-4, V-501 (70°C) pH=5.5         NPs         [1]	DMA)	[100]	_
CTA-4, V-501 (70°C) $nH=5.5$ NPs 11711	EMA) (TA 10 VA 044 (5000) (115 (115 (115 (115 (115 (115 (115 (11	KSPMA-co-	[171]	
	(1) D(1)(1)AA = O(TA + 1) VA + O(A + (CO(C))) = O(VI)A + UV = (1)	EMA)	[ • / • ]	_
<u>β-CD-POEGMA</u> CTA-10, VA-044 (50°C) S, WLM, ULV [181]	CD-POEGMA CTA-10, VA-044 (50°C) S, WLM, ULV	CD-POEGMA	[181]	_
	PEGMA-co- CTA-8, V-50 (52°C) S, WLM, ULV [1	EGMA-co-	[182]	_

	PFBMA				
			CTA-18, CTA-19, CTA-21, CTA-26, CTA-27, CTA-29, CTA-32, V 501 (70°C) V 50 (70°C)	S, WLM, ULV, L	[183- 193]
	PDMA	Ы	$V-501(70^{\circ}C), V-50(70^{\circ}C)$ CTA-18 SPTP (405nm 0.45 mW/cm <sup>2</sup> 70°C)	S TV ULV L	[192]
		<u></u>	CTA-1, CTA-20, Eosin Y/ Ascorbic acid (530nm, 2.65	S, 1 V, OL V, L	[192]
		PET	mW/cm <sup>2</sup> )	S, WLM	195]
DAAM	PEO	TI	CTA-28, V-50 (70°C)	S, WLM, ULV, L	[196]
	PHPMAc	PI	CTA-7, SPTP (405nm, 0.20 mW/cm <sup>2</sup> , 25°C).	S, WLM, ULV, L	[197- 199]
	PVBTMAC	—ті	CTA-26, V-501 (70°C)	S, WLM, ULV	[200]
	PVP	11	CTA-34, V-50 (60°C)	S, ULV	[201]
	PHPMAc- <i>b</i> - PHisMA	PI	CTA-7, SPTP (405–577 nm, 0.20 mW/cm <sup>2</sup> , 25°C)	L	[202]
		TI	CTA-17, CTA-21, CTA-22,	NG	[42,
	PDMA	DI	APS (60°C), V-50 ( $/0^{\circ}$ C) CTA 21 SPTP ( $405$ nm 0.45 mW/cm <sup>2</sup> 37 or 70°C)	S WIM III V	[203]
NIDAm	PPEGA	Sono	CTA-14 (490 kHz 40 W 45°C)	NG	[204]
NIF AIII	PDMA-b-P(DMA-	Dono		<u>a</u>	[203]
	coAA)		СТА-19, V-501 (70°С)	S, WLM, BW, ULV	[206]
	PDEGA-b-PDMA	TI	CTA-27, VA-044 (40°C)	NG	[207]
	PDMA	_	CTA-31, CTA-33, V-50 (60°C)	S, F	[208]
		EI	CTA-7, HRP/ACAC/ $H_2O_2$ (30°C)	SNPs	[209]
MEA	PPEGMA	RI	CTA-7, KPS/NaAs (30 or 40°C)	SNPs	[210]
	II LOWA	EI	CTA-7, HRP/ACAC/H <sub>2</sub> O <sub>2</sub> (30°C)	SNPs	[209]
	PEO	TI	CTA-21, V-501 (70°C)	S, WLM, ULV	[211]
	PGMA		CTA-5, KPS/NaAs, pH=3 (30°C)	S, WLM, ULV	[212]
НВА	PEO	RI	CTA-4, KPS/NaAs, pH=3 (30°C)	S, WLM, ULV	[213]
IIDA	PNAEP		CTA-21, KPS/NaAs, pH=3 (30°C)	S, WLM, ULV	[214]
	PHEA	TI	CTA-23, VA-044 (44°C)	S, WLM, ULV	[215]
	PPEGA	PI	CTA-24, DOXO/TEA (485nm, 2.2 mW/cm <sup>2</sup> , 37°C)	S	[216]
DEAA	PEO-b-PDMA		CTA-21, V-50 (70°C)	NG	[217]
			CTA-21, V-50 (60°C)	NG	[218]
DMAEMA	PEO		CTA-21, V-501 (65°C)	NG	[219]
META-PE		_	CTA-7, V-50 (80°C)	S, WLM, L	[220]
	PPEGMA		CTA-7, V-50 (80°C)	S, WLM, L	[220]
NAT	PNAM	T1	CTA-24, VA-044, (50°C, 90°C)	S, WLM, ULV, L	[221, 222]
PEGMA	PEO		CTA-7, CTA-10, V-50 (70°C)	NG	[223]
MAm-FGD	PGMA	_	<u>CTA-4, V-501 (70°C)</u>	Sp	[224]
CMAm	PDMA		CTA-25, VA-044 (45°C)	S, WLM, ULV	[225, 226]
CysMA	PHPMAc	-PI	CTA-7, SPTP (405nm, 0.20 mW/cm <sup>2</sup> , 25°C)	S, L, ULV	[227]
NAGA	PPEGA		CTA-14, PP-OH (365nm, 3°C)	NG	[228]
AEDIBr	PADEIBr	-ті	CTA-19, V-501 (70°C)	S, WLM, ULV	[229]
	PGMA		CTA-3, VA-044 (50°C)	W	[230]
HPMA +		PI	CTA-7, SPTP (405nm, 0.5 mW/cm <sup>2</sup> , GOx, RT)	S, WLM, ULV, FV	[127]
GlyMA		EI	CTA-7, $HRP/ACAC/H_2O_2$ (RT)	ULV	[129]
	$\frac{PEO + PQDMA}{PEO}$	TI	CTA-4, VA-044 (50°C) pH4.	S, WLM	[231]
	PEO	_	CTA-7, SPTP (405nm, 0.5 mW/cm <sup>2</sup> , RT)	ULV	[138]
	PPEGMA	PI	CTA-10, SPTP (405nm, 0.5 mW/cm <sup>2</sup> , RT)	S, WLM, ULV	[232]
HPMA + DMEAMA	PEO		CTA-7, SPTP (405nm, 0.5 mW/cm <sup>2</sup> , RT)	ULV	[137]
DIVIEANIA		PET	CTA-21, Ru(bpy) <sub>3</sub> Cl <sub>2</sub> ·6H <sub>2</sub> O (9 mW/cm <sup>2</sup> , 25°C)	S, WLM, ULV	[233]
	PGMA		CTA-10, SPTP (405nm, 0.5 mW/cm <sup>2</sup> , GOx, RT).	ULV	[234]
HPMA + METAC	PEO	РI	CTA-7, SPTP (405nm, 0.5 mW/cm <sup>2</sup> , RT)	ULV	[137]
HPMA + MTC	PGMA	TI	CTA-3, V-501 (70°C)	S, WLM, ULV	[235]
HPMA + NiPAM	PEO	PI	CTA-7, SPTP (405nm, 0.5 mW/cm <sup>2</sup> , 37°C)	ULV	[236]

HPMA + RBMA	PPEGMA	PET	CTA-10, CTA-15, RB (595nm, 0.2 mW/cm <sup>2</sup> , RT)	S	[237]
MEA + PEGA		TI	CTA-30, V-50 (70°C)	NG	[238]
AAm + AN	AN	11	CTA-20, VA-044 (45°C)	S, WLM	[239]
DAAM		DET	CTA 11 7nTMDvD (505nm DT)	S WI M	[240]
+PEGMA	TEI	CTA-11, ZhTWH YI (595hhh, K1).	S, WLW	[240]	
HBA +	ΡΟΜΑ	RI	$CT\Delta_2$ KPS/NaAs nH-3 (30°C)	S WIM III V	[241]
DAAM	M	KI	C11-21, Ki 5/10/16, pi1-5 (50 C)	5, WEN, OL V	[241]

109

## 110 **2.1. Chain transfer agent (CTA).**

A judicious selection of the appropriate CTA depending on the nature of the monomer to polymerize is 111 crucial to ensure a good control of the RAFT polymerization.[242] It was reported recently that the control 112 degree of the RAFT polymerization can impact the nano-objects morphology in aqueous dispersion RAFT 113 **PISA.**[243, 244] CTAs are organosulfur compounds containing thiocarbonylthio function and carrying an 114 activating/stabilizing Z-group that influences the addition step, and a fragmenting R-group that influences 115 116 the fragmentation and re-initiation steps. Among the four types of CTAs, dithioesters and trithiocarbonates 117 have been mainly used in aqueous dispersion RAFT PISA (Fig. 2). This may be explained by the nature of 118 the monomers, more activated monomers (MAM) such as (meth)acrylate and (meth)acrylamide that are 119 compatible with those CTAs. To the best of our knowledge, 2-(ethoxycarbonothioyl)sulfanyl propionic 120 acid methyl ester (CTA-34) is the only thiocarbonate-type CTA reported in the literature to mediate the polymerization of a low activated monomer (vinylpyrrolidone, VP) in aqueous dispersion RAFT 121 PISA.[201] Dithiocarbamates have been never reported yet. Fig. 2 shows the wide range of functionalized 122 CTAs reported in the context of aqueous dispersion RAFT PISA. Some of these CTAs have been 123 extensively used, thanks to their commercial availability such as 4-cyanopentanoic acid dithiobenzoate 124 (CTA-1), 2-cyano-2-propyl dithiobenzoate (CTA-3), and 4-cyano-4-(dodecylsul-fanylthio-carbonyl)-125 sulfanylpentanoic acid (CTA-10), or to their facile synthesis route such as 4-cyano-4-(2-phenylethanesul-126 fanylthiocarbonyl)-sulfanylpentanoic acid (CTA-4), 4-cyano-4-(ethylsulfanylthio-carbonyl)-127 sulfanylpentanoic acid (CTA-7). Note that CTA-10 could be considered as a universal CTA model due to 128 129 its compatibility with all RAFT-initiation systems (Table 1). It is well known that the rate of production of radicals affects both the control and the rate of polymerization.[99] Although temperature has been widely 130 used as stimulus since the early works on aqueous dispersion RAFT PISA,[102] other activation ways 131 132 have been successfully investigated such as light, ultrasound, redox, and enzymatic. As shown in Fig. 3, 133 the activation of aqueous dispersion RAFT PISA has been achieved *via* two pathways: i) using an external 134 radical source, and ii) taking advantage of the CTA dissociation.





CN

4-cyano-4-(propylsulfanylthio-

carbonyl)-sulfanylpentanoic acid (CTA-9)

CN

4-cyano-4-(propyllsulfanylthio-

carbonyl)-sulfanylpentanoic acid

mercaptothiazoline amide

(CTA-13)

HO





4-cyanopentanoic acid dithiobenzoate (CTA-1)



4-cyano-4-(2-phenylethylsulfanylthiocarbonyl)sulfanylpropionic acid ethyl ester (CTA-5)



2-cyano-5-((4-fluorophenyl)amino)-5oxopentan-2-yl ethyl carbonotrithioate (CTA-8)



4-cyano-4-(propylsulfanylthiocarbonyl)-sulfanylpentanoic acid N-succinimidyl ester (CTA-12)



4-cyano-4-(dodecylsulfanylthio-carbonyl)sulfanylpentanoic acid Ethyl (2,4,6trimethylbenzoyl) phenylphosphinate ester (CTA-16)



2-(ethylsulfanylthiocarbonyl)-sulfanyl-2-methylpropionic acid (CTA-19)



2-(phenylethylsulfanylthiocarbonyl)sulfanyl-2-methylpropionic acid (CTA-23)



2-(ethylsulfanylthiocarbonyl)- sulfanylpropionic acid methyl ester (CTA-27)



135 (CTA-31) 136 Fig. 2 Chain transfer agents

4-cyano-4-(phenylcarbonothioylthio)pentanoic acid N-succinimidyl ester (CTA-2)







4-cyano-4-(ethylsulfanylthiocarbonyl)-sulfanylpentanoic acid (CTA-7)

4-cyano-4-(2-carboxyethylsulfanylthiocarbonyl)-sulfanylpentanoic acid (CTA-11)

4-cyano-4-(dodecylsulfanylthiocarbonyl)-sulfanylpentanoic acid azidopropyl (CTA-15)



bis (benzylsulfanylthiocarbonyl)sulfanylpentanoic acid ethyl ester (CTA-18)



2-(dodecylsulfanylthiocarbonyl)sulfanyl-2-methylpropionic acid (CTA-21)

2-(butylsulfanylthiocarbonyl)sulfanylpropionic acid ethyl ester (CTA-25)

s<sup>\_C<sub>12</sub>H<sub>25</sub></sup>



(CTA-32) x=1 (R=C<sub>17</sub>H<sub>35</sub>) (CTA-33)

in

aqueous

sulfanyl-2-methylpropionic acid (CTA-22)

2-(2-carboxyethylsulfanylthiocarbonyl)-













RAFT

dispersion

PISA.

bis-(propyl trithiocarbonate) benzyl (CTA-17)

4-cyano-4-(2-phenylethylsulfanylthiocarbonyl)-

sulfanylpentanoic acid morpholinylethyl amide

(CTA-6)

2-(1-isobutylsulfanylthiocarbonyl)sulfanyl-2-methylpropionic acid (CTA-20)

HC

2-(butylsulfanylthiocarbonyl)sulfanylpropionic acid (CTA-24)

ö

2-(ethylsulfanylthiocarbonyl)- sulfanyl-

propionic acid N-succinimidyl ester

(CTA-28)

ő

z

HO

used

2-cyano-5-oxo-5-(prop2-yn-1-ylamino) pentan-2-yldode cylcarbonotrithioate (CTA-14)

4-cyano-4-(dodecylsulfanylthio-

carbonyl)-sulfanylpentanoic acid

(CTA-10)

CN

#### 137 **2.2. RAFT** activation *via* exogeneous radical sources.

This method requires external initiation systems that forms radical to initiate RAFT polymerization. Fig.
3A summarizes the different initiation systems reported in the literature in the context of aqueous dispersion RAFT PISA.

Thermally initiated (TI) RAFT PISA involves the formation of radicals by decomposition upon heating of 141 water-soluble azo compounds such as 4,4 - azobiscyanovaleric acid (V-501), 2,2-azobis(2-(2-imidazolin-2-142 yl)propane) dihydrochloride (VA-044), and 2,2'-azobis(2-methylpropionamidine) dihydrochloride (V-50) 143 that are the commonly thermal initiators employed in aqueous dispersion RAFT PISA (Fig. 3a). PISA 144 145 involving V-501 and V-50 initiators are generally conducted above 60°C regarding their high 10-hour halflife temperature of 69°C and 56°C, respectively. However, VA-044 is suitable to perform PISA at 146 relatively lower temperature (in the range of  $37^{\circ}C - 50^{\circ}C$ ) thanks to its 10-hour half-life temperature of 147 44°C. Due to the facile experimental protocol requested, TI-RAFT polymerization, which is by far the 148 149 most reported technique in PISA, was used in various aqueous dispersion RAFT PISA formulations to produce large set of nano-objects morphologies (Table 1). However, the relatively high temperature 150 requested could unable performing TI-RAFT PISA of monomers forming thermo-sensitive polymers or in 151 the presence of biomolecules.[103, 245] 152

Photo-initiated (PI) RAFT PISA: during the last decade, light has grasped great interest of researchers as a 153 potential alternative to temperature, due to its ability to carry out polymerization under mild reaction 154 conditions (at ambient or below) and for its temporal (by turning on/off the light) and spatial (partial or 155 total irradiation of the reaction medium) control of polymerization.[101, 103, 246-248] In a similar way as 156 157 for TI-RAFT, PI-RAFT involves formation of radicals by decomposition of photoinitiators that absorb 158 light at UV-visible (mostly around 405nm). Phosphine oxides-based compounds, including sodium phenyl-2,4,6-trimethyl-benzoylphosphinate (SPTP), phenylbis(2,4,6-trimethylbenzoyl) phosphine oxide (SBAPO), 159 and ethyl phenyl(2,4,6-trimethylbenzoyl)phosphinate (TPO-L) as well as 2-hydroxy-4-2-(hydroxyl 160 161 ethoxy)-2 methylpropio-phenone (PP-OH) are successfully used in aqueous dispersion PI-RAFT PISA 162 (Table 1). Ultra-fast RAFT polymerization rate was observed, few minutes are needed to achieve full monomers conversion, [132, 158, 192] which is an additional advantage for PI-RAFT PISA compared to 163 TI-RAFT PISA. 164

165 *Other radical sources*: alternative initiation approaches have been investigated to activate aqueous 166 dispersion RAFT PISA under mild conditions. Briefly, due to the relatively low activation energy, Redox-167 initiated (RI) RAFT polymerization[151, 210, 212-214, 241] involving redox reaction between potassium 168 persulfate (KPS) and L-ascorbic acid sodium salt (NaAs) to form radicals has been investigated in the 169 context of aqueous dispersion RAFT PISA at room temperature. In Enzymatic-initiated (EI) RAFT 170 polymerization, horseradish peroxidase (HRP) enzyme has been used to catalyse the oxidation of 171 acetylacetone (ACAC) to ACAC radical by hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). [129, 147, 209] Recently, ultrasound was used as stimulus to activate RAFT polymerization in aqueous dispersion RAFT PISA (Sono-RAFT). In Sono-RAFT, no exogenous initiator is required, because radicals are generated *in-situ* by the homolysis of water. [148, 205]. Although the potential of these approaches for aqueous dispersion RAFT PISA, only few works have been reported in this context. The absence of a stimulus to drive the spatiotemporal control of RI and EI RAFT polymerizations and the specific equipment required for Sono RAFT polymerization are the main drawbacks of these initiation methods compared to the PI-RAFT.[102] 178



Fig. 3 Initiation systems used in aqueous dispersion RAFT PISA. The double-headed red arrows represent where
the species break down to form the radicals.

#### 182 **2.3. RAFT** activation via a photoactivation of CTA.

Alternative pathways for RAFT activation under mild conditions using light as stimulus have been recently
investigated in aqueous dispersion RAFT PISA, thanks to the well-known thiocarbonylthio
photoactivity.[249]

- The first method involves a direct dissociation (DD) of CTA under irradiation, thanks to the high 186 absorption of CTA at UV region (~320 nm,  $\pi \rightarrow \pi^*$  electronic transition) and at visible region (~430 nm n 187  $\rightarrow \pi^*$ ), resulting in the fragmentation of the weakest C–S bond generating a thivl radical and an active 188 carbon radical able to initiate and control the chain propagation (Fig. 3f). In this case, the CTA acts as 189 190 iniferter (initiator-transfer-terminator). This approach was successfully used in many aqueous dispersion 191 RAFT PISA[132, 140-142, 155, 156] as summarized in Table-1. DD of CTA under UV-visible irradiation 192 is an attractive technique since no exogeneous initiator is required to initiate the polymerization. However, 193 a slow RAFT polymerization is communally observed under visible light due to the slow C-S dissociation. 194 Under UV irradiation, CTA can undergo a photolytic degradation resulting in a loose of polymerization 195 control.[250]
- An efficient RAFT activation under visible light radiation has recently been developed by photoredox 196 catalysis to minimize the unwanted photolysis of CTAs. In this approach (Fig. 3g), called photo-electron 197 transfer RAFT polymerization (PET-RAFT),[251-254] the polymerization is carried out by irradiating a 198 199 photocatalyst (PC) under an appropriate wavelength to generate an excited species (PC\*) enables reducing the CTA by an electron transfer into an anion radical and a radical that propagates the polymerization. 200 201 After a few successive additions of monomers, the catalytic cycle is completed by regenerating the dormant polymer chain and the PC in its ground state. [255, 256] Various water-soluble PC such as zinc 202 203 meso-tetra(N-methyl-4-pyridyl) tetrachloride (ZnTMPyP),[144,2401 porphine tris(bipyridine)ruthenium(II) chloride (Ru(bpy)3Cl2),[143, 145, 161, 233] Eosin Y (EY),[194] 204 205 tetrasulfonated zinc phthalocyanine  $(ZnPcS_4^{-}), [146]$  and Rose Bengal [237] have been successfully used to 206 catalyse aqueous dispersion RAFT PISA (Table-1).
- 207

## 208 **3. Hydrophilic steric stabilizer block.**

A wide variety of functional hydrophilic polymers has been used to conduct aqueous dispersion RAFT PISA and ultimately to stabilize the formulated nano-objects. **Fig. 4** summarizes the chemical structures of the steric stabilizers reported to date in the literature. The most commonly are: i) neutral such as poly(glycerol methacrylate) (PGMA),[114, 124] poly(ethylene oxide) (PEO),[131, 132, 135, 147, 148] and poly(2-hydroxypropyl methacrylamide) (PHPMAc),[154, 197, 227] ii) cationic such as poly(2dimethylamino)ethyl methacrylate)) (PQDMA),[43, 168] iii) anionic such as poly(3-sulfopropyl potassium methacrylate) (PKSPMA),[43, 171] and iv) zwitterionics such as poly(2-methacryloyloxyethyl

- 216 phosphorylcholine) (PMPC),[130, 164, 165] poly(potassium 3-sulfopropyl methacrylate) cysteine)
- 217 (PcysMA),[166] and poly(sulfobetaine methacrylate) (PSBMA).[167] Mixture of homopolymers[130, 166,
- 218 173-177, 231] or copolymers[168, 171, 180, 182, 206, 207, 217] were also used.



220 Fig. 4. Steric stabilizer blocks used for aqueous dispersion PISA.

Although the scope of aqueous dispersion PISA is widely focused on steric stabilizer blocks based on 221 222 synthetic polymers, some reports describe the successful preparation of nanostructures from natural polymers such as dextran [155-159] and  $\beta$ -cyclodextrin, [181] bovine serum albumin (BSA) [161] as well as 223 polylysine as bio-inspired peptide.[170] Recently, we demonstrated for the first time the suitability of the 224 aqueous dispersion PISA process to prepare polysaccharides-based nano-objects of advanced 225 226 morphologies, in contrast to solvent-switch methods that led mainly to spheres. [29, 32] To do so, graft copolymers composed of dextran as a hydrophilic backbone bearing poly(2-hydroxypropyl methacrylate) 227 grafts, were prepared via PI-RAFT PISA at room temperature. [155, 157-159] Our investigations revealed 228 that this model system can form well-defined vesicles of modulable membrane thickness over a wide range 229 of solid content.[158] 230

Two strategies have been reported in the literature to prepare reactive steric stabilizers used in PISA process. The first one, and probably the most used, involves TI-RAFT polymerization of vinyl derivatives monomers to produce the corresponding macromolecular chain transfer agent (macroCTA) such as PGMA and PHPMAc. The second is based on the functionalization of a pre-synthesized polymers by CTA. For instance, CTA group is introduced on PEO by esterification of its terminal hydroxyl function, while for dextran, BSA and polylysine, the CTA group is introduced through an amidation coupling.

237

#### 238 4. Water-soluble monomers

#### 239 **4.1. Overview**

The essential criterion to well conduct an aqueous dispersion PISA is the initial water-solubility of the 240 monomer used that leads to a water-insoluble polymer. Thus, a limited number of monomers respecting 241 242 this condition was known in 2014 when Warren and Armes reported the first review on aqueous dispersion RAFT PISA.[71] At this time, the literature examples included N-isopropylacrylamide (NIPAM),[42] 243 244 N,N'-diethylacrylamide (DEAA),[41, 217, 218] 2-methoxyethyl acrylate (MEA),[210] 2-hydroxypropyl methacrylate (HPMA), [43, 114-117, 121, 123, 131, 165, 168, 171, 177, 180] and di(ethylene glycol) 245 methyl ether methacrylate (DEGMA).[223] Moreover, regardless of the polymerization methods used to 246 grow the core-forming block, only HPMA enables a direct access to advanced nano-objects morphologies 247 (worms and vesicles).[114-117, 121, 123, 131, 165, 177, 180] During the last ten years, great efforts have 248 been made to overcome these limitations either by playing on the experimental conditions or by 249 developing new monomers. The monomers currently reported for aqueous dispersion RAFT PISA are 250 summarized in the Fig. 5A and the experimental conditions used for their polymerization are summarized 251 252 in the Table-1. In the following paragraphs, recent progress made for previous monomers (NIPAM, MEA, DEEA and DEGMA) as well as newly developed ones for aqueous dispersion RAFT PISA will be 253 described. A particular attention will be given to HPMA, the monomer of reference and diacetone 254 255 acrylamide (DAAM) the emerging water-soluble monomer.



258 Fig. 5. Water-soluble monomers and (B) crosslinkers used in aqueous dispersion RAFT PISA.

#### **4.2. Water-soluble (meth)acrylates**

N,N'-dimethylaminoethyl methacrylate (DMAEMA) is a water soluble monomer forming a cationic 260 polymer with thermo- and pH-responsive behavior.[257] DMAEMA is probably the first methacrylate 261 monomer used in aqueous dispersion RAFT PISA. Yan and Tao[219] reported the chain extension of PEO 262 macro-RAFT agent with DMAEMA at 65°C, above the lower critical solubility temperature (LCST) of 263 264 poly(N,N'-dimethylaminoethyl methacrylate) - PDMAEMA,[258] in the presence of a diacrylamide symmetric crosslinker (N,N'-methylene bis-(acrylamide), MBA). However, only spherical nanogel (NG) 265 has been formed when cooling down the formulation to RT, probably due to the action of MBA that 266 267 prevents chain reorganization into higher order morphology. By introducing a permanent cationic charge in 2-(methacryloyloxy)ethyltrimethyl-ammonium hexafluorophosphate 268 in **DMAEMA** resulting  $([META^+], [PF_6^-])$  An and coworkers [220] have modulated the water-solubility of  $([META^+], [PF_6^-])$  and 269 the water-insolubility of its corresponding homopolymer. Thus, in an elegant way, the authors succeeded 270 in producing stable spheres, worms and high-density network structure by chain extending poly(ethylene 271 glycol)methyl ether methacrylate (PEGMA) macro-RAFT agent with ( $[META^+], [PF_6^-]$ ) in water and 272 without adding any crosslinker. By taking advantage from the facile self-assembly of HPMA-based 273 diblock amphiphilic copolymers into advanced morphologies, as it will be discussed below, several 274 research groups have been interested in studying the DMAEMA/HPMA binary system to drive the 275 formation of higher-order morphologies.[137, 232-234] In a first study, Tan and coworkers[232] observed 276 277 various morphologies, such as worms and vesicles, by chain extension of poly(poly(ethylene glycol)) methyl ether methacrylate), PPEGMA, macro-CTA with HPMA/DMAEMA co-monomers at RT using PI-278 279 RAFT polymerization. In a second study, Guragain and Perez-Mercader[233] extended PEO-b-PDMAEMA diblock copolymer spheres with HPMA to produce worms that can undergo a morphological 280 transition to vesicles by switching the pH of the medium. 281

2-Methoxyethyl acrylate (MEA) is a highly water-soluble monomer forming a water-insoluble poly(2-282 methoxyethyl acrylate) - PMEA. Such a feature has appealed the attention of researchers for developing 283 novel system for aqueous dispersion PISA.<sup>[209-211]</sup> An and co-workers<sup>[210]</sup> reported formulation of 284 285 crosslinked spherical nanoparticles (NPs) by chain extending PPEGMA with MEA, in the presence of a 286 poly(ethylene glycol) diacrylate (PEGDA) crosslinker at 30°C by using a low temperature initiation system (RI-RAFT polymerization in the presence of KPS/NaAs). As we previously mentioned, crosslinking the 287 core of nano-objects during the PISA using a symmetric crosslinker leads to a frozen spherical 288 289 morphology. [223, 238] In this context, Maeda and coworkers [211] have successfully observed vesicles at high solid content (20% to 30%), while worms were observed only at 15% w/w, by performing PISA of 290 291 MEA from PEO macro-CTA at 70°C without adding any crosslinker. To increase the experimental 292 window of worms, and more particularly fiber morphology, Rieger and coworkers [208] used an endfunctionalized poly(N,N- dimethylacrylamide) (PDMA) macro-CTA with a hydrogen- bonded bis-urea to
 drive PISA of MEA in water towards fiber.

Hydroxypropyl methacrylate (HPMA) is a water-miscible [259] monomer, up to 13% at 25°C, forming a 295 296 water-insoluble homopolymer (poly(hydroxypropyl methacrylate) - PHPMA) from an average degree of polymerization of ~15. PHPMA exhibits glass transition temperature ( $T_{o}$ ) of 95°C in bulk which decreases 297 to 47°C by plasticization in water (values determined for PHPMA of an average molar mass of 30000 298 g/mol).[114] HPMA is a commercially available monomer prepared by reacting propylene oxide with 299 methacrylic acid resulting in HPMA (75 mol%) and 25 mol% of its isomer (2-hydroxyisopropyl 300 methacrylate - HIPMA) due to the poor regioselectivity of the epoxide ring opening reaction.[259] To the 301 302 best of our knowledge, the contribution of each isomer in the self-assembly has not yet been studied. Employed for the first time in 2010,[43] HPMA has become arguably the most studied monomer in the 303 304 context of aqueous dispersion PISA. As summarized in Table 1, more than 60 publications have been reported to date on the formulation of nano-objects composed of PHPMA as core-forming block. The 305 306 particularity of HPMA that makes it a highly attractive monomer for PISA comes probably from its 307 capacity to form large set of morphologies including common morphologies (micelle or sphere (S) wormlike micelles (WLM) and unilamellar vesicle (ULV)) as well as complex ones (framboidal vesicle (FV), 308 309 jellyfish (J), tubular vesicle (TV), multi-compartment vesicle (MCV) and donut-like vesicle (DV)). Fig. 6 summarizes some images of such various morphologies. In the following paragraphs, the discussion will 310 be limited to a few key works on HPMA in the context of aqueous dispersion RAFT PISA. 311

312 HPMA was successfully polymerized from a variety of functional steric stabilizers in aqueous dispersion RAFT PISA to prepare nano-objects with variable surface chemistry. Nano-objects with neutral stabilizers, 313 including PGMA[116, 121, 123, 125, 129] and PEO,[131, 134, 135, 140, 260] were designed for 314 biomedical applications, [116, 131, 134, 135, 260] for stabilization of oil-in-water pickering 315 emulsions, [121, 123, 125] and to improve the resistance against surfactants. [129, 140] On the other hand, 316 polyelectrolyte-based stabilizers block including cationic [168-170, 175] and anionic [166, 171, 172] ones 317 were used to enhance the electrostatically stabilization of nanoparticles[166, 169-172] and for 318 antimicrobial activity.[170] However, only spheres are observed when using polyectrolytes, due to the 319 strong electrostatic repulsion between the charged chains forming the steric stabilizer. Various strategies 320 321 have been used to reduce the charges around such nano-objects and thus to reach higher morphologies, such as the use of statistical copolymer-based stabilizers by copolymerization of the ionic monomer with a 322 nonionic comonomer,[168] or the use of binary mixtures of polyelectrolytic stabilizer and nonionic 323 324 one.[166, 174, 176, 231] HPMA has also been used as a model monomer to study the mechanism of PISA in aqueous dispersion from natural stabilizers such as BSA [161] and dextran.[155-159] Note that aqueous 325 dispersion RAFT PISA was beneficial in achieving advanced morphology (including vesicles) using 326 dextran as polysaccharide stabilizer[155-159] in contrast to nanoprecipitation, [157] which only led to 327

328	spherical nanoparticles. On the other hand, HPMA has been used to study the impact of the ACPs
329	architecture on the resulted nano-objects morphology. Comparing with linear ACPs, it was demonstrated
330	that branched ACPs[139] did not facilitate the formation of higher-order morphologies, while the
331	bottlebrush[153] or graft[158, 159] ACPs can promote the formation of higher-order morphologies when
332	targeting similar degrees of polymerization. Further studies are needed to generalize these observations.
333	



Fig. 6 A selection of some TEM images of PHPMA core-forming block nano-objects prepared by aqueous
dispersion RAFT PISA. Images reproduced and adapted with permission: BW (ref [114]. Copyright 2011,
American Chemical Society), DV (ref [126]. Copyright 2020, American Chemical Society), EULV (ref [160].
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339 [117]. Copyright 2012, American Chemical Society), GULV (ref [132]. Copyright 2017, Royal Society of Chemistry), J (ref [114]. Copyright 2011, American Chemical Society), L (ref [138]. Copyright 2019, Royal 340 Society of Chemistry), LR (ref [165]. Copyright 2011, Royal Society of Chemistry), MLV (ref [131]. Copyright 341 2014, American Chemical Society), MCV (ref [159]. Copyright 2022, American Chemical Society), OM (ref [155]. 342 343 Copyright 2018, Royal Society of Chemistry), R (ref [134]. Copyright 2019, American Chemical Society), S (ref 344 [114]. Copyright 2011, American Chemical Society), SNP (ref [156]), Sp (ref [178]. Copyright 2020, American Chemical Society), TV (ref [126]. Copyright 2020, American Chemical Society), ULV (ref [158]. Copyright 2021, 345 American Chemical Society), and WLM (ref [116]. Copyright 2012, American Chemical Society). 346

347

PGMA-b-PHPMA was used as model to investigate the polymerization behavior in aqueous dispersion TI-348 RAFT PISA (Fig. 7A).[114] A non-linear evolution of the semilogarithmic plot, displaying mainly two 349 350 distinct regimes was observed for the first time. To sum up, the reaction medium remained homogeneous in the first regime, revealing a modest growth of PHPMA that resulted in the formation of water-soluble 351 352 diblock copolymers. The medium became turbid at a specific time called onset of nucleation, indicating 353 that PHPMA block has reached its water-soluble critical size. At this time, monodisperse micelles were observed by DLS and TEM. In the second regime, the dispersion became progressively heterogeneous and 354 355 opaque, involving an increase in the slope of the semilogarithmic plot. This unusual kinetic profile is 356 attributed to the diffusion of the unreacted HPMA monomers into nascent micelles, resulting in an increase in the local monomer concentration and thus an enhancement of the polymerization rate. TEM images of 357 aliquots extracted in the second regime enabled authors to establish a remarkable mechanistic of the *in situ* 358 359 structural evolution from micellar nucleation to the final vesicular morphology. These images reveal that "jellyfish-like" structure is the key intermediate in the transition WLM to ULV as shown in **Fig. 7B**. 360

361 Morphology transition beyond the vesicular one has been studied by many research groups. By using *in* situ SAXS studies, Armes and coll.[119] demonstrated that the overall vesicle diameter remained constant 362 while the vesicle membranes continue to thicken, suggesting an 'inward growth' mechanism of 363 polymersomes during the final stages of the HPMA polymerization from PGMA at 70°C (Fig. 7C). Using 364 the same PISA system but at room temperature under 405nm irradiation, Tan and coll.[126] observed that 365 polymersomes fused to form tubular and donut-like polymersomes (Fig. 7D). By polymerizing HPMA 366 from dextran backbone via PI-RAFT PISA, our group[159] observed the formation of large compound 367 nano-objects (LCNs) resulting from a fusion of multicompartment vesicles as intermediate structures, 368 369 when targeting long PHPMA grafts degrees of polymerization (Fig. 7E). We attributed the formation of 370 this unprecedented intermediate structure to a local phase separation of dextran and PHMPA/HPMA, 371 which leads to aqueous phase domains.



373

Fig. 7 (A) HPMA polymerization kinetics obtained for the targeted  $PGMA_{47}$ -b-PHPMA<sub>200</sub> diblock copolymer 374 nano-objects prepared via aqueous dispersion TI-RAFT PISA at 70 °C and 10% w/w solids. The inset shows a 375 semilogarithmic plot. Reproduced with permission from ref [114] Copyright 2011, American Chemical Society. (B) 376 Suggested mechanism for the WLM-to-V transformation during the synthesis of PGMA<sub>47</sub>-b-PHPMA<sub>200</sub> by aqueous 377 dispersion TI-RAFT PISA at 70 °C. Reproduced with permission from ref [114] Copyright 2011, American 378 Chemical Society (C) Schematic representation of the 'vesicle inward growth' mechanism when targeting PGMA<sub>45</sub>-379 b-PHPMA<sub>200</sub> vesicles via aqueous dispersion TI-RAFT PISA at 70 °C. Reproduced with permission from ref [119] 380 Copyright 2020, Royal Society of Chemistry. (D) Schematic Illustration for the growth of polymersomes during the 381 synthesis of PGMA<sub>62</sub>-b-PHPMA<sub>1000</sub> by aqueous dispersion PI-RAFT PISA at 25 °C and 20% w/w. Reproduced with 382 383 permission from ref [126] Copyright 2020, American Chemical Society (E) CryEM images of HPMA and schematic Illustration for the transformation of MCV intermediate structure to LCN morphology during the 384 385 synthesis of Dex-g<sup>12</sup>-PHPMA<sub>500</sub> by aqueous dispersion PI-RAFT PISA at 25 °C and 10% w/w. Reproduced with permission from ref [159] Copyright 2022, American Chemical Society. 386

387

## 388 4.3. Water-soluble acrylamides

389 *N-isopropylacrylamide (NIPAm)* is a water-soluble monomer forming a thermoresponsive polymer, 390 poly(N-isopropyl-acrylamide) (PNIPAm), that undergoes a coil-to-globule transition in water at 391 temperature higher than its lower critical solution temperature (LCST) ( $\approx$ 32 °C).[261] Inspired from the 392 work of Jo and coll.[262] that exploited the particular thermosensitive characteristics of PNIPAm to 393 produce crosslinked nanogels via aqueous dispersion ATRP mediated PISA, Stucky and coll.[42] were the first preparing PNIPAm-based nanogels (NG) by aqueous dispersion RAFT PISA. In this pioneering work, 394 NIPAm was growth in water from a preformed PDMA macromolecular RAFT agent at 70°C. 2 mol% of 395 MBA crosslinker (relative to NIPAm) was voluntary added at the beginning of the polymerization to 396 397 progressively crosslink the core of nano-objects and avoid the dissolution of nano-objects after cooling down to room temperature (RT). NGs have been also observed by others when NIPAm was copolymerized 398 with MBA even by modifying both the chemical nature of the hydrophilic steric stabilizer 399 (poly(oligo(ethylene glycol) methyl ether acrylate), PPEGA, instead of PDMA)[205] and the architecture 400 of copolymer (tribloc instead of dibloc).[203] Different strategies were employed to prepare higher-order 401 morphologies of PNIPAm-based block copolymers nanoparticles. Sumerlin and coll. [206] have selectively 402 403 crosslinked the corona of the nanoparticles instead of the core. To do this, authors prepared nano-objects at 404 70°C by polymerizing NIPAm from a water-soluble copolymer composed of acrylic acid (AA) and DMA, followed by an immediate crosslinking of the corona at 70°C prior cooling via the reaction of the 405 carboxylic acid groups of AA with ethylene diamine (EDA) in the presence of a carbodiimide. Tan and 406 407 coll.[263] have successfully prepared advanced nano-objects morphology by copolymerizing NIPAm and 408 a small amount of asymmetric allyl acrylamide (ALAM), containing two vinyl groups of different reactivities (Fig. 5B), from PDMA under purple irradiation (405nm) at 70°C. Unlike MBA, which reacts 409 and crosslinks the nanoparticles-core from the biggening of the polymerization, ALAM delays the 410 crosslinking step at the end of the polymerization, thanks to the difference of reactivity between the 411 412 acrylamide function that reacts at the early stage of polymerization and the allyl function that reacts at the late polymerization stage. [186] We suspect that the development of this second strategies can improve the 413 attractivity of NIPAm for aqueous dispersion PISA. 414

N,N-diethylacrylamide (DEAA) is another water-soluble monomer forming thermosensitive poly(N,N 415 diethyl-acrylamide) homopolymer (PDEAA) exhibiting LCST at 32 °C.[264] DEAA is one of the first 416 monomer carried out in aqueous dispersion PISA. In 2007, Charleux and coll.[41] reported preparation of 417 crosslinked core PDEAA-based NG directly in water. In this seminal work, a hydrophilic poly(sodium 418 acrylate) alkoxyamine macroinitiator was chain extended by DEAA via NMP. Few years later, the same 419 team carried TI-RAFT polymerization of DEAA in the presence of MBA, at 60°C-70°C in water from 420 PEO-b-PDAM copolymer as hydrophilic steric stabilizer.[217, 218] Only spherical NGs have been 421 422 observed as judged by transmission electron microscopy images. Ten years later, Boyer and coll.[264] 423 used PDEAA NGs as model to prepare *in situ* doxorubicin (DOX)-loaded nanoparticles for drug delivery 424 by aqueous dispersion PI-RAFT PISA under mild conditions (37°C). In addition to being an efficient 425 antitumor drug, the authors discover that DOX plays an unexpected role of photocatalyst in the presence of triethylamine, leading to an acceleration of the rate of polymerization. Despite the potential of DEAA, this 426

427 monomer is still underexploited in aqueous dispersion PISA, and that more effort is needed to find the428 adequate experimental conditions allowing access to advanced morphologies.

Diacetone acrylamide (DAAM) is an emerging commercially available water-soluble monomer that forms 429 430 a water-insoluble polymer (PDAAM) from an average degree of polymerization of 50.[265] During the last seven years, DAAM has grasped attention of scientists due to its potential to produce polymeric nano-431 objects of advanced morphology in aqueous dispersion PISA. DAAM was used for the first time in 2015, 432 simultaneously but separately, by An's group[188] and Cai's group[197] in aqueous dispersion RAFT 433 PISA to produce vesicles and micelles, respectively. DAAM is undoubtedly the second most studied 434 monomer in the context of aqueous dispersion PISA. DAAM was chain extended by RAFT polymerization 435 using light[194, 197] and temperature[188] as stimulus from various hydrophilic macro-RAFT agents 436 including neutrals (PDMA, [188] PHPMAc, [197] PEO[196] and poly(N-vinylpyrrolidone) (PVP)[201]) 437 and cationic ones (Poly((vinylbenzyl) trimethylammonium chloride) (PVBTMAC)[200] and poly(N-(2-438 hydroxypropyl) methacrylamide)-block-poly(histamine methacrylamide hydrochloride) (PHPMAc-b-439 440 PHisMA)[202] to produce nano-objects of various advanced morphology and surface functionality. To the 441 best of our knowledge, DAAM is the only monomer that has been chain extended in aqueous dispersion 442 PISA from PVP, a hydrophilic steric stabilizer prepared by the polymerization of N-vinylpyrrolidone (VP), 443 a less activated monomer.[201] To do so, VP was polymerized using xanthate chain transfer (CTA-34, Fig. 444 2) via PI-RAFT. Subsequently, PVP was chain extended with DAAM in water at 60°C via TI-RAFT polymerization. Spheres and vesicles were observed in this case, although the uncontrolled RAFT 445 polymerization behavior during PISA due to the undesirable side reactions that PVP can undergoes in 446 aqueous media. Crosslinking approaches have been exploited (Fig. 5B) both in-situ [186, 187, 189] and 447 post-polymerization[183, 193], to increase the stability of PDAAM-core forming nano-objects. An. and 448 449 coll.[187] have reported another original approach to improve the colloidal stability of vesicles against harsh conditions such as solvent dilution, freeze-drying and surfactants competitors. To do so, the 450 crosslinked PDAM-b-PDAAM diblock copolymer vesicles were served as a platform from which third 451 hydrophilic blocks, neutral PDAM, anionic poly(2-acrylamido-2-methyl-1-propanesulfonic acid sodium 452 salt), or cationic poly(3-acrylamidopropyl trimethylammonium chloride), were initiated to generate 453 454 crosslinked triblock copolymer vesicles without any morphological transitions whatever the size of the third hydrophilic block. 455

Bilayer-based morphologies (lamellae or vesicle) are relatively easily obtained with DAAM *via* PISA, which is a considerable advantage compared to other monomers. However, anisotropic morphologies (worms) are rarely observed with DAAM. In this context, Cai and coll.[199] reported the formation of pore-switchable nanotubes at high solid content (>35%) derived by hydrogen bound interactions. Armes and coll.[183] demonstrated that the chain extension of PDMA with DAAM can produce pure worms in an extremely narrow window in the phase diagram composed of large domains of both vesicles and sphere/worm mixtures. In this case, the PDAAM-core forming nano-objects was stabilized by post-

polymerization using adipic acid dihydrazide (ADH), which reacts selectively with the pendent ketone 463 464 groups on the hydrophobic PDAAM chains to form hydrazone moieties. Simultaneously, Sumerlin and coworkers.[193] demonstrated that tuning the hydrophilicity/hydrophobicity of PDAAM-core forming 465 block nano-objects during the PISA process, through the copolymerization of DAAM with a DAM 466 hydrophilic monomer, favored the formation of worms of controlled length. Due to the thermoresponsive 467 behavior of PDAAM-based amphiphilic copolymers, the nano-objects were crosslinked immediately 468 before cooling by reacting a difunctional alkoxyamine (O,O'-1,3-propanediyl bishydroxyamino 469 hydrochlorid) with ketone functional groups of the DAAM units to produce hydrolytically stable oxime 470 linkages. This allowed to lock the morphology prepared during PISA at high temperature and to prevent 471 the system from any morphology transition. Recently, it was found that the morphology of nano-objects 472 473 resulted from the PISA of DAAM can be impacted by the experimental conditions [192, 198] and the 474 copolymer architecture.[184, 190, 196] Cai and coll.[198] studied the aqueous dispersion PI-RAFT PISA of DAAM monomer using PHPMAc macro-CTA and SPTP initiator in water at 25 - 60 °C. The authors 475 observed lamellae (L) when conducting PISA at RT, while WLM or ULV were observed at higher 476 temperatures. Moreover, reversible thermoresponsive ULV-to-L-to-ULV and ULV-to-V-to-ULV 477 transitions were observed upon a cooling-heating cycle. By taking advantage of the difference in 478 479 polymerization rates between TI-PISA and PI-PISA, Tan and coll.[192] demonstrated that the polymerization kinetics has a strong impact on the self-assembled structures formed in PISA of DAAM. 480 By performing PISA at the same temperature (70 $^{\circ}$ C), they found that the relatively low polymerization rate 481 482 of TI-RAFT PISA provides sufficient time for block copolymers to relax and reorganize from spheres to 483 worms and vesicles, while kinetically trapped spheres were formed by PI-RAFT PISA due to its ultrafast 484 rate.

485 Novel synthetic acrylamides. The limited number of monomers that meet requirements of aqueous dispersion PISA has prompted many research groups to develop new synthetic monomers. In this context, 486 Pascual and coll.[266] have explored for the first time the well-known synthetic water-soluble monomer N-487 acryloyl glycinamide (NAGA) that forms poly(N-acryloyl glycinamide), PNAGA, a homopolymer 488 exhibiting an upper critical solution temperature (UCST) behavior. Thermosensitive NGs was prepared by 489 490 the chain extension of PPEGA hydrophilic stabilizer with NAGA at 3°C via PI-RAFT PISA under UV irradiation ( $\lambda = 365$  nm) in the presence of PP-OH photoinitiator. NGs were crosslinked *in situ* by MBA 491 crosslinker during polymerization at 3°C to prevent NGs from dissolution at high temperature. 492 493 Simultaneously, Rieger and coll.[239] reported another UCST thermosensitive nano-objects prepared by 494 chain extension of PDMA via RAFT copolymerization of acrylamide (AAm) and acrylonitrile (AN) in 495 water at 45°C, without adding a crosslinker. WLM and S that underwent either reversible 496 aggregation/dissociation (S to individual chains) or reversible morphological transition (WLM to S) have been observed upon heating, depending on the length and the molar fraction of AN in the poly(AAm-co-497 498 AN) core forming block. Inspired by this work, the same team has recently developed a novel water-

soluble monomer N-cyanomethylacrylamide (CMAm)[267] by merging both AN and AAm in a single 499 monomer. Various nano-objects including S, WLM and ULV were successfully prepared via chain 500 extension of PDMA with CMAm in water at 45°C.[268] Semasarilar and coll.[224] have developed a 501 novel tripeptide-containing methacrylamide (MAm-FGD) based on the phenylalanine-glycine-aspartic acid 502 sequence. Chain extension of PGMA used as macromolecular chain agent with MAm-FGD in water at 503 504 70°C resulted in the formation of unprecedented morphology of large dendritic and bow-tie shaped fibrous spherulitic (Sp) structures. *N-acryloylthiomorpholine (NAT)*[269] is another novel hydrophilic monomer 505 forming hydrophobic homopolymer even at very short length (X=25) recently used by Brendel and 506 coll.[221] A wide range of nano-objects morphologies behavior has been formed by chain extension of 507 poly(N-acryloylmorpholine) (PNAM), a hydrophilic macro-CTA prepared by polymerizing N-508 acryloylmorpholine (NAM) that exhibits a chemical structure similar to NAT. Moreover, the nano-objects 509 morphology can be tuned from S to ULV by increasing the polymerization temperature used during the 510 PISA process. Cai and coworkers. [227, 270] reported successful aqueous dispersion RAFT PISA of 511 cationic core-forming monomers, 2-aminoethylacrylamide hydrochloride (AEAM)[270] and cystamine 512 methacrylamide hydrochloride (CysMA),[227] using a nonionic PHPMAc macro-CTA in the presence of 513 an anionic poly(2-acrylamido-2- methylpropanesulfonic acid) (PAMPS) which induces in situ polyion 514 complexation and morphology transition depending on the size of core-forming block. 515

#### 516 Conclusions

517 In summary, aqueous dispersion RAFT PISA process has reached some maturity in terms of production of 518 highly ordered nano-objects. Activation of the polymerization in this process was demonstrated using 519 different stimuli and in the presence of various steric stabilizers. Extensive investigations have been carried out using traditional water-soluble monomers forming water-insoluble polymers, and novel monomer for 520 advanced polymer properties. Diagrams of morphology are now known for some specific model systems. 521 It was established that the morphology could be modulated, for some system, by varying the core-forming 522 block size or the solid content. Aqueous dispersion RAFT PISA process remains currently empirical, and it 523 is important now to carry out deeper studies to rationalize the large data generated during the past decade 524 or so. Many fundamental questions remain unanswered: is it possible to predict the morphology of nano-525 objects according to the chemical nature of the hydrophilic stabilizing block and/or to the monomers used 526 ? How can we modulate the morphology of nano-objects by playing on the architecture of the final 527 copolymer? 528

#### 529 Conflicts of interest

530 There are no conflicts to declare.

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#### 534 **References**

- 535 [1] H. Kawaguchi, Functional polymer microspheres, Prog. Polym. Sci. 25(8) (2000) 1171-1210.
- 536 [2] D.E. Discher, A. Eisenberg, Polymer Vesicles, Science 297(5583) (2002) 967-973.
- 537 [3] A. Blanazs, S.P. Armes, A.J. Ryan, Self-Assembled Block Copolymer Aggregates: From Micelles to Vesicles and their Biological
- 538 Applications, Macromol. Rapid Commun. 30(4-5) (2009) 267-277.
- [4] U. Tritschler, S. Pearce, J. Gwyther, G.R. Whittell, I. Manners, 50th Anniversary Perspective: Functional Nanoparticles from the Solution
   Self-Assembly of Block Copolymers, Macromolecules 50(9) (2017) 3439-3463.
- 541 [5] M.E. Davis, Z. Chen, D.M. Shin, Nanoparticle therapeutics: an emerging treatment modality for cancer, Nat. Rev. Drug. Discov 7(9) (2008) 771-782.
- [6] J.J. Shi, P.W. Kantoff, R. Wooster, O.C. Farokhzad, Cancer nanomedicine: progress, challenges and opportunities, Nat. Rev. Cancer 17(1)
  (2017) 20-37.
- [7] P. Tanner, P. Baumann, R. Enea, O. Onaca, C. Palivan, W. Meier, Polymeric Vesicles: From Drug Carriers to Nanoreactors and Artificial
  Organelles, Accounts Chem. Res. 44(10) (2011) 1039-1049.
- [8] H.M. Shewan, J.R. Stokes, Review of techniques to manufacture micro-hydrogel particles for the food industry and their applications, J.
  Food Eng. 119(4) (2013) 781-792.
- 549 [9] A. Plucinski, Z. Lyu, B. Schmidt, Polysaccharide nanoparticles: from fabrication to applications, J. Mater. Chem. B 9(35) (2021) 7030-550 7062.
- [10] S. Förster, M. Konrad, From self-organizing polymers to nano-and biomaterials, J. Mater. Chem. 13(11) (2003) 2671-2688.
- 552 [11] Y. Mai, A. Eisenberg, Self-assembly of block copolymers, Chem. Soc. Rev. 41(18) (2012) 5969-5985.
- 553 [12] G. Riess, Micellization of block copolymers, Prog. Polym. Sci. 28(7) (2003) 1107-1170.
- [13] J. Rodriguez-Hernandez, F. Checot, Y. Gnanou, S. Lecommandoux, Toward 'smart' nano-objects by self-assembly of block copolymers in
   solution, Prog. Polym. Sci. 30(7) (2005) 691-724.
- [14] F.H. Schacher, P.A. Rupar, I. Manners, Functional Block Copolymers: Nanostructured Materials with Emerging Applications, Angew.
   Chem.-Int. Edit. 51(32) (2012) 7898-7921.
- 558 [15] J.Z. Du, R.K. O'Reilly, Advances and challenges in smart and functional polymer vesicles, Soft Matter 5(19) (2009) 3544-3561.
- [16] E. Rideau, R. Dimova, P. Schwille, F.R. Wurm, K. Landfester, Liposomes and polymersomes: a comparative review towards cell
   mimicking, Chem. Soc. Rev. 47(23) (2018) 8572-8610.
- [17] M.J. Derry, L.A. Fielding, S.P. Armes, Polymerization-induced self-assembly of block copolymer nanoparticles via RAFT non-aqueous
   dispersion polymerization, Prog. Polym. Sci. 52 (2016) 1-18.
- 563 [18] N.A. Lynd, A.J. Meuler, M.A. Hillmyer, Polydispersity and block copolymer self-assembly, Prog. Polym. Sci. 33(9) (2008) 875-893.
- [19] Y.Q. Lu, J.P. Lin, L.Q. Wang, L.S. Zhang, C.H. Cai, Self-Assembly of Copolymer Micelles: Higher-Level Assembly for Constructing
   Hierarchical Structure, Chem. Rev. 120(9) (2020) 4111-4140.
- [20] C.K. Wong, X.L. Qiang, A.H.E. Muller, A.H. Groschel, Self-Assembly of block copolymers into internally ordered microparticles, Prog.
   Polym. Sci. 102 (2020) 22.
- 568 [21] A.D. Jenkins, R.G. Jones, G. Moad, Terminology for reversible-deactivation radical polymerization previously called "controlled" radical
   569 or "living" radical polymerization (IUPAC Recommendations 2010), Pure Appl. Chem. 82(2) (2010) 483-491.
- 570 [22] G. Moad, E. Rizzardo, S.H. Thang, Living Radical Polymerization by the RAFT Process A Second Update, Aust. J. Chem. 62(11) (2009)
  571 1402-1472.
- 572 [23] P.M. Kazmaier, K.A. Moffat, M.K. Georges, R.P.N. Veregin, G.K. Hamer, FREE-RADICAL POLYMERIZATION FOR NARROW-
- 573 POLYDISPERSITY RESINS SEMIEMPIRICAL MOLECULAR-ORBITAL CALCULATIONS AS A CRITERION FOR SELECTING STABLE FREE-RADICAL 574 REVERSIBLE TERMINATORS, Macromolecules 28(6) (1995) 1841-1846.
- [24] M.K. Georges, R.P.N. Veregin, G.K. Hamer, P.M. Kazmaier, BREATHING NEW LIFE INTO THE FREE-RADICAL POLYMERIZATION PROCESS,
   Macromolecular Symposia 88 (1994) 89-103.
- 577 [25] J.-S. Wang, K. Matyjaszewski, Controlled/"living" radical polymerization. atom transfer radical polymerization in the presence of 578 transition-metal complexes, J. Am. Chem. Soc. 117(20) (1995) 5614-5615.
- 579 [26] M. Kato, M. Kamigaito, M. Sawamoto, T. Higashimura, Polymerization of Methyl Methacrylate with the Carbon
- Tetrachloride/Dichlorotris- (triphenylphosphine)ruthenium(II)/Methylaluminum Bis(2,6-di-tert-butylphenoxide) Initiating System:
   Possibility of Living Radical Polymerization, Macromolecules 28(5) (1995) 1721-1723.
- [27] J. Chiefari, Y.K. Chong, F. Ercole, J. Krstina, J. Jeffery, T.P.T. Le, R.T.A. Mayadunne, G.F. Meijs, C.L. Moad, G. Moad, E. Rizzardo, S.H.
   Thang, Living free-radical polymerization by reversible addition-fragmentation chain transfer: The RAFT process, Macromolecules 31(16)
  - 584 (1998) 5559-5562.
- 585 [28] K. Matyjaszewski, Controlled radical polymerization, Curr. Opin. Solid State Mat. Sci. 1(6) (1996) 769-776.
- [29] K. Ferji, C. Nouvel, J. Babin, M.-H. Li, C. Gaillard, E. Nicol, C. Chassenieux, J.-L. Six, Polymersomes from Amphiphilic Glycopolymers
   Containing Polymeric Liquid Crystal Grafts, ACS Macro Lett. 4(10) (2015) 1119-1122.
- 588 [30] M. Nguyen, K. Ferji, S. Lecommandoux, C. Bonduelle, Amphiphilic Nucleobase-Containing Polypeptide Copolymers—Synthesis and Self-589 Assembly, Polymers 12(6) (2020) 1357.

- 590 [31] R.C. Hayward, D.J. Pochan, Tailored Assemblies of Block Copolymers in Solution: It Is All about the Process, Macromolecules 43(8) (2010) 3577-3584.
- 592 [32] J.L. Six, K. Ferji, Polymerization induced self-assembly: an opportunity toward the self-assembly of polysaccharide-containing
- 593 copolymers into high-order morphologies, Polym. Chem. 10(1) (2019) 45-53.
- 594 [33] A. Peyret, J.F. Trant, C.V. Bonduelle, K. Ferji, N. Jain, S. Lecommandoux, E.R. Gillies, Synthetic glycopolypeptides: synthesis and self-595 assembly of poly(gamma-benzyl-L-glutamate)-glycosylated dendron hybrids, Polym. Chem. 6(45) (2015) 7902-7912.
- 596 [34] M. El Founi, S.M.A. Soliman, R. Vanderesse, S. Acherar, E. Guedon, I. Chevalot, J. Babin, J.-L. Six, Light-sensitive dextran-covered PNBA
- 597 nanoparticles as triggered drug delivery systems: Formulation, characteristics and cytotoxicity, J. Colloid Interface Sci. 514 (2018) 289-298.
- 598 [35] K. Ferji, I. Hamouda, C. Chassenieux, B. Nadal, B. Dubertret, C. Gaillard, E. Nicol, Fast and effective quantum-dots encapsulation and 599 protection in PEO based photo-cross-linked micelles, J. Colloid Interface Sci. 476 (2016) 222-229.
- 600 [36] T.P.T. Dao, F. Fernandes, E. Ibarboure, K. Ferji, M. Prieto, O. Sandre, J.F. Le Meins, Modulation of phase separation at the micron scale 601 and nanoscale in giant polymer/lipid hybrid unilamellar vesicles (GHUVs), Soft Matter 13(3) (2017) 627-637.
- [37] T.P.T. Dao, A. Brulet, F. Fernandes, M. Er-Rafik, K. Ferji, R. Schweins, J.P. Chapel, F.M. Schmutz, M. Prieto, O. Sandre, J.F. Le Meins,
   Mixing Block Copolymers with Phospholipids at the Nanoscale: From Hybrid Polymer/Lipid Wormlike Micelles to Vesicles Presenting Lipid
   Nanodomains, Langmuir 33(7) (2017) 1705-1715.
- 605 [38] S.M.A. Soliman, M. El Founi, R. Vanderesse, S. Acherar, K. Ferji, J. Babin, J.-L. Six, Light-sensitive dextran-covered PNBA nanoparticles to 606 continuously or discontinuously improve the drug release, Colloids Surf. B 182 (2019) 110393.
- 607 [39] W.-M. Wan, C.-Y. Hong, C.-Y. Pan, One-pot synthesis of nanomaterials via RAFT polymerization induced self-assembly and morphology 608 transition, Chem. Commun. (39) (2009) 5883-5885.
- [40] W.-M. Wan, X.-L. Sun, C.-Y. Pan, Morphology Transition in RAFT Polymerization for Formation of Vesicular Morphologies in One Pot,
   Macromolecules 42(14) (2009) 4950-4952.
- 611 [41] G. Delaittre, M. Save, B. Charleux, Nitroxide-mediated aqueous dispersion polymerization: From water-soluble macroalkoxyamine to 612 thermosensitive nanogels, Macromol. Rapid Commun. 28(15) (2007) 1528-1533.
- [42] Z. An, Q. Shi, W. Tang, C.-K. Tsung, C.J. Hawker, G.D. Stucky, Facile RAFT Precipitation Polymerization for the Microwave-Assisted
   Synthesis of Well-Defined, Double Hydrophilic Block Copolymers and Nanostructured Hydrogels, J. Am. Chem. Soc. 129(46) (2007) 14493 14499.
- [43] Y. Li, S.P. Armes, RAFT Synthesis of Sterically Stabilized Methacrylic Nanolatexes and Vesicles by Aqueous Dispersion Polymerization,
   Angew. Chem. Int. Ed. 49(24) (2010) 4042-4046.
- 618 [44] B. Charleux, G. Delaittre, J. Rieger, F. D'Agosto, Polymerization-Induced Self-Assembly: From Soluble Macromolecules to Block
- 619 Copolymer Nano-Objects in One Step, Macromolecules 45(17) (2012) 6753-6765.
- [45] V.J. Cunningham, A.M. Alswieleh, K.L. Thompson, M. Williams, G.J. Leggett, S.P. Armes, O.M. Musa, Poly(glycerol monomethacrylate)–
   Poly(benzyl methacrylate) Diblock Copolymer Nanoparticles via RAFT Emulsion Polymerization: Synthesis, Characterization, and Interfacial
- 622 Activity, Macromolecules 47(16) (2014) 5613-5623.
- 623 [46] S.L. Canning, T.J. Neal, S.P. Armes, pH-Responsive Schizophrenic Diblock Copolymers Prepared by Polymerization-Induced Self-
- 624 Assembly, Macromolecules 50(16) (2017) 6108-6116.
- [47] J. Ma, H.M. Andriambololona, D. Quemener, M. Semsarilar, Membrane preparation by sequential spray deposition of polymer PISA
   nanoparticles, J. Membr. Sci. 548 (2018) 42-49.
- [48] C.J. Ferguson, R.J. Hughes, B.T.T. Pham, B.S. Hawkett, R.G. Gilbert, A.K. Serelis, C.H. Such, Effective ab Initio Emulsion Polymerization
   under RAFT Control, Macromolecules 35(25) (2002) 9243-9245.
- [49] M. Chenal, L. Bouteiller, J. Rieger, Ab initio RAFT emulsion polymerization of butyl acrylate mediated by poly(acrylic acid)
   trithiocarbonate, Polym. Chem. 4(3) (2013) 752-762.
- [50] S. Samanta, S.L. Banerjee, S.K. Ghosh, N.K. Singha, Smart Polyacrylate Emulsion Based on a New ABC-Type Triblock Copolymer via
   RAFT-Mediated Surfactant-Free Miniemulsion Polymerization: Its Multifunctional Properties, ACS Appl. Mater. Interfaces 11(47) (2019)
   44722-44734.
- 634 [51] C. Boussiron, M. Le Bechec, J. Sabalot, S. Lacombe, M. Save, Photoactive rose bengal-based latex via RAFT emulsion polymerization-635 induced self-assembly, Polym. Chem. 12(1) (2021) 134-147.
- [52] F.L. Hatton, J.R. Lovett, S.P. Armes, Synthesis of well-defined epoxy-functional spherical nanoparticles by RAFT aqueous emulsion
   polymerization, Polym. Chem. 8(33) (2017) 4856-4868.
- 638 [53] F.L. Hatton, A.M. Park, Y. Zhang, G.D. Fuchs, C.K. Ober, S.P. Armes, Aqueous one-pot synthesis of epoxy-functional diblock copolymer 639 worms from a single monomer: new anisotropic scaffolds for potential charge storage applications, Polym. Chem. 10(2) (2019) 194-200.
- 640 [54] X. Yan, V. La Padula, S. Favre-Bonte, J. Bernard, Heptyl mannose decorated glyconanoparticles with tunable morphologies through
- polymerization induced self-assembly. Synthesis, functionalization and interactions with type 1 piliated E. coli, Eur. Polym. J. 112 (2019)
   170-175.
- [55] X. Dai, L. Yu, Y. Zhang, L. Zhang, J. Tan, Polymerization-Induced Self-Assembly via RAFT-Mediated Emulsion Polymerization of
   Methacrylic Monomers, Macromolecules 52(19) (2019) 7468-7476.
- 645 [56] S. Varlas, J.C. Foster, P.G. Georgiou, R. Keogh, J.T. Husband, D.S. Williams, R.K. O'Reilly, Tuning the membrane permeability of
- 646 polymersome nanoreactors developed by aqueous emulsion polymerization-induced self-assembly, Nanoscale 11(26) (2019) 12643-12654.
- 647 [57] J. Rieger, G. Osterwinter, C. Bui, F. Stoffelbach, B. Charleux, Surfactant-Free Controlled/Living Radical Emulsion (Co)polymerization of
- n-Butyl Acrylate and Methyl Methacrylate via RAFT Using Amphiphilic Poly(ethylene oxide)-Based Trithiocarbonate Chain Transfer Agents,
   Macromolecules 42(15) (2009) 5518-5525.
- 650 [58] J. Engström, H. Asem, H. Brismar, Y. Zhang, M. Malkoch, E. Malmström, In Situ Encapsulation of Nile Red or Doxorubicin during RAFT-
- Mediated Emulsion Polymerization via Polymerization-Induced Self-Assembly for Biomedical Applications, Macromol. Chem. Phys. 221(5)
   (2020) 1900443.

- 653 [59] Q. Zhang, G. Yu, W.-J. Wang, H. Yuan, B.-G. Li, S. Zhu, Switchable Block Copolymer Surfactants for Preparation of Reversibly
- 654 Coagulatable and Redispersible Poly(methyl methacrylate) Latexes, Macromolecules 46(4) (2013) 1261-1267.
- [60] A.M. dos Santos, J. Pohn, M. Lansalot, F. D'Agosto, Combining Steric and Electrostatic Stabilization Using Hydrophilic MacroRAFT
   Agents in an Ab Initio Emulsion Polymerization of Styrene, Macromol. Rapid Commun. 28(12) (2007) 1325-1332.
- [61] D.E. Ganeva, E. Sprong, H. de Bruyn, G.G. Warr, C.H. Such, B.S. Hawkett, Particle Formation in ab Initio RAFT Mediated Emulsion
  Polymerization Systems, Macromolecules 40(17) (2007) 6181-6189.
- 659 [62] M. Manguian, M. Save, B. Charleux, Batch Emulsion Polymerization of Styrene Stabilized by a Hydrophilic Macro-RAFT Agent,
- 660 Macromol. Rapid Commun. 27(6) (2006) 399-404.
- [63] F. Lauterbach, V. Abetz, An eco-friendly pathway to thermosensitive micellar nanoobjects via photoRAFT PISA: the full guide to poly(N-acryloylpyrrolidin)-block-polystyrene diblock copolymers, Soft Matter 16(9) (2020) 2321-2331.
- [64] S.Y. Khor, N.P. Truong, J.F. Quinn, M.R. Whittaker, T.P. Davis, Polymerization-Induced Self-Assembly: The Effect of End Group and
- Initiator Concentration on Morphology of Nanoparticles Prepared via RAFT Aqueous Emulsion Polymerization, ACS Macro Lett. 6(9) (2017)
   1013-1019.
- 666 [65] W. Zhang, F. D'Agosto, O. Boyron, J. Rieger, B. Charleux, Toward a Better Understanding of the Parameters that Lead to the Formation
- of Nonspherical Polystyrene Particles via RAFT-Mediated One-Pot Aqueous Emulsion Polymerization, Macromolecules 45(10) (2012) 4075 4084.
- 669 [66] J. Bernard, M. Save, B. Arathoon, B. Charleux, Preparation of a xanthate-terminated dextran by click chemistry: Application to the
- 670 synthesis of polysaccharide-coated nanoparticles via surfactant-free ab initio emulsion polymerization of vinyl acetate, J. Polym. Sci. A:
   671 Polym. Chem. 46(8) (2008) 2845-2857.
- [672 [67] S. Binauld, L. Delafresnaye, B. Charleux, F. D'Agosto, M. Lansalot, Emulsion Polymerization of Vinyl Acetate in the Presence of Different
   Hydrophilic Polymers Obtained by RAFT/MADIX, Macromolecules 47(10) (2014) 3461-3472.
- [68] P. Galanopoulo, P.-Y. Dugas, M. Lansalot, F. D'Agosto, Poly(ethylene glycol)-b-poly(vinyl acetate) block copolymer particles with
   various morphologies via RAFT/MADIX aqueous emulsion PISA, Polym. Chem. 11(23) (2020) 3922-3930.
- [69] S.L. Canning, G.N. Smith, S.P. Armes, A Critical Appraisal of RAFT-Mediated Polymerization-Induced Self Assembly, Macromolecules
   49(6) (2016) 1985-2001.
- [70] X. Wang, Z.S. An, New Insights into RAFT Dispersion Polymerization-Induced Self-Assembly: From Monomer Library, Morphological
   Control, and Stability to Driving Forces, Macromol. Rapid Commun. 40(2) (2019) 14.
- [71] N.J. Warren, S.P. Armes, Polymerization-Induced Self-Assembly of Block Copolymer Nano-objects via RAFT Aqueous Dispersion
   Polymerization, J. Am. Chem. Soc. 136(29) (2014) 10174-10185.
- [72] J.-T. Sun, C.-Y. Hong, C.-Y. Pan, Recent advances in RAFT dispersion polymerization for preparation of block copolymer aggregates,
   Polym. Chem. 4(4) (2013) 873-881.
- 684 [73] A.B. Lowe, RAFT alcoholic dispersion polymerization with polymerization-induced self-assembly, Polymer 106 (2016) 161-181.
- [74] J. Rieger, Guidelines for the Synthesis of Block Copolymer Particles of Various Morphologies by RAFT Dispersion Polymerization,
   Macromol. Rapid Commun. 36(16) (2015) 1458-1471.
- [75] J. Huang, Y. Guo, S. Gu, G. Han, W. Duan, C. Gao, W. Zhang, Multicompartment block copolymer nanoparticles: recent advances and
   future perspectives, Polym. Chem. 10(25) (2019) 3426-3435.
- [76] Q. Xu, C. Tian, L. Zhang, Z. Cheng, X. Zhu, Photo-Controlled Polymerization-Induced Self-Assembly (Photo-PISA): A Novel Strategy Using
   In Situ Bromine-Iodine Transformation Living Radical Polymerization, Macromol. Rapid Commun. 40(2) (2019) 1800327.
- [77] E.R. Jones, M. Semsarilar, A. Blanazs, S.P. Armes, Efficient Synthesis of Amine-Functional Diblock Copolymer Nanoparticles via RAFT
   Dispersion Polymerization of Benzyl Methacrylate in Alcoholic Media, Macromolecules 45(12) (2012) 5091-5098.
- 693 [78] W. Zhao, G. Gody, S. Dong, P.B. Zetterlund, S. Perrier, Optimization of the RAFT polymerization conditions for the in situ formation of 694 nano-objects via dispersion polymerization in alcoholic medium, Polym. Chem. 5(24) (2014) 6990-7003.
- 695 [79] W.-J. Zhang, C.-Y. Hong, C.-Y. Pan, Artificially smart vesicles with superior structural stability: Fabrication, characterizations, and 696 transmembrane traffic, ACS Appl. Mater. Interfaces 9(17) (2017) 15086-15095.
- [80] C. Huang, J. Tan, Q. Xu, J. He, X. Li, D. Liu, L. Zhang, Adding a solvophilic comonomer to the polymerization-induced self-assembly of
  block copolymer and homopolymer: a cooperative strategy for preparing large compound vesicles, Rsc Advances 7(73) (2017) 4606946081.
- [81] G.H. Teo, P.B. Zetterlund, S.C. Thickett, Interfacial crosslinking of self-assembled triblock copolymer nanoparticles via alkoxysilane
   hydrolysis and condensation, J. Polym. Sci. A: Polym. Chem. 57(18) (2019) 1897-1907.
- [82] D. Das, D. Gerboth, A. Postma, S. Srinivasan, H. Kern, J. Chen, D.M. Ratner, P.S. Stayton, A.J. Convertine, Synthesis of zwitterionic,
   hydrophobic, and amphiphilic polymers via RAFT polymerization induced self-assembly (PISA) in acetic acid, Polym. Chem. 7(39) (2016)
- **704** 6133-6143.
- [83] B. Darmau, M.J. Rymaruk, N.J. Warren, R. Bening, S.P. Armes, RAFT dispersion polymerization of benzyl methacrylate in non-polar
   media using hydrogenated polybutadiene as a steric stabilizer block, Polym. Chem. 11(47) (2020) 7533-7541.
- 707 [84] L.A. Fielding, M.J. Derry, V. Ladmiral, J. Rosselgong, A.M. Rodrigues, L.P.D. Ratcliffe, S. Sugihara, S.P. Armes, RAFT dispersion
- polymerization in non-polar solvents: facile production of block copolymer spheres, worms and vesicles in n-alkanes, Chem. Sci. 4(5) (2013)
   2081-2087.
- [85] Y. Kang, A. Pitto-Barry, H. Willcock, W.-D. Quan, N. Kirby, A.M. Sanchez, R.K. O'Reilly, Exploiting nucleobase-containing materials –
   from monomers to complex morphologies using RAFT dispersion polymerization, Polym. Chem. 6(1) (2015) 106-117.
- 711 Information protogies using KAPT dispersion polymerization, Polymerization, Polymerization, 2013) 106-117.
   712 [86] L. Zhang, C. Song, J. Yu, D. Yang, M. Xie, One-pot synthesis of polymeric nanoparticle by ring-opening metathesis polymerization, J.
- 713 Polym. Sci. A: Polym. Chem. 48(22) (2010) 5231-5238.

- 714 [87] A. Xu, Q. Lu, Z. Huo, J. Ma, B. Geng, U. Azhar, L. Zhang, S. Zhang, Synthesis of fluorinated nanoparticles via RAFT dispersion
- polymerization-induced self-assembly using fluorinated macro-RAFT agents in supercritical carbon dioxide, RSC Adv. 7(81) (2017) 51612 51620.
- [88] Q. Zhang, S. Zhu, Ionic liquids: versatile media for preparation of vesicles from polymerization-induced self-assembly, ACS Macro Lett.
   4(7) (2015) 755-758.
- [89] C. Gao, H. Zhou, Y. Qu, W. Wang, H. Khan, W. Zhang, In situ synthesis of block copolymer nanoassemblies via polymerization-induced
   self-assembly in poly (ethylene glycol), Macromolecules 49(10) (2016) 3789-3798.
- 721 [90] K.M. Burridge, T.A. Wright, R.C. Page, D. Konkolewicz, Photochemistry for Well-Defined Polymers in Aqueous Media: From
- Fundamentals to Polymer Nanoparticles to Bioconjugates, Macromol Rapid Commun 39(12) (2018) e1800093.
- 723 [91] N. An, X. Chen, J. Yuan, Non-thermally initiated RAFT polymerization-induced self-assembly, Polym. Chem. 12(22) (2021) 3220-3232.
- [92] C. Liu, C.-Y. Hong, C.-Y. Pan, Polymerization techniques in polymerization-induced self-assembly (PISA), Polym. Chem. 11(22) (2020)
   3673-3689.
- [93] S.R. Mane, Trending methods employed for polymerization induced self-assembly, New Journal of Chemistry 44(17) (2020) 6690-6698.
   [94] M. Lages, N. Gil, P. Galanopoulo, J. Mougin, C. Lefay, Y. Guillaneuf, M. Lansalot, F. D'Agosto, J. Nicolas, Degradable Vinyl Copolymer
- Nanoparticles/Latexes by Aqueous Nitroxide-Mediated Polymerization-Induced Self-Assembly, Macromolecules 55(21) (2022) 9790-9801.
- 729 [95] C. Grazon, P. Salas-Ambrosio, S. Antoine, E. Ibarboure, O. Sandre, A.J. Clulow, B.J. Boyd, M.W. Grinstaff, S. Lecommandoux, C.
- Bonduelle, Aqueous ROPISA of α-amino acid N-carboxyanhydrides: polypeptide block secondary structure controls nanoparticle shape
   anisotropy, Polym. Chem. 12(43) (2021) 6242-6251.
- [96] S. Jimaja, S. Varlas, Y. Xie, J.C. Foster, D. Taton, A.P. Dove, R.K. O'Reilly, Nickel-Catalyzed Coordination Polymerization-Induced Self Assembly of Helical Poly(aryl isocyanide)s, ACS Macro Lett. 9(2) (2020) 226-232.
- [97] G. Moad, E. Rizzardo, A 20th anniversary perspective on the life of RAFT (RAFT coming of age), Polymer International 69(8) (2020) 658661.
- [98] F. D'Agosto, J. Rieger, M. Lansalot, RAFT-Mediated Polymerization-Induced Self-Assembly, Angew. Chem.-Int. Edit. 59 (2020) 8368–
   8392.
- 738 [99] S. Perrier, 50th Anniversary Perspective: RAFT Polymerization-A User Guide, Macromolecules 50(19) (2017) 7433-7447.
- [100] D. Liu, J. He, L. Zhang, J. Tan, 100th Anniversary of Macromolecular Science Viewpoint: Heterogenous Reversible Deactivation Radical
   Polymerization at Room Temperature. Recent Advances and Future Opportunities, ACS Macro Lett. 8(12) (2019) 1660-1669.
- 740 Forymentation at Norm remperature, Recent Advances and Future Opportunities, Acs Macro Lett. 8(12) (2019) 1000-1009.
   741 [101] S. Li, G. Han, W. Zhang, Photoregulated reversible addition–fragmentation chain transfer (RAFT) polymerization, Polym. Chem. 11(11)
- 742 (2020) 1830-1844.
- [102] N.J.W. Penfold, J. Yeow, C. Boyer, S.P. Armes, Emerging Trends in Polymerization-Induced Self-Assembly, ACS Macro Lett. 8(8) (2019)
   1029-1054.
- [103] J. Yeow, C. Boyer, Photoinitiated Polymerization-Induced Self-Assembly (Photo-PISA): New Insights and Opportunities, Adv. Sci. 4(7)
   (2017) 1700137.
- [104] S. Li, G. Han, W. Zhang, Cross-linking approaches for block copolymer nano-assemblies via RAFT-mediated polymerization-induced
   self-assembly, Polym. Chem. 11(29) (2020) 4681-4692.
- 749 [105] W.-J. Zhang, J. Kadirkhanov, C.-H. Wang, S.-G. Ding, C.-Y. Hong, F. Wang, Y.-Z. You, Polymerization-induced self-assembly for the 750 fabrication of polymeric nano-objects with enhanced structural stability by cross-linking, Polym. Chem. 11(22) (2020) 3654-3672.
- 750 Tablication of polyment nano-objects with enhanced structural stability by closs-initing, Polym. Chem. 11(22) (2020) 3034-3072.
   751 [106] D. Le, D. Keller, G. Delaittre, Reactive and Functional Nanoobjects by Polymerization-Induced Self-Assembly, Macromol. Rapid
   752 Commun. 40(2) (2019) 21.
- [107] J. Cao, Y. Tan, Y. Chen, L. Zhang, J. Tan, Expanding the Scope of Polymerization-Induced Self-Assembly: Recent Advances and New
   Horizons, Macromol. Rapid Commun. 42 (2021) 2100498.
- [108] B. Niu, Y. Chen, L. Zhang, J. Tan, Organic–inorganic hybrid nanomaterials prepared via polymerization-induced self-assembly: recent
   developments and future opportunities, Polym. Chem. 13(18) (2022) 2554-2569.
- [109] H. Phan, V. Taresco, J. Penelle, B. Couturaud, Polymerisation-induced self-assembly (PISA) as a straightforward formulation strategy
   for stimuli-responsive drug delivery systems and biomaterials: recent advances, Biomater. Sci. 9(1) (2021) 38-50.
- [110] S. Pearce, J. Perez-Mercader, PISA: construction of self-organized and self-assembled functional vesicular structures, Polym. Chem.
   12(1) (2021) 29-49.
- [111] W.J. Zhang, C.Y. Hong, C.Y. Pan, Polymerization-Induced Self-Assembly of Functionalized Block Copolymer Nanoparticles and Their
   Application in Drug Delivery, Macromol. Rapid Commun. 40(2) (2019) 1800279.
- [112] S.Y. Khor, J.F. Quinn, M.R. Whittaker, N.P. Truong, T.P. Davis, Controlling Nanomaterial Size and Shape for Biomedical Applications via
   Polymerization-Induced Self-Assembly, Macromol. Rapid Commun. 40(2) (2019) 1800438.
- [113] G. Cheng, J. Perez-Mercader, Polymerization-Induced Self-Assembly for Artificial Biology: Opportunities and Challenges, Macromol.
   Rapid Commun. 40(2) (2019) 1800513.
- [114] A. Blanazs, J. Madsen, G. Battaglia, A.J. Ryan, S.P. Armes, Mechanistic Insights for Block Copolymer Morphologies: How Do Worms
   Form Vesicles?, J. Am. Chem. Soc. 133(41) (2011) 16581-16587.
- 769 [115] A. Blanazs, A. Ryan, S. Armes, Predictive phase diagrams for RAFT aqueous dispersion polymerization: effect of block copolymer
   770 composition, molecular weight, and copolymer concentration, Macromolecules 45(12) (2012) 5099-5107.
- [116] A. Blanazs, R. Verber, O.O. Mykhaylyk, A.J. Ryan, J.Z. Heath, C.I. Douglas, S.P. Armes, Sterilizable gels from thermoresponsive block
- 772 copolymer worms, J. Am. Chem. Soc. 134(23) (2012) 9741-9748.
- 773 [117] P. Chambon, A. Blanazs, G. Battaglia, S. Armes, Facile synthesis of methacrylic ABC triblock copolymer vesicles by RAFT aqueous
- dispersion polymerization, Macromolecules 45(12) (2012) 5081-5090.

- [118] N.J. Warren, O.O. Mykhaylyk, A.J. Ryan, M. Williams, T. Doussineau, P. Dugourd, R. Antoine, G. Portale, S.P. Armes, Testing the
- Vesicular Morphology to Destruction: Birth and Death of Diblock Copolymer Vesicles Prepared via Polymerization-Induced Self-Assembly, J.
   Am. Chem. Soc. 137(5) (2015) 1929-1937.
- 778 [119] A. Czajka, S.P. Armes, In situ SAXS studies of a prototypical RAFT aqueous dispersion polymerization formulation: monitoring the
- evolution in copolymer morphology during polymerization-induced self-assembly, Chem. Sci. 11(42) (2020) 11443-11454.
- [120] J.R. Lovett, N.J. Warren, L.P. Ratcliffe, M.K. Kocik, S.P. Armes, pH-responsive non-ionic diblock copolymers: Ionization of carboxylic
   acid end-groups induces an order-order morphological transition, Angew. Chem. Int. Ed. 54(4) (2015) 1279-1283.
- [121] K.L. Thompson, C.J. Mable, A. Cockram, N.J. Warren, V.J. Cunningham, E.R. Jones, R. Verber, S.P. Armes, Are block copolymer worms
   more effective Pickering emulsifiers than block copolymer spheres?, Soft Matter 10(43) (2014) 8615-8626.
- [122] N. Penfold, J. Lovett, N. Warren, P. Verstraete, J. Smets, S. Armes, pH-Responsive non-ionic diblock copolymers: protonation of a
   morpholine end-group induces an order–order transition, Polym. Chem. 7(1) (2016) 79-88.
- [123] K.L. Thompson, P. Chambon, R. Verber, S.P. Armes, Can Polymersomes Form Colloidosomes?, J. Am. Chem. Soc. 134(30) (2012)
   12450-12453.
- [124] J. Tan, D. Liu, Y. Bai, C. Huang, X. Li, J. He, Q. Xu, X. Zhang, L. Zhang, An insight into aqueous photoinitiated polymerization-induced
   self-assembly (photo-PISA) for the preparation of diblock copolymer nano-objects, Polym. Chem. 8(8) (2017) 1315-1327.
- [125] J. He, D. Liu, J. Tan, L. Zhang, Sodium Bis(acyl)phosphane oxide (SBAPO): An efficient photoinitiator for blue light initiated aqueous
   RAFT dispersion polymerization, Polymer 145 (2018) 70-79.
- [126] Q. Zhang, R. Zeng, Y. Zhang, Y. Chen, L. Zhang, J. Tan, Two Polymersome Evolution Pathways in One Polymerization-Induced Self Assembly (PISA) System, Macromolecules 53(20) (2020) 8982-8991.
- [127] J. Tan, D. Liu, Y. Bai, C. Huang, X. Li, J. He, Q. Xu, L. Zhang, Enzyme-assisted photoinitiated polymerization-induced self-assembly: An
   oxygen-tolerant method for preparing block copolymer nano-objects in open vessels and multiwell plates, Macromolecules 50(15) (2017)
   5798-5806.
- [128] Q. Zhang, R. Wang, Y. Chen, L. Zhang, J. Tan, Block Copolymer Vesicles with Tunable Membrane Thicknesses and Compositions
   Prepared by Aqueous Seeded Photoinitiated Polymerization-Induced Self-Assembly at Room Temperature, Langmuir 38(8) (2022) 2699 2710.
- 800 [129] Q. Xu, Y. Zhang, X. Li, J. He, J. Tan, L. Zhang, Enzyme catalysis-induced RAFT polymerization in water for the preparation of epoxy-801 functionalized triblock copolymer vesicles, Polym. Chem. 9(39) (2018) 4908-4916.
- [130] D.L. Beattie, O.O. Mykhaylyk, S.P. Armes, Enthalpic incompatibility between two steric stabilizer blocks provides control over the
   wesicle size distribution during polymerization-induced self-assembly in aqueous media, Chem. Sci. 11(39) (2020) 10821-10834.
- 804 [131] N.J. Warren, O.O. Mykhaylyk, D. Mahmood, A.J. Ryan, S.P. Armes, RAFT Aqueous Dispersion Polymerization Yields Poly(ethylene
- glycol)-Based Diblock Copolymer Nano-Objects with Predictable Single Phase Morphologies, J. Am. Chem. Soc. 136(3) (2014) 1023-1033.
   [132] L.D. Blackman, K.E. Doncom, M.I. Gibson, R.K. O'Reilly, Comparison of photo-and thermally initiated polymerization-induced self-
- 807 assembly: a lack of end group fidelity drives the formation of higher order morphologies, Polym. Chem. 8(18) (2017) 2860-2871.
- 808 [133] J.W. Yu, J. Jung, Y.-M. Choi, J.H. Choi, J. Yu, J.K. Lee, N.-H. You, M. Goh, Enhancement of the crosslink density, glass transition
- temperature, and strength of epoxy resin by using functionalized graphene oxide co-curing agents, Polym. Chem. 7(1) (2016) 36-43.
- [134] N.J. Penfold, J.R. Whatley, S.P. Armes, Thermoreversible block copolymer worm gels using binary mixtures of PEG stabilizer blocks,
   Macromolecules 52(4) (2019) 1653-1662.
- [135] J. Tan, H. Sun, M. Yu, B.S. Sumerlin, L. Zhang, Photo-PISA: shedding light on polymerization-induced self-assembly, ACS Macro Lett.
   4(11) (2015) 1249-1253.
- [136] J. Tan, D. Liu, X. Zhang, C. Huang, J. He, Q. Xu, X. Li, L. Zhang, Facile preparation of hybrid vesicles loaded with silica nanoparticles via
   aqueous photoinitiated polymerization-induced self-assembly, RSC Adv. 7(37) (2017) 23114-23121.
- [137] J. Tan, Q. Xu, Y. Zhang, C. Huang, X. Li, J. He, L. Zhang, Room Temperature Synthesis of Self-Assembled AB/B and ABC/BC Blends by
   Photoinitiated Polymerization-Induced Self-Assembly (Photo-PISA) in Water, Macromolecules 51(18) (2018) 7396-7406.
- [138] Y. Zhang, J. He, X. Dai, L. Yu, J. Tan, L. Zhang, Combining the power of heat and light: temperature-programmed photoinitiated RAFT
   dispersion polymerization to tune polymerization-induced self-assembly, Polym. Chem. 10(28) (2019) 3902-3911.
- 820 [139] D. Liu, Y. Chen, L. Zhang, J. Tan, Efficient Preparation of Branched Block Copolymer Assemblies by Photoinitiated RAFT Self-
- 821 Condensing Vinyl Dispersion Polymerization, Macromolecules 53(22) (2020) 9725-9735.
- [140] S. Varlas, L.D. Blackman, H.E. Findlay, E. Reading, P.J. Booth, M.I. Gibson, R.K. O'Reilly, Photoinitiated polymerization-induced self assembly in the presence of surfactants enables membrane protein incorporation into vesicles, Macromolecules 51(16) (2018) 6190-6201.
- [141] M.A. Touve, D.B. Wright, C. Mu, H. Sun, C. Park, N.C. Gianneschi, Block Copolymer Amphiphile Phase Diagrams by High-Throughput
   Transmission Electron Microscopy, Macromolecules 52(15) (2019) 5529-5537.
- [142] N. Zaquen, J. Yeow, T. Junkers, C. Boyer, P.B. Zetterlund, Visible Light-Mediated Polymerization-Induced Self-Assembly Using
   Continuous Flow Reactors, Macromolecules 51(14) (2018) 5165-5172.
- [143] A.N. Albertsen, J.K. Szymański, J. Pérez-Mercader, Emergent properties of giant vesicles formed by a polymerization-induced self assembly (PISA) reaction, Scientific reports 7(1) (2017) 1-8.
- [144] S. Xu, J. Yeow, C. Boyer, Exploiting Wavelength Orthogonality for Successive Photoinduced Polymerization-Induced Self-Assembly and
   Photo-Crosslinking, ACS Macro Lett. 7(11) (2018) 1376-1382.
- [145] K. Ren, J. Perez-Mercader, Thermoresponsive gels directly obtained via visible light-mediated polymerization-induced self-assembly
   with oxygen tolerance, Polym. Chem. 8(23) (2017) 3548-3552.
- [146] Z. Wu, W. Fang, C. Wu, N. Corrigan, T. Zhang, S. Xu, C. Boyer, An aqueous photo-controlled polymerization under NIR wavelengths:
   synthesis of polymeric nanoparticles through thick barriers, Chem. Sci. 13(39) (2022) 11519-11532.
- 836 [147] J. Tan, Q. Xu, X. Li, J. He, Y. Zhang, X. Dai, L. Yu, R. Zeng, L. Zhang, Enzyme-PISA: An Efficient Method for Preparing Well-Defined
- 837 Polymer Nano-Objects under Mild Conditions, Macromol. Rapid Commun. 39(9) (2018) 1700871.

- [148] J. Wan, B. Fan, Y. Liu, T. Hsia, K. Qin, T. Junkers, B.M. Teo, S.H. Thang, Room temperature synthesis of block copolymer nano-objects
   with different morphologies via ultrasound initiated RAFT polymerization-induced self-assembly (sono-RAFT-PISA), Polym. Chem. 11(21)
- 840 (2020) 3564-3572.
- [149] J. Tan, Y. Bai, X. Zhang, L. Zhang, Room temperature synthesis of poly(poly(ethylene glycol) methyl ether methacrylate)-based diblock
   copolymer nano-objects via Photoinitiated Polymerization-Induced Self-Assembly (Photo-PISA), Polym. Chem. 7(13) (2016) 2372-2380.
- 843 [150] J. Tan, Y. Bai, X. Zhang, C. Huang, D. Liu, L. Zhang, Low-Temperature Synthesis of Thermoresponsive Diblock Copolymer Nano-Objects
- via Aqueous Photoinitiated Polymerization-Induced Self-Assembly (Photo-PISA) using Thermoresponsive Macro-RAFT Agents, Macromol.
   Rapid Commun. 37(17) (2016) 1434-1440.
- 846 [151] Y. Du, S. Jia, Y. Chen, L. Zhang, J. Tan, Type I Photoinitiator-Functionalized Block Copolymer Nanoparticles Prepared by RAFT-
- 847 Mediated Polymerization-Induced Self-Assembly, ACS Macro Lett. 10(2) (2021) 297-306.
  - [152] J. Wu, L. Zhang, Y. Chen, J. Tan, Linear and Star Block Copolymer Nanoparticles Prepared by Heterogeneous RAFT Polymerization
     Using an ω,ω-Heterodifunctional Macro-RAFT Agent, ACS Macro Lett. 11(7) (2022) 910-918.
  - [153] D. Liu, S. Yang, S. Peng, Y. Chen, L. Zhang, J. Tan, Simultaneous Synthesis and Self-Assembly of Bottlebrush Block Copolymers at Room
     Temperature via Photoinitiated RAFT Dispersion Polymerization, Macromol. Rapid Commun. 43(8) (2022) 2100921.
- [154] L.P. Ratcliffe, M.J. Derry, A. Ianiro, R. Tuinier, S.P. Armes, A single thermoresponsive diblock copolymer can form spheres, worms or
   vesicles in aqueous solution, Angew. Chem. Int. Ed. 131(52) (2019) 19140-19146.
- 854 [155] K. Ferji, P. Venturini, F. Cleymand, C. Chassenieux, J.-L. Six, In situ glyco-nanostructure formulation via photo-polymerization induced 855 self-assembly, Polym. Chem. 9(21) (2018) 2868-2872.
- [156] V.L. Romero Castro, B. Nomeir, A.A. Arteni, M. Ouldali, J.-L. Six, K. Ferji, Dextran-Coated Latex Nanoparticles via Photo-RAFT Mediated
   Polymerization Induced Self-Assembly, Polymers 13(23) (2021) 4064.
- [157] D. Ikkene, A.A. Arteni, M. Ouldali, J.-L. Six, K. Ferji, Self-assembly of amphiphilic copolymers containing polysaccharide: PISA versus
   nanoprecipitation, and the temperature effect, Polym. Chem. 11(29) (2020) 4729-4740.
- [158] D. Ikkene, A.A. Arteni, M. Ouldali, G. Francius, A. Brûlet, J.-L. Six, K. Ferji, Direct Access to Polysaccharide-Based Vesicles with a
   Tunable Membrane Thickness in a Large Concentration Window via Polymerization-Induced Self-Assembly, Biomacromolecules 22(7)
   (2021) 3128-3137.
- 863 [159] D. Ikkene, A.A. Arteni, C. Boulogne, J.-L. Six, K. Ferji, Multicompartment Vesicles: A Key Intermediate Structure in Polymerization 864 Induced Self-Assembly of Graft Copolymers, Macromolecules 55(11) (2022) 4268-4275.
- 865 [160] S. Varlas, P.G. Georgiou, P. Bilalis, J.R. Jones, N. Hadjichristidis, R.K. O'Reilly, Poly(sarcosine)-Based Nano-Objects with Multi-Protease
- Resistance by Aqueous Photoinitiated Polymerization-Induced Self-Assembly (Photo-PISA), Biomacromolecules 19(11) (2018) 4453-4462.
  [161] C. Ma, X. Liu, G. Wu, P. Zhou, Y. Zhou, L. Wang, X. Huang, Efficient way to generate protein-based nanoparticles by in-situ
- 868 photoinitiated polymerization-induced self-assembly, ACS Macro Lett. 6(7) (2017) 689-694.
- [162] R. Gibson, S. Armes, O. Musa, A. Fernyhough, End-group ionisation enables the use of poly (N-(2-methacryloyloxy) ethyl pyrrolidone)
   as an electrosteric stabiliser block for polymerisation-induced self-assembly in aqueous media, Polym. Chem. 10(11) (2019) 1312-1323.
- [163] L. Yang, M. Liang, C. Cui, X. Li, L. Li, X. Pan, H.S. Yazd, M. Hong, J. Lu, Y.C. Cao, Enhancing the Nucleolytic Resistance and Bioactivity of
   Functional Nucleic Acids by Diverse Nanostructures through in Situ Polymerization-Induced Self-assembly, ChemBioChem 22(4) (2021) 754 759.
- [164] S. Sugihara, A. Blanazs, S.P. Armes, A.J. Ryan, A.L. Lewis, Aqueous dispersion polymerization: a new paradigm for in situ block
   copolymer self-assembly in concentrated solution, J. Am. Chem. Soc. 133(39) (2011) 15707-15713.
- [165] S. Sugihara, S.P. Armes, A. Blanazs, A.L. Lewis, Non-spherical morphologies from cross-linked biomimetic diblock copolymers using
   RAFT aqueous dispersion polymerization, Soft Matter 7(22) (2011) 10787-10793.
- [166] V. Ladmiral, A. Charlot, M. Semsarilar, S.P. Armes, Synthesis and characterization of poly (amino acid methacrylate)-stabilized diblock
   copolymer nano-objects, Polym. Chem. 6(10) (2015) 1805-1816.
- [167] K.E. Doncom, N.J. Warren, S.P. Armes, Polysulfobetaine-based diblock copolymer nano-objects via polymerization-induced self assembly, Polym. Chem. 6(41) (2015) 7264-7273.
- [168] M. Semsarilar, V. Ladmiral, A. Blanazs, S. Armes, Cationic polyelectrolyte-stabilized nanoparticles via RAFT aqueous dispersion
   polymerization, Langmuir 29(24) (2013) 7416-7424.
- [169] M.A. Rahman, Y. Cha, L. Yuan, P. Pageni, T. Zhu, M.S. Jui, C. Tang, Polymerization-induced self-assembly of metallo-polyelectrolyte
   block copolymers, J. Polym. Sci. 58(1) (2020) 77-83.
- [170] L. Luppi, T. Babut, E. Petit, M. Rolland, D. Quemener, L. Soussan, M. Moradi, M. Semsarilar, Antimicrobial polylysine decorated nano structures prepared through polymerization induced self-assembly (PISA), Polym. Chem. 10(3) (2019) 336-344.
- [171] M. Semsarilar, V. Ladmiral, A. Blanazs, S. Armes, Anionic polyelectrolyte-stabilized nanoparticles via RAFT aqueous dispersion
   polymerization, Langmuir 28(1) (2012) 914-922.
- 890 [172] S.M. North, S.P. Armes, Aqueous solution behavior of stimulus-responsive poly (methacrylic acid)-poly (2-hydroxypropyl
- 891 methacrylate) diblock copolymer nanoparticles, Polym. Chem. 11(12) (2020) 2147-2156.
- 892 [173] L. Romero-Azogil, N.J. Penfold, S.P. Armes, Tuning the hydroxyl functionality of block copolymer worm gels modulates their
- thermoresponsive behavior, Polym. Chem. 11(31) (2020) 5040-5050.
- [174] L.A. Fielding, C.T. Hendley IV, E. Asenath-Smith, L.A. Estroff, S.P. Armes, Rationally designed anionic diblock copolymer worm gels are
   useful model systems for calcite occlusion studies, Polym. Chem. 10(37) (2019) 5131-5141.
- 896 [175] M. Williams, N. Penfold, S. Armes, Cationic and reactive primary amine-stabilised nanoparticles via RAFT aqueous dispersion
- 897 polymerisation, Polym. Chem. 7(2) (2016) 384-393.
- 898 [176] M. Williams, N. Penfold, J. Lovett, N. Warren, C. Douglas, N. Doroshenko, P. Verstraete, J. Smets, S. Armes, Bespoke cationic nano-
- 899 objects via RAFT aqueous dispersion polymerisation, Polym. Chem. 7(23) (2016) 3864-3873.

- 900 [177] V. Ladmiral, M. Semsarilar, I. Canton, S.P. Armes, Polymerization-Induced Self-Assembly of Galactose-Functionalized Biocompatible 901 Diblock Copolymers for Intracellular Delivery, J. Am. Chem. Soc. 135(36) (2013) 13574-13581.
- [178] T.T. Dao, L. Vezenkov, G. Subra, M. Amblard, M. In, J.-F. Le Meins, F. Aubrit, M.-A. Moradi, V. Ladmiral, M. Semsarilar, Self-Assembling
   Peptide—Polymer Nano-Objects via Polymerization-Induced Self-Assembly, Macromolecules 53(16) (2020) 7034-7043.
- 904 [179] L. Ratcliffe, K. Bentley, R. Wehr, N. Warren, B. Saunders, S. Armes, Cationic disulfide-functionalized worm gels, Polym. Chem. 8(38) 905 (2017) 5962-5971.
- 906 [180] J. Rosselgong, A. Blanazs, P. Chambon, M. Williams, M. Semsarilar, J. Madsen, G. Battaglia, S. Armes, Thiol-functionalized block 907 copolymer vesicles, ACS Macro Letters 1(8) (2012) 1041-1045.
- 908 [181] B. Fan, Y. Liu, J. Wan, S. Crawford, S.H. Thang, Polymerization-Induced Self-Assembly (PISA) and "Host–Guest" Complexation-Directed
   909 Polymer/Gold Nanocomposites, ACS Mater. Lett. 2(5) (2020) 492-498.
- 910 [182] H. Phan, M. Cossutta, C. Houppe, C. Le Coeur, S. Prevost, I. Cascone, J. Courty, J. Penelle, B. Couturaud, Polymerization-Induced Self-
- 911 Assembly (PISA) for in situ drug encapsulation or drug conjugation in cancer application, J. Colloid Interface Sci. 618 (2022) 173-184.
- [183] S.J. Byard, M. Williams, B.E. McKenzie, A. Blanazs, S.P. Armes, Preparation and cross-linking of all-acrylamide diblock copolymer nano objects via polymerization-induced self-assembly in aqueous solution, Macromolecules 50(4) (2017) 1482-1493.
- 914 [184] G. Mellot, P. Beaunier, J.M. Guigner, L. Bouteiller, J. Rieger, F. Stoffelbach, Beyond simple AB diblock copolymers: application of
   915 bifunctional and trifunctional RAFT agents to PISA in water, Macromol. Rapid Commun. 40(2) (2019) 1800315.
- 916 [185] S. Parkinson, N.S. Hondow, J.S. Conteh, R.A. Bourne, N.J. Warren, All-aqueous continuous-flow RAFT dispersion polymerisation for 917 efficient preparation of diblock copolymer spheres, worms and vesicles, React. Chem. Eng. 4(5) (2019) 852-861.
- 918 [186] Q. Qu, G. Liu, X. Lv, B. Zhang, Z. An, In situ cross-linking of vesicles in polymerization-induced self-assembly, ACS Macro Lett. 5(3) 919 (2016) 316-320.
- [187] L. Zhang, Q. Lu, X. Lv, L. Shen, B. Zhang, Z. An, In situ cross-linking as a platform for the synthesis of triblock copolymer vesicles with
   diverse surface chemistry and enhanced stability via RAFT dispersion polymerization, Macromolecules 50(5) (2017) 2165-2174.
- 922 [188] W. Zhou, Q. Qu, Y. Xu, Z. An, Aqueous polymerization-induced self-assembly for the synthesis of ketone-functionalized nano-objects
   923 with low polydispersity, ACS Macro Lett. 4(5) (2015) 495-499.
- [189] X. Wang, J. Zhou, X. Lv, B. Zhang, Z. An, Temperature-Induced Morphological Transitions of Poly(dimethylacrylamide)–Poly(diacetone
   acrylamide) Block Copolymer Lamellae Synthesized via Aqueous Polymerization-Induced Self-Assembly, Macromolecules 50(18) (2017)
   7222-7232.
- 927 [190] P. Biais, P. Beaunier, F. Stoffelbach, J. Rieger, Loop-stabilized BAB triblock copolymer morphologies by PISA in water, Polym. Chem.
  928 9(35) (2018) 4483-4491.
- 929 [191] P. Biais, O. Colombani, L. Bouteiller, F. Stoffelbach, J. Rieger, Unravelling the formation of BAB block copolymer assemblies during
   930 PISA in water, Polym. Chem. 11(28) (2020) 4568-4578.
- [192] X. Luo, S. Zhao, Y. Chen, L. Zhang, J. Tan, Switching between thermal initiation and photoinitiation redirects RAFT-mediated
   polymerization-induced self-assembly, Macromolecules 54(6) (2021) 2948-2959.
- [193] C.A. Figg, R.N. Carmean, K.C. Bentz, S. Mukherjee, D.A. Savin, B.S. Sumerlin, Tuning Hydrophobicity To Program Block Copolymer
   Assemblies from the Inside Out, Macromolecules 50(3) (2017) 935-943.
- [194] J. Yeow, R. Chapman, J. Xu, C. Boyer, Oxygen tolerant photopolymerization for ultralow volumes, Polym. Chem. 8(34) (2017) 50125022.
- 937 [195] N. Zaquen, H. Zu, A.M.N.B.P.H.A. Kadir, T. Junkers, P.B. Zetterlund, C. Boyer, Scalable Aqueous Reversible Addition–Fragmentation
   938 Chain Transfer Photopolymerization-Induced Self-Assembly of Acrylamides for Direct Synthesis of Polymer Nanoparticles for Potential Drug
- 939 Delivery Applications, ACS Appl. Polym. Mater. 1(6) (2019) 1251-1256.
- [196] X. Wang, C.A. Figg, X. Lv, Y. Yang, B.S. Sumerlin, Z. An, Star architecture promoting morphological transitions during polymerization induced self-assembly, ACS Macro Lett. 6(4) (2017) 337-342.
- [197] Y. Jiang, N. Xu, J. Han, Q. Yu, L. Guo, P. Gao, X. Lu, Y. Cai, The direct synthesis of interface-decorated reactive block copolymer
   nanoparticles via polymerisation-induced self-assembly, Polym. Chem. 6(27) (2015) 4955-4965.
- 944 [198] Y. Ma, P. Gao, Y. Ding, L. Huang, L. Wang, X. Lu, Y. Cai, Visible light initiated thermoresponsive aqueous dispersion polymerization-945 induced self-assembly, Macromolecules 52(3) (2019) 1033-1041.
- [199] P. Gao, H. Cao, Y. Ding, M. Cai, Z. Cui, X. Lu, Y. Cai, Synthesis of Hydrogen-Bonded Pore-Switchable Cylindrical Vesicles via Visible Light-Mediated RAFT Room-Temperature Aqueous Dispersion Polymerization, ACS Macro Lett. 5(12) (2016) 1327-1331.
- [200] V. Baddam, L. Valinen, H. Tenhu, Thermoresponsive Polycation-Stabilized Nanoparticles through PISA. Control of Particle Morphology
   with a Salt, Macromolecules 54(9) (2021) 4288-4299.
- [201] P.G. Georgiou, H.L. Marton, A.N. Baker, T.R. Congdon, T.F. Whale, M.I. Gibson, Polymer Self-Assembly Induced Enhancement of Ice
   Recrystallization Inhibition, J. Am. Chem. Soc. 143(19) (2021) 7449-7461.
- [202] L. Cao, Q. Zhao, Q. Liu, L. Ma, C. Li, X. Wang, Y. Cai, Electrostatic manipulation of triblock terpolymer nanofilm compartmentalization
   during aqueous photoinitiated polymerization-induced self-assembly, Macromolecules 53(6) (2020) 2220-2227.
- [203] M. Morimura, S. Ida, M. Oyama, H. Takeshita, S. Kanaoka, Design of Hydrogels with Thermoresponsive Crosslinked Domain Structures
   via the Polymerization-Induced Self-Assembly Process and Their Thermoresponsive Toughening in Air, Macromolecules 54(4) (2021) 1732 1741.
- 957 [204] J. He, D. Lin, Y. Chen, L. Zhang, J. Tan, One-Step Preparation of Thermo-Responsive Poly (N-isopropylacrylamide)-Based Block
- 958 Copolymer Nanoparticles by Aqueous Photoinitiated Polymerization-Induced Self-Assembly, Macromol. Rapid Commun. (2021) 2100201.
   959 [205] S. Piogé, T.N. Tran, T.G. McKenzie, S. Pascual, M. Ashokkumar, L. Fontaine, G. Qiao, Sono-RAFT Polymerization-Induced Self-Assembly
   960 in Aqueous Dispersion: Synthesis of LCST-type Thermosensitive Nanogels, Macromolecules 51(21) (2018) 8862-8869.
- 961 [206] C.A. Figg, A. Simula, K.A. Gebre, B.S. Tucker, D.M. Haddleton, B.S. Sumerlin, Polymerization-induced thermal self-assembly (PITSA),
- 962 Chem. Sci. 6(2) (2015) 1230-1236.

- 963 [207] Y. Li, Z. Ye, L. Shen, Y. Xu, A. Zhu, P. Wu, Z. An, Formation of Multidomain Hydrogels via Thermally Induced Assembly of PISA-964 Generated Triblock Terpolymer Nanogels, Macromolecules 49(8) (2016) 3038-3048.
- 965 [208] G. Mellot, J.m. Guigner, L. Bouteiller, F. Stoffelbach, J. Rieger, Templated PISA: Driving Polymerization-Induced Self-Assembly towards
   966 Fibre Morphology, Angew. Chem. Int. Ed. 131(10) (2019) 3205-3209.
- 967 [209] B. Zhang, X. Wang, A. Zhu, K. Ma, Y. Lv, X. Wang, Z. An, Enzyme-initiated reversible addition–fragmentation chain transfer 968 polymerization, Macromolecules 48(21) (2015) 7792-7802.
- 969 [210] G. Liu, Q. Qiu, W. Shen, Z. An, Aqueous dispersion polymerization of 2-methoxyethyl acrylate for the synthesis of biocompatible 970 nanoparticles using a hydrophilic RAFT polymer and a redox initiator, Macromolecules 44(13) (2011) 5237-5245.
- 971 [211] S. Sugihara, A.H. Ma'Radzi, S. Ida, S. Irie, T. Kikukawa, Y. Maeda, In situ nano-objects via RAFT aqueous dispersion polymerization of 2 972 methoxyethyl acrylate using poly(ethylene oxide) macromolecular chain transfer agent as steric stabilizer, Polymer 76 (2015) 17-24.
- 973 [212] J.M. Cumming, O.J. Deane, S.P. Armes, Reversible Addition-Fragmentation Chain Transfer Aqueous Dispersion Polymerization of 4-
- 974 Hydroxybutyl Acrylate Produces Highly Thermoresponsive Diblock Copolymer Nano-Objects, Macromolecules 55(3) (2022) 788–798.
- 975 [213] O.J. Deane, J. Jennings, S.P. Armes, Shape-shifting thermoreversible diblock copolymer nano-objects via RAFT aqueous dispersion
   976 polymerization of 4-hydroxybutyl acrylate, Chem. Sci. 12(41) (2021) 13719-13729.
- 977 [214] O.J. Deane, J. Jennings, T.J. Neal, O.M. Musa, A. Fernyhough, S.P. Armes, Synthesis and Aqueous Solution Properties of Shape-Shifting
   978 Stimulus-Responsive Diblock Copolymer Nano-Objects, Chem. Mater. 33(19) (2021) 7767-7779.
- 979 [215] D.L. Beattie, O.J. Deane, O.O. Mykhaylyk, S.P. Armes, RAFT aqueous dispersion polymerization of 4-hydroxybutyl acrylate: effect of
   980 end-group ionization on the formation and colloidal stability of sterically-stabilized diblock copolymer nanoparticles, Polym. Chem. 13
   981 (2022) 655-667.
- [216] L. Zhang, L. Xie, S. Xu, R.P. Kuchel, Y. Dai, K. Jung, C. Boyer, Dual Role of Doxorubicin for Photopolymerization and Therapy,
   Biomacromolecules 21(9) (2020) 3887-3897.
- 984 [217] C. Grazon, J. Rieger, N. Sanson, B. Charleux, Study of poly(N, N-diethylacrylamide) nanogel formation by aqueous dispersion
- polymerization of N, N-diethylacrylamide in the presence of poly(ethylene oxide)-b-poly(N, N-dimethylacrylamide) amphiphilic
   macromolecular RAFT agents, Soft Matter 7(7) (2011) 3482-3490.
- 987 [218] J. Rieger, C. Grazon, B. Charleux, D. Alaimo, C. Jerome, Pegylated Thermally Responsive Block Copolymer Micelles and Nanogels via In
   988 Situ RAFT Aqueous Dispersion Polymerization, J. Polym. Sci. A: Polym. Chem. 47(9) (2009) 2373-2390.
- 989 [219] L. Yan, W. Tao, One-step synthesis of pegylated cationic nanogels of poly (N, N'-dimethylaminoethyl methacrylate) in aqueous
   990 solution via self-stabilizing micelles using an amphiphilic macroRAFT agent, Polymer 51(10) (2010) 2161-2167.
- [220] B. Zhang, X. Lv, Z. An, Modular monomers with tunable solubility: synthesis of highly incompatible block copolymer nano-objects via
   RAFT aqueous dispersion polymerization, ACS Macro Lett. 6(3) (2017) 224-228.
- 993 [221] F.H. Sobotta, M. Kuchenbrod, S. Hoeppener, J.C. Brendel, One polymer composition, various morphologies: the decisive influence of 994 conditions on the polymerization-induced self-assembly (PISA) of N-acryloyl thiomorpholine, Nanoscale 12(39) (2020) 20171-20176.
- 995 [222] R. Takahashi, S. Miwa, F.H. Sobotta, J.H. Lee, S. Fujii, N. Ohta, J.C. Brendel, K. Sakurai, Unraveling the kinetics of the structural
- development during polymerization-induced self-assembly: decoupling the polymerization and the micelle structure, Polym. Chem. 11(8)
   (2020) 1514-1524.
- 998 [223] W. Shen, Y. Chang, G. Liu, H. Wang, A. Cao, Z. An, Biocompatible, antifouling, and thermosensitive core- shell nanogels synthesized
   999 by RAFT aqueous dispersion polymerization, Macromolecules 44(8) (2011) 2524-2530.
- 1000 [224] T.T. Dao, L. Vezenkov, G. Subra, V. Ladmiral, M. Semsarilar, Nano-assemblies with core-forming hydrophobic polypeptide via
- 1001 polymerization-induced self-assembly (PISA), Polym. Chem. 12(1) (2021) 113-121.
- 1002 [225] N. Audureau, F. Coumes, C. Veith, C. Guibert, J.-M. Guigner, F. Stoffelbach, J. Rieger, Synthesis and Characterization of Temperature-1003 Responsive N-Cyanomethylacrylamide-Containing Diblock Copolymer Assemblies in Water, Polymers 13(24) (2021) 4424.
- 1004 [226] N. Audureau, F. Coumes, J.-M. Guigner, C. Guibert, F. Stoffelbach, J. Rieger, Dual Thermo- and pH-Responsive N-
- 1005 Cyanomethylacrylamide-Based Nano-Objects Prepared by RAFT-Mediated Aqueous Polymerization-Induced Self-Assembly,
- 1006 Macromolecules (2022).
- [227] L. Huang, Y. Ding, Y. Ma, L. Wang, Q. Liu, X. Lu, Y. Cai, Colloidal Stable PIC Vesicles and Lamellae Enabled by Wavelength-Orthogonal
   Disulfide Exchange and Polymerization-Induced Electrostatic Self-Assembly, Macromolecules 52(12) (2019) 4703-4712.
- 1009 [228] T.N. Tran, S. Piogé, L. Fontaine, S. Pascual, Hydrogen-Bonding UCST-Thermosensitive Nanogels by Direct Photo-RAFT
- 1010 Polymerization-Induced Self-Assembly in Aqueous Dispersion, Macromol. Rapid Commun. 41(13) (2020) 2000203.
- 1011 [229] J. Depoorter, X. Yan, B. Zhang, G. Sudre, A. Charlot, E. Fleury, J. Bernard, All poly (ionic liquid) block copolymer nanoparticles from
- antagonistic isomeric macromolecular blocks via aqueous RAFT polymerization-induced self-assembly, Polym. Chem. 12(1) (2021) 82-91.
   [230] J. Lovett, L. Ratcliffe, N. Warren, S. Armes, M. Smallridge, R. Cracknell, B. Saunders, A robust cross-linking strategy for block
- 1014 copolymer worms prepared via polymerization-induced self-assembly, Macromolecules 49(8) (2016) 2928-2941.
- 1015 [231] N.J. Penfold, Y. Ning, P. Verstraete, J. Smets, S.P. Armes, Cross-linked cationic diblock copolymer worms are superflocculants for 1016 micrometer-sized silica particles, Chem. Sci. 7(12) (2016) 6894-6904.
- 1017 [232] J. Tan, X. Zhang, D. Liu, Y. Bai, C. Huang, X. Li, L. Zhang, Facile Preparation of CO2-Responsive Polymer Nano-Objects via Aqueous
- 1018 Photoinitiated Polymerization-Induced Self-Assembly (Photo-PISA), Macromol. Rapid Commun. 38(13) (2017) 1600508.
- [233] S. Guragain, J. Perez-Mercader, Light-mediated one-pot synthesis of an ABC triblock copolymer in aqueous solution via RAFT and the
   effect of pH on copolymer self-assembly, Polym. Chem. 9(29) (2018) 4000-4006.
- 1021 [234] L. Yu, Y. Zhang, X. Dai, Q. Xu, L. Zhang, J. Tan, Open-air preparation of cross-linked CO 2-responsive polymer vesicles by enzyme-
- assisted photoinitiated polymerization-induced self-assembly, Chem. Commun. 55(79) (2019) 11920-11923.
- [235] L.P. Ratcliffe, C. Couchon, S.P. Armes, J.M. Paulusse, Inducing an order–order morphological transition via chemical degradation of
   amphiphilic diblock copolymer nano-objects, Biomacromolecules 17(6) (2016) 2277-2283.

- 1025 [236] J. He, J. Cao, Y. Chen, L. Zhang, J. Tan, Thermoresponsive block copolymer vesicles by visible light-initiated seeded polymerization-1026 induced self-assembly for temperature-regulated enzymatic nanoreactors, ACS Macro Lett. 9(4) (2020) 533-539. 1027 [237] A.B. Korpusik, Y. Tan, J.B. Garrison, W. Tan, B.S. Sumerlin, Aptamer-Conjugated Micelles for Targeted Photodynamic Therapy Via 1028 Photoinitiated Polymerization-Induced Self-Assembly, Macromolecules 54 (2021) 7354–7363. 1029 [238] G. Liu, Q. Qiu, Z. An, Development of thermosensitive copolymers of poly (2-methoxyethyl acrylate-co-poly (ethylene glycol) methyl 1030 ether acrylate) and their nanogels synthesized by RAFT dispersion polymerization in water, Polym. Chem. 3(2) (2012) 504-513. 1031 [239] N. Audureau, F. Coumes, J.-M. Guigner, T.P.T. Nguyen, C. Ménager, F. Stoffelbach, J. Rieger, Thermoresponsive properties of poly 1032 (acrylamide-co-acrylonitrile)-based diblock copolymers synthesized (by PISA) in water, Polym. Chem. 11(37) (2020) 5998-6008. 1033 [240] S. Xu, T. Zhang, R.P. Kuchel, J. Yeow, C. Boyer, Gradient Polymerization–Induced Self-Assembly: A One-Step Approach, Macromol. 1034 Rapid Commun. 41(1) (2020) 1900493. 1035 [241] S.J. Byard, C.T. O'Brien, M.J. Derry, M. Williams, O.O. Mykhaylyk, A. Blanazs, S.P. Armes, Unique aqueous self-assembly behavior of a 1036 thermoresponsive diblock copolymer, Chem. Sci. 11(2) (2020) 396-402. 1037 [242] G. Moad, Y.K. Chong, A. Postma, E. Rizzardo, S.H. Thang, Advances in RAFT polymerization: the synthesis of polymers with defined 1038 end-groups, Polymer 46(19) (2005) 8458-8468. 1039 [243] Y. Zhang, L. Yu, X. Dai, L. Zhang, J. Tan, Structural Difference in Macro-RAFT Agents Redirects Polymerization-Induced Self-Assembly, 1040 ACS Macro Lett. 8(9) (2019) 1102-1109. 1041 [244] S. Han, J. Wu, Y. Zhang, J. Lai, Y. Chen, L. Zhang, J. Tan, Utilization of Poor RAFT Control in Heterogeneous RAFT Polymerization, 1042 Macromolecules 54(10) (2021) 4669-4681. 1043 [245] P. Lertturongchai, M.I.A. Ibrahim, A. Durand, P. Sunintaboon, K. Ferji, Synthesis of Thermoresponsive Copolymers with Tunable UCST-1044 Type Phase Transition Using Aqueous Photo-RAFT Polymerization, Macromol. Rapid Commun, 41(9) (2020) 2000058. 1045 [246] S.C. Ligon, B. Husar, H. Wutzel, R. Holman, R. Liska, Strategies to Reduce Oxygen Inhibition in Photoinduced Polymerization, Chem. 1046 Rev. 114(1) (2014) 557-589. 1047 [247] D. Liu, W. Cai, L. Zhang, C. Boyer, J. Tan, Efficient Photoinitiated Polymerization-Induced Self-Assembly with Oxygen Tolerance 1048 through Dual-Wavelength Type I Photoinitiation and Photoinduced Deoxygenation, Macromolecules 53(4) (2020) 1212-1223. 1049 [248] S. Shanmugam, J. Xu, C. Boyer, Aqueous RAFT Photopolymerization with Oxygen Tolerance, Macromolecules 49(24) (2016) 9345-1050 9357. 1051 [249] T.G. McKenzie, Q. Fu, M. Uchiyama, K. Satoh, J. Xu, C. Boyer, M. Kamigaito, G.G. Qiao, Beyond Traditional RAFT: Alternative Activation 1052 of Thiocarbonylthio Compounds for Controlled Polymerization, Adv. Sci. 3(9) (2016) 1500394. 1053 [250] T.G. McKenzie, L.P.D. Costa, Q. Fu, D.E. Dunstan, G.G. Qiao, Investigation into the photolytic stability of RAFT agents and the 1054 implications for photopolymerization reactions, Polym. Chem. 7(25) (2016) 4246-4253. 1055 [251] S. Shanmugam, J. Xu, C. Boyer, Photoinduced Electron Transfer–Reversible Addition–Fragmentation Chain Transfer (PET-RAFT) 1056 Polymerization of Vinyl Acetate and N-Vinylpyrrolidinone: Kinetic and Oxygen Tolerance Study, Macromolecules 47(15) (2014) 4930-4942. 1057 [252] J. Yeow, J.T. Xu, C. Boyer, Polymerization-Induced Self-Assembly Using Visible Light Mediated Photoinduced Electron Transfer-1058 Reversible Addition-Fragmentation Chain Transfer Polymerization, ACS Macro Lett. 4(9) (2015) 984-990. 1059 [253] J. Yeow, S. Shanmugam, N. Corrigan, R.P. Kuchel, J.T. Xu, C. Boyer, A Polymerization-Induced Self-Assembly Approach to 1060 Nanoparticles Loaded with Singlet Oxygen Generators, Macromolecules 49(19) (2016) 7277-7285. 1061 [254] J. Xu, K. Jung, N.A. Corrigan, C. Boyer, Aqueous photoinduced living/controlled polymerization: tailoring for bioconjugation, Chem. 1062 Sci. 5(9) (2014) 3568-3575. 1063 [255] J.T. Xu, K. Jung, A. Atme, S. Shanmugam, C. Boyer, A Robust and Versatile Photoinduced Living Polymerization of Conjugated and 1064 Unconjugated Monomers and Its Oxygen Tolerance, J. Am. Chem. Soc. 136(14) (2014) 5508-5519. 1065 [256] B. Nomeir, O. Fabre, K. Ferji, Effect of Tertiary Amines on the Photoinduced Electron Transfer-Reversible Addition-Fragmentation 1066 Chain Transfer (PET-RAFT) Polymerization, Macromolecules 52(18) (2019) 6898-6903. 1067 [257] M. Vamvakaki, G.F. Unali, V. Bütün, S. Boucher, K.L. Robinson, N.C. Billingham, S.P. Armes, Effect of Partial Quaternization on the 1068 Aqueous Solution Properties of Tertiary Amine-Based Polymeric Surfactants: Unexpected Separation of Surface Activity and Cloud Point 1069 Behavior, Macromolecules 34(20) (2001) 6839-6841. 1070 [258] V. Bütün, S.P. Armes, N.C. Billingham, Synthesis and aqueous solution properties of near-monodisperse tertiary amine methacrylate 1071 homopolymers and diblock copolymers, Polymer 42(14) (2001) 5993-6008. 1072 [259] M. Save, J.V.M. Weaver, S.P. Armes, P. McKenna, Atom Transfer Radical Polymerization of Hydroxy-Functional Methacrylates at 1073 Ambient Temperature: Comparison of Glycerol Monomethacrylate with 2-Hydroxypropyl Methacrylate, Macromolecules 35(4) (2002) 1074 1152-1159. 1075 [260] J. Tan, Q. Xu, X. Li, J. He, Y. Zhang, X. Dai, L. Yu, R. Zeng, L. Zhang, Enzyme-PISA: An Efficient Method for Preparing Well-Defined 1076 Polymer Nano-Objects under Mild Conditions, Macromol Rapid Commun 39(9) (2018) e1700871. 1077 [261] A. Halperin, M. Kroger, F.M. Winnik, Poly(N-isopropylacrylamide) Phase Diagrams: Fifty Years of Research, Angew. Chem.-Int. Edit. 1078 54(51) (2015) 15342-15367. 1079 [262] K.H. Kim, J. Kim, W.H. Jo, Preparation of hydrogel nanoparticles by atom transfer radical polymerization of N-isopropylacrylamide in 1080 aqueous media using PEG macro-initiator, Polymer 46(9) (2005) 2836-2840. 1081 [263] J. He, D.N. Lin, Y. Chen, L. Zhang, J.B. Tan, One-Step Preparation of Thermo-Responsive Poly(N-isopropylacrylamide)-Based Block 1082 Copolymer Nanoparticles by Aqueous Photoinitiated Polymerization-Induced Self-Assembly, Macromol. Rapid Commun. 42(18) (2021) 7. 1083 [264] I. Idziak, D. Avoce, D. Lessard, D. Gravel, X.X. Zhu, Thermosensitivity of Aqueous Solutions of Poly(N,N-diethylacrylamide), 1084 Macromolecules 32(4) (1999) 1260-1263. 1085 [265] C.M. Schilli, A.H.E. Müller, E. Rizzardo, S.H. Thang, Y.K. Chong, RAFT Polymers: Novel Precursors for Polymer—Protein Conjugates,
- 1086 Advances in Controlled/Living Radical Polymerization, American Chemical Society2003, pp. 603-618.

- 1087 [266] T.N. Tran, S. Piogé, L. Fontaine, S. Pascual, Hydrogen-Bonding UCST-Thermosensitive Nanogels by Direct Photo-RAFT Polymerization-1088 Induced Self-Assembly in Aqueous Dispersion, Macromol Rapid Commun 41(13) (2020) e2000203.
- 1089 [267] N. Audureau, C. Veith, F. Coumes, T.P.T. Nguyen, J. Rieger, F. Stoffelbach, RAFT-Polymerized N-Cyanomethylacrylamide-Based 1090 (Co)polymers Exhibiting Tunable UCST Behavior in Water, Macromol Rapid Commun 42(23) (2021) e2100556.
- 1091 [268] N. Audureau, F. Coumes, C. Veith, C. Guibert, J.M. Guigner, F. Stoffelbach, J. Rieger, Synthesis and Characterization of Temperature-1092 Responsive N-Cyanomethylacrylamide-Containing Diblock Copolymer Assemblies in Water, Polymers 13 (2021) 4424.
- 1092 (269) P. Hennaux, A. Laschewsky, Novel nonionic surfactants based on sulfoxides. 2. Homo- and copolymers, Colloid Polym. Sci. 281(9)
- 1094 (2003) 807-814.
- 1095 [270] Q. Yu, Y. Ding, H. Cao, X. Lu, Y. Cai, Use of Polyion Complexation for Polymerization-Induced Self-Assembly in Water under Visible
- 1096 Light Irradiation at 25 °C, ACS Macro Lett. 4(11) (2015) 1293-1296.
- 1097